

DECLARATION

I, Akiko KOSEMURA, of HIRAKI & ASSOCIATES, do solemnly and sincerely declare as follows:

1. That I am well acquainted with the English and Japanese languages and am competent to translate from Japanese into English.
2. That I have executed, with the best of my ability, a true and correct translation into English of Japanese Patent Application No. 329115/2003 filed on September 19, 2003, a copy of which I attach herewith.

This 16th day of July, 2010


Akiko KOSEMURA

[Title of Document] CLAIMS

[Claim 1]

A replicon RNA, comprising a nucleotide sequence containing at least the 5' untranslated region, the nucleotide sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein and the 3' untranslated region on the genomic RNA of hepatitis C virus of genotype 2a.

[Claim 2]

The replicon RNA of claim 1, containing at least one selection marker gene or a reporter gene, and at least one IRES sequence.

[Claim 3]

A replicon RNA, comprising a nucleotide sequence containing the 5' untranslated region comprising the nucleotide sequence represented by SEQ ID NO: 9 or 10; at least one selection marker gene or a reporter gene; an IRES sequence; the nucleotide sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein on the genomic RNA of hepatitis C virus of genotype 2a; and the 3' untranslated region comprising the nucleotide sequence represented by SEQ ID NO: 11 or 12.

[Claim 4]

The replicon RNA of any one of claims 1 to 3, wherein the genomic RNA of hepatitis C virus of genotype 2a is an RNA comprising the nucleotide sequence represented by SEQ ID NO: 3 or 5.

[Claim 5]

A replicon RNA, comprising the following RNA (a) or (b):

(a) an RNA comprising the nucleotide sequence represented by SEQ ID NO: 1 or 2; and

(b) an RNA comprising a nucleotide sequence derived from the nucleotide sequence represented by SEQ ID NO: 1 or 2 by deletion, substitution or addition of 1 to 10 nucleotides, and being capable of autonomous replication.

[Claim 6]

A replicon-replicating cell, which is prepared by introducing the replicon RNA of any one of claims 1 to 5 into a cell.

[Claim 7]

The replicon-replicating cell of claim 6, wherein the cell is a eukaryotic cell.

[Claim 8]

The replicon-replicating cell of claim 7, wherein the eukaryotic cell is a human liver-derived cell, a human uterine cervix-derived cell or a human fetal kidney-derived cell.

[Claim 9]

The replicon-replicating cell of claim 7, wherein the eukaryotic cell is any one cell selected from the group consisting of an Huh7 cell, an HepG2 cell, an IMY-N9 cell, an HeLa cell and a 293 cell.

[Claim 10]

The replicon RNA of any one of claims 1 to 5, which is for producing or evaluating a therapeutic agent or a diagnostic agent against hepatitis C virus infection.

[Claim 11]

The replicon-replicating cell of any one of claims 6 to 9, which is for producing or evaluating a therapeutic agent or a diagnostic agent against hepatitis C virus infection.

[Claim 12]

The replicon RNA of any one of claims 1 to 5, which is for producing a vaccine against hepatitis C virus infection.

[Claim 13]

The replicon-replicating cell of any one of claims 6 to 9, which is for producing a vaccine against hepatitis C virus infection.

[Claim 14]

A method of producing a replicon RNA of hepatitis C virus of genotype 2a, comprising extracting the replicon RNA from the replicon-replicating cell of any

one of claims 6 to 9.

[Claim 15]

A method of producing a viral protein of hepatitis C virus of genotype 2a, comprising culturing the replicon-replicating cell of any one of claims 6 to 9, and obtaining the viral protein from the resulting culture product.

[Claim 16]

A method of screening for a substance promoting or suppressing the replication of hepatitis C virus, comprising culturing the replicon-replicating cell of any one of claims 6 to 9 in the presence of a test substance, and detecting the replication of a replicon RNA in the resulting culture product.

[Claim 17]

A method of increasing the replication efficiency of the replicon RNA of hepatitis C virus of genotype 2a, comprising performing once or more the following: obtaining a replicated replicon RNA from the replicon-replicating cell of any one of claims 6 to 9, and introducing the thus obtained replicated replicon RNA into a cell that is different from the replicon-replicating cell so as to prepare a new replicon-replicating cell.

[Claim 18]

The method of claim 17, wherein the replication efficiency increases to become at least two times greater than that of the replicon RNA that is introduced at the beginning into the replicon-replicating cell.

[Claim 19]

A method of producing a replicon RNA of hepatitis C virus of genotype 2a having increased replication efficiency, comprising performing once or more the following: obtaining a replicated replicon RNA from the replicon-replicating cell of any one of claims 6 to 9, and introducing the thus obtained replicated replicon RNA into a cell that is different from the replicon-replicating cell so as to prepare a new replicon-replicating cell; and obtaining a replicated replicon RNA from the finally obtained replicon-replicating cell.

[Claim 20]

A method of producing a replicon RNA of hepatitis C virus of genotype 2a having increased replication efficiency, comprising detecting a nucleotide mutation or an amino acid mutation between the replicon RNA that is produced so as to have an increased replication efficiency by the method of claim 19 and the replicon RNA that is introduced at the beginning into the replicon-replicating cell; and introducing the thus detected nucleotide mutation or amino acid mutation into a replicon RNA whose replication efficiency is to be increased.

[Claim 21]

A replicon RNA, comprising a nucleotide sequence derived from the nucleotide sequence represented by SEQ ID NO: 1 by at least one mutation selected from the group consisting of the following (a) to (u):

- (a) a mutation from A to G at nucleotide site 7157;
- (b) a mutation from C to U at nucleotide site 4955;
- (c) a mutation from A to G at nucleotide site 4936;
- (d) a mutation from A to G at nucleotide site 5000;
- (e) a mutation from A to G at nucleotide site 7288;
- (f) a mutation from G to U at nucleotide site 5901;
- (g) a mutation from A to U at nucleotide site 6113;
- (h) a mutation from A to G at nucleotide site 2890;
- (i) a mutation from C to A at nucleotide site 6826;
- (j) a mutation from C to A at nucleotide site 6887;
- (k) a mutation from U to A at nucleotide site 6580;
- (l) a mutation from U to C at nucleotide site 7159;
- (m) a mutation from U to A at nucleotide site 7230;
- (n) a mutation from C to A at nucleotide site 6943;
- (o) a mutation from G to A at nucleotide site 5687;
- (p) a mutation from A to G at nucleotide site 6110;
- (q) a mutation from U to C at nucleotide site 5550;

- (r) a mutation from A to G at nucleotide site 7217;
- (s) a mutation from A to G at nucleotide site 3643;
- (t) a mutation from G to A at nucleotide site 5851; and
- (u) a mutation from G to A at nucleotide site 5914.

[Title of Document] DESCRIPTION

[Title Of Invention] A NUCLEIC ACID CONSTRUCT CONTAINING A NUCLEIC ACID DERIVED FROM THE GENOME OF HEPATITIS C VIRUS (HCV) OF GENOTYPE 2a, AND A CELL HAVING SUCH NUCLEIC ACID CONSTRUCT INTRODUCED THEREIN

[Technical Field]

[0001]

The present invention relates to a replicon RNA of the hepatitis C virus of genotype 2a, a replicon-replicating cell wherein the replicon RNA is introduced, and a method of increasing the replication efficiency of the replicon RNA.

[Background Art]

[0002]

The hepatitis C virus (HCV) is a virus belonging to the family Flaviviridae. It has a single-stranded (+) strand sense RNA as its genome and is known to cause hepatitis C. Recent studies have revealed that Hepatitis C virus is classified into a number of types based on genotypes or serotypes. According to the phylogenetic analysis of Simmonds et al., using the nucleotide sequences of the HCV strains, which is currently a mainstream method of classifying HCV genotypes, HCV is classified into 6 genotypes: genotype 1a, genotype 1b, genotype 2a, genotype 2b, genotype 3a and genotype 3b (see Non Patent Literature 1). Each of these types is further classified into several subtypes. The nucleotide sequences of the full-length genomes of a several number of genotypes of HCV have been determined to date (see Patent Literature 1 and Non Patent Literatures 2-5).

[0003]

HCV causes chronic hepatitis by persistent infection. Currently, the main cause of chronic hepatitis observed worldwide is persistent HCV infection. Actually, around 50% of individuals with persistent infection develop chronic hepatitis. Chronic hepatitis in approximately 20% of these patients shifts to liver

cirrhosis over the course of 10 to 20 years, and some of these patients further go on to advanced lethal pathological conditions such as hepatic cancer.

[0004]

Hepatitis C is currently treated mainly by a therapy using interferon- α or interferon- β , or a therapy using in combination interferon- α and ribavirin, the purine-nucleoside derivative. However, even when these therapies are performed, the therapeutic effects are observed in only approximately 60% of all the treated patients. When the therapies are ceased after the exertion of the effects, the disease recrudesces in more than half of the patients. The therapeutic effect of interferones is known to relate to HCV genotypes, and is said to be lower against genotype 1b and higher against genotype 2a (see Non Patent Literature 6).

[0005]

It is an important goal to develop therapeutic agents or prophylactic agents effective against hepatitis C, the incidence rate of which is high in industrial countries, for which currently no causal treatment are present, and which finally bring about serious results. Hence, the development of HCV-specific chemotherapies and vaccine therapies are earnestly desired. A target for the development of an anti-HCV agent may be the suppression of HCV replication or the suppression of infection of cells with HCV.

[0006]

Until recently, propagation of HCV in a cell culture system and infecting cultured cells with HCV have been difficult. Moreover, a chimpanzee has been the only animal that can be infected with HCV and can be used in experiments, so that it has been difficult to carry out studies on the replication mechanism of HCV and the infection mechanism of HCV. However, recently, HCV subgenomic RNA replicons have been prepared as HCV-derived autonomously replicable RNA (see Patent Literature 2 and Non Patent Literatures 7-10), which enables the analysis of the replication mechanism of HCV using cultured cells. These HCV subgenomic RNA replicons are each prepared by substituting structural proteins existing

downstream of HCV IRES in the 5' untranslated region of the HCV genomic RNA of genotype 1b with a neomycin resistance gene and EMCV IRES that has been ligated downstream of the resistance gene. It has been demonstrated that this RNA replicon is autonomously replicated in human hepatic cancer cells, Huh7 cells, when introduced into the Huh7 cells followed by culture in the presence of neomycin.

[0007]

However, regarding such intracellular RNA replication systems for HCV, only those using HCV genomic RNA of genotype 1b have been prepared so far. Since there has been a report that different genotypes of HCV differ also in viral proteins encoded, it may be difficult to sufficiently elucidate the replication mechanism of HCV only by analyzing the subgenomic RNA replicons derived from HCV of genotype 1b. Furthermore, based on the fact that the therapeutic effects of interferons differ depending on the HCV genotypes, it may be particularly difficult to develop an anti-HCV agent having an effect on various types of HCV by the use of only an HCV replication system containing the subgenomic RNA replicon of HCV of genotype 1b.

[0008]

[Patent Literature 1] JP Patent Publication (Kokai) No. 2002-171978 A

[Patent Literature 2] JP Patent Publication (Kokai) No. 2001-17187 A

[Non Patent Literature 1] Simmonds, P. et al, Hepatology, (1994) 10, pp. 1321-1324

[Non Patent Literature 2] Choo et al., Science, (1989) 244, pp. 359-362

[Non Patent Literature 3] Kato et al., J. Med. Virol., (2001) 64(3) pp. 334-339

[Non Patent Literature 4] Okamoto, H et al, J. Gen. Virol., (1992) 73 pp. 673-679

[Non Patent Literature 5] Mori, S. et al, Biochem. Biophys. Res. Commun., (1992) 183, pp. 334-342

[Non Patent Literature 6] Yoshioka et al., Hepatology, (1992) 16(2): pp. 293-299

[Non Patent Literature 7] Lohmann et al., Science, (1999) 285, pp. 110-113

[Non Patent Literature 8] Blight et al., Science, (2000) 290, pp. 1972-1974

[Non Patent Literature 9] Friebe et al., J. Virol., (2001) 75(24): pp. 12047-12057

[Non Patent Literature 10] Ikeda et al., J. Virol., (2002) 76(6): pp. 2997-3006

[Disclosure of Invention]

[Problem to be Solved by Invention]

[0009]

An object of the present invention is to provide an HCV-derived replicon RNA of a HCV genotype for which replicon RNA has not yet been prepared.

[Means for Solving the Problem]

[0010]

As a result of intensive studies to achieve the above object, we have succeeded in preparing the replicon RNA of HCV genotype 2a.

[0011]

That is, the present invention is as follows.

[1] A replicon RNA, comprising a nucleotide sequence containing at least the 5' untranslated region, the nucleotide sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein and the 3' untranslated region on the genomic RNA of hepatitis C virus of genotype 2a. Preferably, this replicon RNA further contains at least one selection marker gene or a reporter gene, and at least one IRES sequence.

[2] A replicon RNA, comprising a nucleotide sequence containing the 5' untranslated region comprising the nucleotide sequence represented by either SEQ ID NO: 9 or 10; at least one selection marker gene or a reporter gene; an IRES sequence; the nucleotide sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein on the genomic RNA of hepatitis C virus of genotype 2a; and the 3' untranslated region comprising the nucleotide sequence represented by either SEQ ID NO: 11 or 12.

[3] The replicon RNA of [1] or [2] above, wherein the genomic RNA of hepatitis C virus of genotype 2a is an RNA comprising the nucleotide sequence represented by

SEQ ID NO: 3 or 5.

[4] A replicon RNA, comprising the following RNA (a) or (b):

(a) an RNA comprising the nucleotide sequence represented by SEQ ID NO: 1 or 2; and

(b) an RNA comprising a nucleotide sequence derived from the nucleotide sequence represented by SEQ ID NO: 1 or 2 by deletion, substitution or addition of 1 to 10 nucleotides, and being capable of autonomous replication.

[5] A replicon-replicating cell, which is prepared by introducing the replicon RNA of any one of [1] to [4] above into a cell. For this replicon-replicating cell, a cell into which the replicon RNA is introduced is preferably a eukaryotic cell, more preferably a human liver-derived cell, a human uterine cervix-derived cell or a human fetal kidney-derived cell, and further more preferably any one cell selected from the group consisting of an Huh7 cell, an HepG2 cell, an IMY-N9 cell, an HeLa cell and a 293 cell.

[6] The replicon RNA of [1] to [4] above, which is for producing or evaluating a therapeutic agent or a diagnostic agent against hepatitis C virus infection.

[7] The replicon-replicating cell of [5] above, which is for producing or evaluating a therapeutic agent or a diagnostic agent against hepatitis C virus infection.

[8] The replicon RNA of [1] to [4] above, which is for producing a vaccine against hepatitis C virus infection.

[9] The replicon-replicating cell of [5] above, which is for producing a vaccine against hepatitis C virus infection.

[10] A method of producing a replicon RNA of hepatitis C virus of genotype 2a, comprising extracting the replicon RNA from the replicon-replicating cell of [5] above.

[11] A method of producing a viral protein of hepatitis C virus of genotype 2a, comprising culturing the replicon-replicating cell of [5] above, and obtaining the viral protein from the resulting culture product.

[12] A method of screening for a substance promoting or suppressing the

replication of hepatitis C virus, comprising culturing the replicon-replicating cell of [5] above in the presence of a test substance, and detecting the replication of a replicon RNA in the resulting culture product.

[13] A method of increasing the replication efficiency of the replicon RNA of hepatitis C virus of genotype 2a, comprising performing once or more the following: obtaining a replicated replicon RNA from the replicon-replicating cell of [5] above, and introducing the thus obtained replicated replicon RNA into a cell that is different from the replicon-replicating cell so as to prepare a new replicon-replicating cell. In this method, it is more preferred that the replication efficiency increases to become preferably at least two times greater than that of the replicon RNA that is introduced at the beginning into the replicon-replicating cell.

[14] A method of producing a replicon RNA of hepatitis C virus of genotype 2a having increased replication efficiency, comprising performing once or more the following: obtaining a replicated replicon RNA from the replicon-replicating cell of [5] above, and introducing the thus obtained replicated replicon RNA into a cell that is different from the replicon-replicating cell so as to prepare a new replicon-replicating cell; and obtaining a replicated replicon RNA from the finally obtained replicon-replicating cell.

[15] A method of producing a replicon RNA of hepatitis C virus of genotype 2a having increased replication efficiency, comprising detecting a nucleotide mutation or an amino acid mutation between the replicon RNA that is produced so as to have an increased replication efficiency by the method of [14] above and the replicon RNA that is introduced at the beginning into the replicon-replicating cell; and introducing the thus detected nucleotide mutation or amino acid mutation into a replicon RNA whose replication efficiency is to be increased.

[16] A replicon RNA, comprising a nucleotide sequence derived from the nucleotide sequence represented by SEQ ID NO: 1 by at least one mutation selected from the group consisting of the following (a) to (u):

- (a) a mutation from A to G at nucleotide site 7157;
- (b) a mutation from C to U at nucleotide site 4955;
- (c) a mutation from A to G at nucleotide site 4936;
- (d) a mutation from A to G at nucleotide site 5000;
- (e) a mutation from A to G at nucleotide site 7288;
- (f) a mutation from G to U at nucleotide site 5901;
- (g) a mutation from A to U at nucleotide site 6113;
- (h) a mutation from A to G at nucleotide site 2890;
- (i) a mutation from C to A at nucleotide site 6826;
- (j) a mutation from C to A at nucleotide site 6887;
- (k) a mutation from U to A at nucleotide site 6580;
- (l) a mutation from U to C at nucleotide site 7159;
- (m) a mutation from U to A at nucleotide site 7230;
- (n) a mutation from C to A at nucleotide site 6943;
- (o) a mutation from G to A at nucleotide site 5687;
- (p) a mutation from A to G at nucleotide site 6110;
- (q) a mutation from U to C at nucleotide site 5550;
- (r) a mutation from A to G at nucleotide site 7217;
- (s) a mutation from A to G at nucleotide site 3643;
- (t) a mutation from G to A at nucleotide site 5851; and
- (u) a mutation from G to A at nucleotide site 5914.

[Effects of Invention]

[0012]

According to the present invention, an HCV-RNA replicon derived from the genotype 2a strain of HCV has been provided for the first time. The replicon-replicating cell according to the present invention can be used as a culture system for the continuous production of RNA and HCV proteins derived from HCV of genotype 2a. Furthermore, the replicon-replicating cell according to the present invention is useful as a test system for screening for various substances that affect

HCV replication and/or the translation of HCV proteins.

[Best Mode for Carrying out Invention]

[0013]

The present invention is explained in detail as follows.

[0014]

1. HCV-derived replicon RNA according to the present invention

The genome of hepatitis C virus (HCV) is a single-stranded (+) strand RNA comprising approximately 9600 nucleotides. This genomic RNA comprises the 5' untranslated region (also denoted as 5' NTR or 5' UTR), a translated region composed of a structural region and a non-structural region and the 3' untranslated region (also denoted as 3' NTR or 3' UTR). HCV structural proteins are encoded in the structural region, and a plurality of non-structural proteins are encoded in the non-structural region.

[0015]

Such HCV structural proteins and non-structural proteins are generated through the translation into a continuous form thereof, a polyprotein, from the translated region, restricted degradation of the polyprotein by protease, and then the release of the structural proteins (Core, E1 and E2) and non-structural proteins (NS2, NS3, NS4A, NS4B, NS5A and NS5B), respectively. Among these structural proteins and non-structural proteins, that is, viral proteins of HCV, Core is a core protein, E1 and E2 are envelope proteins, and non-structural proteins (NS2, NS3, NS4A, NS4B, NS5A and NS5B) are proteins involved in virus's own replication. NS2 is known to have metalloprotease activity, and NS3 is known to have serine protease activity (at one-third of the N terminal side) and helicase activity (at two-thirds of the C-terminal side). Furthermore, NS4A is a cofactor for protease activity of NS3, and NS5B has been reported to have RNA-dependent RNA polymerase activity. Furthermore, the genome of HCV of genotype 2a has already been reported to have a similar gene structure (see Patent Literature 1).

[0016]

We have constructed RNA capable of autonomous replication using such HCV genome of genotype 2a. Specifically, the HCV-derived replicon RNA of the present invention is an RNA construct, which contains the whole or partial RNA of the HCV genome of genotype 2a and is capable of autonomous replication.

[0017]

In this specification, RNA that is prepared by altering the viral genome of HCV and is capable of autonomous replication is referred to as "replicon RNA" or "RNA replicon." RNA that is artificially prepared from HCV of genotype 2a and is capable of autonomous replication is referred to as "replicon RNA derived from HCV of genotype 2a." In this specification, the HCV-derived replicon RNA is also referred to as an HCV-RNA replicon.

[0018]

In the present invention, "hepatitis C virus of genotype 2a" or "HCV of genotype 2a" means hepatitis C virus identified as genotype 2a according to the international classification of Simmonds et al. The "hepatitis C virus of genotype 2a" or the "HCV of genotype 2a" of the present invention encompasses not only a virus having naturally occurring HCV genomic RNA, but also a virus having genomic RNA prepared by artificially altering a naturally occurring HCV genomic sequence. Specific examples of HCV of genotype 2a include viruses of JFH-1 strain and the JCH-1 strain (see Patent Literature 1).

[0019]

Furthermore, "the genomic RNA of hepatitis C virus of genotype 2a" means RNA that comprises the single-stranded (+) strand sense RNA of hepatitis C virus of genotype 2a and has the nucleotide sequence throughout the entire region of its genome. The genomic RNA of hepatitis C virus of genotype 2a is preferably RNA comprising the nucleotide sequence represented by SEQ ID NO: 3 or 5, but is not limited thereto.

[0020]

In the specification of the present application, "5' untranslated region"

(5'NTR or 5'UTR), "a sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein," "a sequence encoding Core protein" (Core region or C region), "a sequence encoding E1 protein" (E1 region), "a sequence encoding E2 protein" (E2 region), "a sequence encoding NS2 protein" (NS2 region), "a sequence encoding NS3 protein" (NS3 region), "a sequence encoding NS4A protein" (NS4A region), "a sequence encoding NS4B protein" (NS4B region), "a sequence encoding NS5A protein" (NS5A region), "a sequence encoding NS5B protein" (NS5B region) and "3' untranslated region" (3' NTR or 3' UTR), and other specific regions or sites are determined based on the nucleotide sequence of SEQ ID NO: 3 of the full-length cDNA (JFH-1 clone) encoding the entire region of the genome of the JFH-1 strain, which is HCV of genotype 2a. The nucleotide sequence of SEQ ID NO: 3 can be obtained from the International DNA Data Bank (DDBJ/EMBL/GenBank) by referring to the accession No. AB047639. Specifically, when a particular HCV RNA sequence is aligned with the nucleotide sequence represented by SEQ ID NO: 3, a sequence to be aligned with nucleotides 1 to 340 on the nucleotide sequence represented by SEQ ID NO: 3 is "5' untranslated region" of the RNA, a sequence to be aligned with the nucleotides 3431 to 9442 on the same are a sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein, a sequence to be aligned with the nucleotides 3431 to 5323 on the same is "a sequence encoding NS3 protein," a sequence to be aligned with the nucleotides 5324 to 5485 on the same is "a sequence encoding NS4A protein," a sequence to be aligned with the nucleotides 5486 to 6268 on the same is a sequence encoding NS4B protein," a sequence to be aligned with the nucleotides 6269 to 7666 on the same is "a sequence encoding NS5A protein," a sequence to be aligned with the nucleotides 7667 to 9442 on the same is "a sequence encoding NS5B protein," and a sequence to be aligned with the nucleotides 9443 to 9678 on the same is "3' untranslated region." Furthermore, in this case, gaps, additions, deletions, substitutions or the like may be present in the "aligned" sequences. Furthermore, the above

"particular HCV" is not limited thereto, and includes the JFH-1 strain or JCH-1 strain, or viral strains that are derivatives thereof.

[0021]

One embodiment of the HCV RNA-replicon according to the present invention is a replicon RNA comprising a nucleotide sequence containing at least the 5' untranslated region, a sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein, and the 3' untranslated region on the genomic RNA of hepatitis C virus of genotype 2a. The replicon RNA may further contain at least one selection marker gene or one reporter gene, and at least one IRES sequence. Furthermore, this replicon RNA may also contain a sequence encoding a viral protein other than NS3, NS4A, NS4B, NS5A and NS5B proteins on the genomic RNA of hepatitis C virus of genotype 2a.

[0022]

Another preferred embodiment of HCV RNA-replicon according to the present invention is a replicon RNA comprising a nucleotide sequence containing the 5' untranslated region comprising the nucleotide sequence represented by SEQ ID NO: 9 or 10, at least one selection marker gene or reporter gene, the IRES sequence, a sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein on the genomic RNA of hepatitis C virus of genotype 2a, and the 3' untranslated region comprising the nucleotide sequence represented by SEQ ID NO: 11 or 12. In this case the nucleotide sequences represented by SEQ ID NO: 9 and 10 are sequences of the 5' untranslated regions of rSGREP-JFH1 (SEQ ID NO: 1) and rSGREP-JCH1 (SEQ ID NO: 2), respectively, which are replicon RNAs according to the present invention. Furthermore, the nucleotide sequences represented by SEQ ID NO: 11 and 12 are sequences of the 3' untranslated regions of rSGREP-JFH1 (SEQ ID NO: 1) and rSGREP-JCH1 (SEQ ID NO: 2), respectively, which are replicon RNAs according to the present invention.

[0023]

A more preferred embodiment of HCV RNA-replicon according to the present invention is a replicon RNA comprised of an RNA comprising the nucleotide sequence represented by SEQ ID NO: 1 or 2. Furthermore, a replicon RNA comprising a nucleotide sequence derived from the nucleotide sequence represented by SEQ ID NO: 1 or 2 by deletion, substitution or addition of 1 to 50, 1 to 30, 1 to 10, 1 to 6, or 1 to several (2 to 5) nucleotides, and being capable of autonomous replication is also included in the scope of the present invention as a preferred embodiment. In the present invention, "capable of autonomous replication" means that when replicon RNA is introduced into a cell, the replicon RNA allows its own full-length sequence to be replicated within the cell. For example, this ability of autonomous replication can be confirmed by transfecting replicon RNA into Huh7 cells, culturing the Huh7 cells, extracting RNA from the cells in the thus resulting culture product and conducting Northern blot hybridization for the extracted RNA using a probe that can specifically detect the transfected replicon RNA so as to detect the presence of the replicon RNA. However, examples of such a method are not limited thereto. Specific procedures for confirming the ability of autonomous replication can be conducted according to descriptions given in the Examples of this specification such as those for measuring the ability of colony formation, those for confirming the expression of HCV proteins or those for detecting replicon RNA.

[0024]

In the present invention, a "selection marker gene" means a gene that can provide a cell with selectivity such that only the cell expressing the gene is selected. A general example of a selection marker gene is an antibiotic resistance gene. In the present invention, preferred examples of a selection marker gene include a neomycin resistance gene, a thymidine kinase gene, a kanamycin resistance gene, a pyrithiamine resistance gene, an adenylyl transferase gene, a Zeocin resistance gene and a puromycin resistance gene. The neomycin resistance gene and the thymidine kinase gene are preferred, and the neomycin

resistance gene is more preferred. However, the selection marker gene in the present invention is not limited to these genes.

[0025]

Furthermore in the present invention, a "reporter gene" means a marker gene encoding a gene product that is a marker for the expression of the gene. General examples of a reporter gene include structural genes of enzymes that catalyze light emitting reaction or color reaction. Preferred examples of the reporter gene in the present invention include a transposon Tn9-derived chloramphenicol acetyltransferase gene, an *Escherichia coli*-derived β glucuronidase or β galactosidase gene, a luciferase gene, a green fluorescence protein gene, an aequorin gene from jellyfish, and a secreted placental alkaline phosphatase (SEAP) gene. However, the reporter gene in the present invention is not limited to these genes.

[0026]

Either only one or both of the above selection marker gene and reporter gene may be contained in replicon RNA.

[0027]

In the present invention, "IRES sequence" means an internal ribosome entry site that allows translation to be initiated by binding ribosomes within the inside of RNA. Preferred examples of IRES sequence in the present invention include, but are not limited to, EMCV IRES (the internal ribosome entry site of encephalomyocarditis virus), FMDV IRES and HCV IRES. EMCV IRES and HCV IRES are more preferred, and EMCV IRES is the most preferred sequence.

[0028]

The replicon RNA according to the present invention may further contain a sequence on the genomic RNA of another HCV strain or HCV of another genotype. For example, the replicon RNA may also contain a fragment of HCV genome of genotype 1b. Examples of another HCV strain include, but are not limited to, HCV-1, HCV-H, HC-J1, HCT-18, H77, DK-7, US11, S14, HCT23, HCV-Th, DR1,

DR4, HCT27, S18, SW1, DK9, H90, TD-6E1, S9, HCV-BK, T10, DK1, HC-J4, HCV-J, HK3, HK8, HK5, HCV-G3, IND5, IND8, P10, D1, D3, SW2, T3, S45, SA10, US6, HCV-JK1, HCV-JK4, HCV-JK3, HCV-JK2, HCV-JT, HC-J2, HCV-T, HK4, HC-G9, Z1, Bi, S. I., Cho, J.M., HCV-J6, T4, T9, US10, HC-J5, T2, HC-J7, DK11, SW3, DK8, T8, HC-J8, S83, HK2, HC-J6, HC-J8, BEBE1, HCV-J6, HCV-J8, HD10-2, BR36-9, S52, S54, S2, BR33-1, HK10, DK12, HCV-TR, BA-1, BA-2, DK13, Z1, Z4, Z6, Z7, HK2, SA1, SA4, SA5, SA7, SA13, SA6, NZL1, SA30, EG-13, HCV-K3a/650, ED43, EUH1480, EUHK2, Th580, VN235, VN405, VN004, JK049, JK046, JFH-1, JCH-1, JCH-2, JCH-3, JCH-4, JCH-5, JCH-6, J6CF and H77.

[0029]

The replicon RNA according to the present invention preferably has the 5' untranslated region on the genomic RNA of HCV of genotype 2a on the 5'-most side, and the 3' untranslated region on the genomic RNA of HCV of genotype 2a on the 3'-most side. A selection marker gene or a reporter gene may be ligated upstream of the IRES sequence, or upstream or downstream of "the sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein," or inserted in the middle of "the sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein."

[0030]

The replicon RNA according to the present invention more preferably has the 5' untranslated region on the genomic RNA of HCV of genotype 2a on the 5'-most side, and a selection marker gene or a reporter gene, the IRES sequence and "the sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein" downstream of the 5' untranslated region in this order, and the 3' untranslated region on the genomic RNA of HCV of genotype 2a on the 3'-most side.

[0031]

Examples of the replicon RNA according to the present invention may

include an RNA containing any foreign gene to be expressed within a cell into which the replicon RNA is introduced, in addition to the sequences as described above. A foreign gene may also be ligated downstream of the 5' untranslated region, or ligated upstream or downstream of a selection marker gene or a reporter gene, or ligated upstream or downstream of "the sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein," or may be inserted in the middle of "the sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein." A replicon RNA containing a foreign gene can express a protein encoded by the foreign gene when it is translated within a cell into which the RNA is introduced. Thus, the replicon RNA containing a foreign gene can be appropriately used also for gene therapy or the like, the purpose of which is to generate a particular gene product within a cell.

[0032]

The replicon RNA according to the present invention may further contain a ribozyme. A ribozyme is inserted to ligate a selection marker gene, a reporter gene or a foreign gene on the 5' side in the replicon RNA to those located on the 3' side thereof including the IRES sequence and "the sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein," so that it enables cleavage and separation of the two by the self-cleavage activity of the ribozyme.

[0033]

In the replicon RNA according to the present invention, the above described selection marker gene, reporter gene, sequences encoding viral proteins on the genomic RNA of hepatitis C virus of genotype 2a, sequences encoding viral proteins of HCV of a genotype other than genotype 2a, a foreign gene or the like are ligated so that they are translated from the replicon RNA in the correct reading frame. Among these sequences, the protein-coding sequences may be ligated to each other via a protease cleavage site and the like, so that after the proteins are expressed as a fusion protein with the polyprotein that is translated from "the sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and

NS5B protein" of hepatitis C virus of genotype 2a, the fusion protein is separated by protease into each protein.

[0034]

2. Preparation of replicon RNA according to the present invention

The HCV RNA-replicon according to the present invention can be prepared using any genetic engineering techniques known by persons skilled in the art. The HCV RNA-replicon can be prepared by, for example, the following method, but the method of preparation is not limited thereto.

[0035]

First, DNA corresponding to the entire region of the genomic RNA of hepatitis C virus of genotype 2a is ligated downstream of an RNA promoter according to a standard procedure so as to prepare a DNA clone. As used herein, "DNA corresponding to RNA" means a DNA having a nucleotide sequence derived from the nucleotide sequence of the RNA by substituting U (uracil) with T (thymine). The above RNA promoter is preferably an RNA promoter contained in a plasmid clone. An example of an RNA promoter is not limited, but T7 RNA promoter is particularly preferred.

[0036]

Next, for the thus prepared DNA clone, for example, the structural region (Core sequence, E1 sequence and E2 sequence) located downstream of the 5' untranslated region and the sequence encoding NS2 protein are substituted with a DNA fragment containing a selection marker gene or a reporter gene and the IRES sequence ligated downstream thereof. In this substitution, portions other than the structural region, such as a fragment on the 3' terminal side of the 5' untranslated region or a part of the sequence encoding NS3 protein may be substituted with a sequence derived from HCV of another genotype.

[0037]

Subsequently, using the DNA clone after the substitution as a template, RNA is synthesized using RNA polymerase. RNA synthesis can be initiated by a

standard procedure from the 5' untranslated region and the IRES sequence. When a template DNA is a plasmid clone, the above DNA region ligated downstream of an RNA promoter is excised by a restriction enzyme from the plasmid clone, and then RNA can be synthesized using the DNA fragment as a template. In addition, preferably the 3' terminus of RNA to be synthesized agrees with the 3' untranslated region of the viral genomic RNA, and no other sequences are added or deleted. The thus synthesized RNA is the replicon RNA according to the present invention.

[0038]

3. Preparation of replicon-replicating cells into which replicon RNA from HCV of genotype 2a is introduced

The replicon RNA that is prepared as described above is introduced into cells in which the replicon RNA should be replicated, so that cells wherein the replicon RNA is continuously replicated can be obtained. In this specification, a cell wherein replicon RNA is continuously replicated is referred to as a "replicon-replicating cell."

[0039]

As a cell into which replicon RNA is introduced, any cell can be used, as long as it can be subcultured. Such a cell is preferably a eukaryotic cell, more preferably a human liver-derived cell, a human uterine cervix-derived cell or a human fetal kidney-derived cell, and further preferably any cell selected from the group consisting of Huh7 cells, HepG2 cells, IMY-N9 cells, HeLa cells and 293 cells. As these cells, commercially available cells may be utilized, these cells may be obtained from cell depositories, or cell lines established from any cells (e.g., cancer cells or stem cells) may also be used.

[0040]

As the above cells, cells that can be mass-cultured are preferably used for the purpose of the mass production of HCV proteins, such as in the case of vaccine production. From such a viewpoint, the cells are preferably those other than Huh7 cells.

[0041]

Introduction of replicon RNA into cells can be performed using any technique known by persons skilled in the art. Examples of such an introduction method include electroporation, a particle gun method, a lipofection method, a calcium phosphate method, a microinjection method and a DEAE sepharose method. The method using electroporation is particularly preferred.

[0042]

A replicon RNA of interest may be introduced alone, or may be introduced after it is mixed with other nucleic acids. To vary the quantity of replicon RNA while keeping RNA quantity to be introduced at a certain level, the replicon RNA of interest is mixed with total cellular RNA extracted from cells into which the RNA is introduced, and then the mixture is used for introduction into cells. The quantity of replicon RNA to be used for introduction into cells may be determined depending on the introduction method employed, and is preferably between 1 picogram and 100 micrograms, and more preferably between 10 picograms and 10 micrograms.

[0043]

When replicon RNA containing a selection marker gene or a reporter gene is used for introduction into cells, cells wherein the replicon RNA is introduced and continuously replicated can be selected utilizing the expression of the selection marker gene or the reporter gene. Specifically, for example, such cells into which replicon RNA has been introduced may be cultured in media whereby the cells can be selected by the expression of the selection marker gene or the reporter gene. As an example, when replicon RNA contains a neomycin resistance gene as a selection marker gene, cells into which replicon RNA has been intracellularly introduced are seeded into a culture dish. After 16 to 24 hours of culture, G418 (neomycin) is added to the culture dish at a concentration of 0.05 milligrams/milliliter to 3.0 milligrams/milliliter. The cells are continuously cultured for preferably 10 days to 40 days and more preferably 14

days to 28 days after seeding, while exchanging the culture solution twice a week. Next, surviving cells are stained with crystal violet, so that cells into which the replicon RNA has been introduced and is being continuously replicated can be selected as formed colonies.

[0044]

Cloned cells can be obtained from the formed colonies by cloning surviving cells by a standard procedure, and then continuing the culture of the cells. The thus obtained cell clone wherein the replicon RNA of interest is continuously replicated is referred to as "a replicon-replicating cell clone" in this specification.

[0045]

Regarding the established cell clone, detection of a replicon RNA that has been replicated from the introduced replicon RNA in the cell clone, confirmation of the presence or the absence of the incorporation of a selection marker gene or a reporter gene in the introduced replicon RNA into a host genomic DNA, and confirmation of the expression of an HCV protein are preferably carried out to confirm the fact that a replicon RNA of interest is actually and continuously replicated.

[0046]

A replicon RNA that has been replicated from the introduced replicon RNA in the cell clone (in this specification, hereinafter conveniently referred to as "replicated replicon RNA") may be detected according to any RNA detection method known by persons skilled in the art. For example, detection can be performed by conducting the Northern hybridization method for total RNA extracted from the cell clone using as a probe a DNA fragment specific to the introduced replicon RNA.

[0047]

Furthermore, the presence or the absence of the incorporation of a selection marker gene or a reporter gene in the introduced replicon RNA into a

host genomic DNA can be confirmed by, for example, performing PCR for the host genomic DNA extracted from the cell clone to amplify at least a part of the selection marker gene or the reporter gene, and then confirming the presence or the absence of the amplified product. However, examples of relevant methods are not limited thereto. A cell clone for which the amplified product is confirmed is considered to have a selection marker gene or a reporter gene incorporated in the host genome. Thus, regarding the cell clone, the replicon RNA itself may not be continuously replicated within the cell. In this case, whether or not the replicon RNA is continuously replicated can be confirmed by conducting an experiment to confirm the expression of an HCV protein, as described below.

[0048]

The expression of an HCV protein can be confirmed by, for example, causing an antibody against an HCV protein to be expressed from the introduced replicon RNA and to react with a protein extracted from a cell clone. This method can be conducted by any protein detection method known by persons skilled in the art. Specifically, for example, a protein sample extracted from the cell clone is blotted onto a nitrocellulose membrane, with which an anti-HCV protein antibody (e.g., an anti-NS3-specific antibody or an antiserum collected from a hepatitis C patient) is reacted, and then the anti-HCV protein antibody is detected. If the HCV protein is detected among proteins extracted from the cell clone, it can be concluded that this cell clone continuously replicate HCV-derived replicon RNA to express the HCV protein.

[0049]

As described above, cell clones confirmed to continuously replicate a replicon RNA of interest (replicon-replicating cell clones) can be obtained. Furthermore in the present invention, replicon RNA can be obtained by any method known by persons skilled in the art, for example, by extracting RNA from the replicon-replicating cell, and then separating replicon RNA from the RNA by an electrophoresis method. The present invention also relates to such a method

of producing replicon RNA. Moreover, preferably, the replicon-replicating cell according to the present invention can be used for producing HCV proteins. Persons skilled in the art can obtain HCV proteins from the replicon-replicating cells according to any standard method. Specifically, for example, a viral protein of hepatitis C virus of genotype 2a can be produced by culturing replicon-replicating cells, collecting proteins from the resulting culture product (including cultured cells and culture media) by a standard procedure, and then selectively obtaining viral proteins from the proteins by detection or the like using an anti-HCV protein antibody.

[0050]

Moreover, when the replicon-replicating cell according to the present invention continuously replicates replicon RNA containing a foreign gene, a protein encoded by the foreign gene can be obtained by the expression thereof using the replicon-replicating cell. Specifically, for example, the protein encoded by a foreign gene can be obtained by culturing replicon-replicating cells, collecting proteins from the resulting culture product (including cultured cells and culture media) by a standard procedure, and then selectively obtaining the protein from among the proteins by detection or the like using an antibody against the protein of interest.

[0051]

4. Introduction of mutation that increases replication efficiency into replicon RNA from HCV of genotype 2a

Mutation producing enhancement of replication efficiency frequently takes place in the replicon RNA that is replicated or generated in the replicon-replicating cell (replicated replicon RNA) according to the present invention. Such a mutation may be an adaptive mutation.

Utilizing this fact, introduction of a mutation enhancing replication efficiency into the replicon RNA according to the present invention can be promoted in the present invention.

[0052]

Specifically, the step comprising obtaining a first replicated replicon RNA by extraction or the like from a first replicon-replicating cell (preferably, a replicon-replicating cell, wherein the replicon RNA according to the present invention has been introduced), and then re-introducing the first replicated replicon RNA into another cell to prepare a second replicon-replicating cell is performed repeatedly once or more, preferably 1 to 10 times, more preferably 1 to 5 times, and further preferably 1 to 2 times, so that the mutation increasing replication efficiency can be introduced at a high frequency into the replicon RNA within the replicon-replicating cells.

[0053]

As a cell into which a replicated replicon RNA is re-introduced, any cell can be used. Such a cell is preferably derived from a biological species that is the same as that of a cell wherein replicon RNA is introduced at the beginning, more preferably derived from the same tissue derived from the same biological species as that of a cell wherein replicon RNA is introduced at the beginning, and further preferably of a cell line that is the same as that for a cell wherein replicon RNA is introduced at the beginning.

[0054]

Therefore in the present invention, using the above method, replicon RNA wherein the mutation increasing replication efficiency is introduced can be produced. Specifically, the step comprising obtaining a first replicated replicon RNA by extraction or the like from a first replicon-replicating cell (preferably, a replicon-replicating cell, into which the replicon RNA according to the present invention has been introduced), and then re-introducing the first replicated replicon RNA into another cell so as to prepare a second replicon-replicating cell is performed repeatedly once or more, preferably 1 to 10 times, more preferably 1 to 5 times, and further preferably 1 to 2 times. Subsequently, the replicated replicon RNA is obtained by extraction or the like from the replicon-replicating

cell finally obtained at the end of the repeated steps, so that replicon RNA with increased replication efficiency can be produced.

[0055]

In the present invention, the replication efficiency of a replicon RNA can be increased at least 2 times, preferably 10 to 100 times, and more preferably 100 to 10000 times by the above method.

[0056]

Regarding the replicon RNA that is produced by such a method so as to have increased replication efficiency, the nucleotide sequence is preferably determined by a known method, for example, by obtaining cDNA by reverse transcription PCR and subjecting such cDNA to sequencing. Furthermore, the thus determined nucleotide sequence or the amino acid sequence encoded by the nucleotide sequence is compared with the nucleotide sequence of replicon RNA that had been introduced at the beginning into cells, so that adaptive mutation can be identified. As adaptive mutation increasing replication efficiency, in particular, nonsynonymous substitution that mutates an amino acid in a viral protein encoded by replicon RNA is preferred.

[0057]

The present invention also provides a method whereby the replicon RNA of hepatitis C virus of genotype 2a having increased replication efficiency can be produced by introducing the thus identified adaptive mutation into replicon RNA, the replication efficiency of which is to be increased, by a standard procedure.

[0058]

The replicon RNA that is produced as described above so as to have increased replication efficiency can be used for producing replicon RNA in large quantity within cells that have been used for the method.

[0059]

The replication efficiency of the replicon RNA according to the present invention can be determined by a method known by persons skilled in the art.

For example, it can be determined according to the following method. Replicon RNAs are transfected in quantities of 0.0001, 0.0003, 0.001, 0.003, 0.01, 0.03, 0.1, 0.3 and 1.0 micrograms, respectively, into Huh7 cells, selective culture with G418 is performed for 21 days in a method similar to the above experimental techniques, and then the number of colonies formed (number of colonies) is counted. The quantity of RNA introduced is compared with the number of colonies formed to determine the range of the quantity of the replicon RNA introduced, within which colony formation increases in a quantity-dependent manner. The number of colonies formed within the range is divided by the quantity of RNA introduced, and the resulting value is regarded as the colony forming activity per microgram. This equation is as follows.

$$\text{Colony forming activity [(Colony Forming Unit, or CFU)/microgram]} = \text{Number of colonies formed [colony]} / \text{quantity of RNA introduced [microgram]}$$

[0060]

The thus calculated colony forming activity is regarded as a value representing the replication efficiency of replicon RNA introduced. Specifically, the higher the colony forming activity, the higher the replication efficiency of the replicon RNA. In addition, the replication efficiency of replicon RNA can also be shown via a colony-forming ability that is represented by the number of copies of the replicon RNA introduced per formed colony. That is, in this case, the ability can be calculated according to the following equation.

$$\text{Colony forming ability} = \text{number of copies of replicon RNA introduced [copy]} / \text{number of formed colonies [colony]}$$

[0061]

5. Other embodiments of the present invention

The replicon RNA-replicating cell according to the present invention can also be used as a test system for, for example, screening for a substance that promotes or suppresses the replication of hepatitis C virus. Specifically, for example, replicon replicating cells are cultured in the presence of a test substance,

replication of the replicon RNA in the resulting culture product is detected, and then whether or not the test substance promotes or suppresses the replication of the replicon RNA is determined, so that a substance that promotes or suppresses the replication of hepatitis C virus can be screened for. In this case, detection of the replication of the replicon RNA in the resulting culture product may be conducted by detecting the quantity of, or the presence or the absence of, the replicon RNA in the RNAs extracted from the replicon RNA-replicating cell, or by detecting the quantity of, or the presence or the absence of, HCV protein contained in the proteins in the culture product or in the replicon RNA-replicating cells contained in the culture product.

[0062]

Such a test cell system using the replicon RNA-replicating cells according to the present invention may be aimed at producing or evaluating a therapeutic agent or a diagnostic agent for treating hepatitis C virus infection. Specific examples of such purposes include the following examples.

[0063]

(1) Search for a substance suppressing the proliferation of HCV of genotype 2a

Examples of a substance suppressing the proliferation of HCV of genotype 2a include organic chemicals directly or indirectly affecting the proliferation of HCV of genotype 2a, and antisense oligonucleotides directly or indirectly affecting the proliferation of HCV or the translation of HCV proteins by hybridizing to a target sequence in the HCV genome of genotype 2a or a complementary strand thereof.

(2) Evaluation of various substances having antiviral action in cell culture

Examples of the various substances include substances obtained through rational drug design or high throughput screening (e.g., an isolated and purified enzyme) and the like.

(3) Identification of a new target for attack for treating patients infected with HCV of genotype 2a

To identify a host cellular protein that plays an important role in proliferation of HCV virus, for example, the replicon-replicating cell according to the present invention can be used.

- (4) Evaluation of the ability of HCV virus to acquire resistance against a drug or the like and identification of mutation concerning such resistance
- (5) Production of a viral protein as an antigen that can be used for developing, producing and evaluating a diagnostic agent or a therapeutic agent for hepatitis C virus infection
- (6) Viral genome replication system for producing HCV virus or virus-like particles that can be used for developing, producing and evaluating a diagnostic agent or a therapeutic agent for hepatitis C virus infection
- (7) Production of a vaccine antigen that can be used as a vaccine against HCV of genotype 2a
- (8) Production of hepatic cell-directed genetic vector that is used after the incorporation of a foreign gene therein for gene therapy

[Examples]

[0064]

The present invention will be described more specifically based on the following examples and drawings. However, the technical scope of the present invention is not limited by these examples.

[0065]

[Example 1] Preparation of replicon RNA

(A) Construction of expression vector

DNA corresponding to the entire region of viral genome of hepatitis C virus JFH-1 strain (genotype 2a) that had been separated from patients with fulminant hepatic failure was obtained from a JFH-1 clone containing the full-length genomic cDNA of the virus strain. The DNA was inserted downstream of T7 RNA promoter sequence that had been inserted in pUC19 plasmid. The thus constructed plasmid DNA is hereinafter referred to as pJFH1. Similarly, DNA

corresponding to the entire region of viral genome of hepatitis C virus JCH-1 strain (genotype 2a) that had been separated from patients with chronic hepatitis was obtained from a JCH-1 clone containing the full-length genomic cDNA of the virus strain. The DNA was inserted downstream of the T7 RNA promoter sequence that had been inserted in pUC19 plasmid. The thus constructed plasmid DNA is hereinafter referred to as pJCH1. In addition, the preparation of the above JFH1 clone and JCH-1 clone is described in Patent Literature 1 and Non Patent Literature 3. Moreover, the nucleotide sequence of the full-length cDNA of JFH-1 clone was registered at the International DNA Data Bank (DDBJ/EMBL/GenBank) under accession No. AB047639, and the nucleotide sequence of the full-length cDNA of the JCH-1 clone under accession No. AB047640.

[0066]

The structures of the thus constructed plasmid DNA pJFH1 and pJCH1 are shown in the upper section of Fig. 1. "T7" represents T7 RNA promoter, and "G" represents dGTP inserted upstream of the 5' end of the inserted JFH-1- or JCH-1-derived DNA and downstream of the 3' end of T7 RNA promoter sequence. A region from "5' NTR" to "3' NTR" is DNA corresponding to the entire genomic region of hepatitis C virus.

[0067]

Next, the structural regions and a part of the non-structural regions of plasmid DNA pJFH1 and pJCH1 were substituted with a neomycin resistance gene (neo; also referred to as a neomycin phosphotransferase gene) and EMCV-IRES (internal ribosome entry site of encephalomyocarditis virus), thereby constructing plasmid DNA pSGREP-JFH1 and pSGREP-JCH1, respectively (lower section of Fig. 1). This construction procedure was conducted according to a previous report (Non Patent Literature 7). Specifically, plasmid pJFH1 and pJCH1 were cleaved with restriction enzymes Age I and Cla I, and between the Age I and Cla I restriction sites, the following fragments were inserted to be ligated; a fragment

was prepared by binding of a sequence ranging from 5' NTR to Core region derived from pJFH-1 with the neomycin resistance gene derived from pRSV5NEO by PCR amplification and then cleaving it with restriction enzymes Age I and Pme I, and, a fragment was prepared by binding of sequences ranging from EMCV IRES to NS3 region by PCR amplification and then cleaving it with restriction enzymes Pme I and Cla I.

[0068]

Moreover, a mutation that mutates an amino acid motif GDD to GND, corresponding to the active center of RNA polymerase encoded by the NS5B region, was introduced into the NS5B region in pSGREP-JFH1, thereby preparing a mutant plasmid clone pSGREP-JFH1/GND.

[0069]

Moreover, a mutation that results in the deletion of a sequence of 10 continuous amino acids containing an amino acid motif GDD corresponding to the active center of RNA polymerase encoded by the NS5B region was introduced into the NS5B region in pSGREP-JFH1, thereby preparing a mutant plasmid clone pSGREP-JFH1/dGDD.

[0070]

The above-prepared mutant clones pSGREP-JFH1/GND and pSGREP-JFH1/dGDD cannot express active NS5B protein, which is required for the replication of replicon RNA, because the amino acid sequence of the active site of NS5B protein encoded by these clones has mutated.

[0071]

(B) Preparation of replicon RNA

To prepare template DNA for use in synthesis of replicon RNA, the above-constructed expression vectors pSGREP-JFH1, pSGREP-JCH1, pSGREP-JFH1/GND and pSGREP-JFH1/dGDD were each cleaved with a restriction enzyme Xba I.

[0072]

Subsequently, 10 to 20 µg each of these Xba I-cleaved fragments was contained in 50 µl of a reaction solution, and then further treated by 30 minutes of incubation at 30°C with 20 U of Mung Bean Nuclease. Mung Bean Nuclease is an enzyme catalyzing a reaction for selectively degrading a single-stranded portion of double-stranded DNA. Generally, when RNA synthesis is performed using directly the above Xba I-cleaved fragment as a template, a replicon RNA having four nucleotides of CUGA, a part of the recognition sequence of Xba I, excessively added to the 3' terminus would be synthesized. Hence, in this example, Xba I-cleaved fragments were treated with Mung Bean Nuclease, so as to remove the four nucleotides of CUGA from the fragments. The solutions containing Xba I-cleaved fragments, which had been treated with Mung Bean Nuclease, were treated to remove proteins according to a general method, so that Xba I-cleaved fragments, from which the four nucleotides of CUGA had been removed, were purified and used as template DNAs.

[0073]

Next, from the template DNA, RNA was synthesized in vitro using T7 RNA polymerase. For this RNA synthesis, MEGAscript from Ambion, Inc. was used. Reaction was carried out using 20 µl of a reaction solution containing 0.5 to 1.0 micrograms of the template DNA according to the instructions of the manufacturer.

[0074]

After completion of RNA synthesis, DNase (2 U) was added to the reaction solution to conduct reaction at 37°C for 15 minutes. RNA extraction using acidic phenol was further performed to remove the template DNA. RNAs (replicon RNAs) synthesized in this manner from the above template DNAs derived from pSGREP-JFH1, pSGREP-JCH1, pSGREP-JFH1/GND and pSGREP-JFH1/dGDD were respectively named rSGREP-JFH1, rSGREP-JCH1, rSGREP-JFH1/GND and rSGREP-JFH1/dGDD. Regarding the nucleotide sequences of these replicon RNAs, the nucleotide sequence of rSGREP-JFH1 is shown in SEQ ID NO: 1 and

Fig. 2, that of rSGREP-JCH1 is shown in SEQ ID NO: 2 and Fig. 3, that of rSGREP-JFH1/GND is shown in SEQ ID NO: 7, and that of rSGREP-JFH1/dGDD is shown in SEQ ID NO: 8.

[0075]

[Example 2] Establishment of replicon-replicating cell clone

(C) Transfection of replicon RNA, determination of colony-forming ability of transfected cells and establishment of cell clones

Each of the above-synthesized replicon RNAs (rSGREP-JFH1, rSGREP-JCH1, rSGREP-JFH1/GND and rSGREP-JFH1/dGDD) was mixed in different quantities with total cellular RNA extracted from Huh7 cells so as to have a total RNA quantity of 10 μ g. Subsequently, the mixed RNA was introduced into Huh7 cells by the electroporation method. The Huh7 cells subjected to the electroporation treatment were seeded into culture dishes, and then cultured for 16 hours to 24 hours. G418 (neomycin) was then added to the culture dishes at different concentrations. Thereafter, culture was continued while exchanging the culture solutions twice a week. After 21 days of culture following seeding, surviving cells were stained with crystal violet. The number of stained colonies was counted, and then the number of colonies obtained per μ g of the transfected replicon RNA was calculated.

[0076]

For rSGREP-JFH1 or rSGREP-JCH1-transfected cells, for which colony formation had been observed, colonies of the surviving cells were further cloned from the above culture dishes after 21 days of culture, and were continuously cultured. By such cloning of colonies, several strains of cell clones could be established.

[0077]

For the established cell clones, detection of the replicated replicon RNA, confirmation of the presence or the absence of the incorporation of the neomycin resistance gene into the host genomic DNA, and confirmation of the expression of

HCV proteins were performed as described later, in Example 4. Cell clones for which the replication of the replicon had been confirmed in the cells were regarded as replicon-replicating cell clones.

[0078]

(D) Colony-forming ability in each transfected cell

As a result of the above transfection, for rSGREP-JFH1-transfected Huh7 cells, the colony-forming ability per μg of the transfected replicon RNA was 94700 CFU (Colony Forming Unit)/ μg -RNA when G418 concentration was 1.0 mg/ml (the left column in Fig. 4). In contrast, colony formation was not observed in the Huh7 cells, into which rSGREP-JFH1/dGDD and rSGREP-JFH1/GND had each been transfected (the central column and the right column in Fig. 4). This suggests that the colony-forming ability confirmed for the Huh7 cells, into which rSGREP-JFH1 replicon RNA had been transfected, depends on the activity of NS5B (RNA polymerase) expressed by rSGREP-JFH1. Specifically, it was considered that in cells that had formed colonies, rSGREP-JFH1 replicon RNA autonomously replicated due to the action of NS5B expressed by rSGREP-JFH1, and the neomycin resistance gene was continuously expressed to maintain G418 resistance, so that cell growth was enabled.

[0079]

On the other hand, in the Huh7 cells, into which rSGREP-JCH1 had been transfected, no colony formation was observed in the case of 1 to 0.5 mg/ml G418 concentrations (Fig. 5). When G418 concentration was lowered to 0.25 mg/ml, colony formation was observed in the Huh7 cells, into which rSGREP-JCH1 had been transfected as well.

[0080]

Furthermore, Xba I-cleaved fragment of the expression vector pSGREP-JFH1 obtained in (B) above was used as a template DNA for RNA synthesis without treating the fragment with Mung Bean Nuclease, so as to synthesize replicon RNA. This replicon RNA was transfected to Huh7 cells in a manner

similar to that in (C) above. The replicon RNA that had been prepared without performing Mung Bean Nuclease treatment had the four nucleotides of CUGA excessively added to the 3' terminus.

[0081]

As a result, the colony-forming ability of the Huh7 cells, into which the replicon RNA prepared without treatment with Mung Bean Nuclease had been transfected, decreased to 512 CFU/ μ g-RNA (the left side in Fig. 6). This result revealed that the sequence on the 3' terminus of the replicon RNA affects the colony-forming ability of the transfected cells.

[0082]

[Example 3]

(E) Re-transfection of replicated replicon RNA derived from replicon-replicating cells

From the replicon-replicating cell clones that had been established by transfection of rSGREP-JFH1 into Huh7 cells according to descriptions of Example 2, total RNA was extracted by a standard procedure. The number of copies of the replicated replicon RNA contained in the cellular RNA was determined by Northern blot analysis and a quantitative RT-PCR method.

[0083]

Northern blot analysis was performed according to the description in Molecular Cloning, A laboratory Manual, 2nd edition, J. Sambrook, E. F. Fritsch, T. Maniatis, Cold Spring Harbor Laboratory Press (1989). Specifically, RNA extracted from the cells was subjected to denaturing agarose electrophoresis. After electrophoresis, the RNA was transferred onto a positively charged nylon membrane. The ³²P-labeled DNA or RNA probe prepared from pSGREP-JFH1 was hybridized to the RNA transferred to the membrane as described above. Next the membrane was washed, and then exposed to a film, so as to detect a replicon-specific RNA band.

[0084]

Detection of the replicon RNA by quantitative RT-PCR was conducted by detecting the 5' untranslated region RNA within HCV RNA according to Takeuchi T, Katsume A, Tanaka T, Abe A, Inoue K, Tsukiyama-Kohara K, Kawaguchi R, Tanaka S and Kohara M., Real-time detection system for quantification of Hepatitis C virus genome, *Gastroenterology* 116: 636-642 (1999). Specifically, the replicon RNA contained in RNA extracted from the cells was amplified by PCR using synthetic primers: R6-130-S17, 5'-CGGGAGAGCCATAGTGG-3' (SEQ ID NO: 13) and R6-290-R19, 5'-AGTACCACAAGGCCTTTCG-3' (SEQ ID NO: 14); TaqMan Probe; R6-148-S21FT, 5'-CTGCGGAACCGGTGAGTACAC-3' (SEQ ID NO: 15) and an EZ rTth RNA PCR kit, and then detected using an ABI Prism 7700 sequence detector system.

[0085]

Next, aliquots of total cellular RNAs extracted from clone 6 (among the above-mentioned replicon-replicating cell clones) and pool clones (prepared by collecting replicon-replicating cells that had formed colonies from whole one dish and culturing them) were each introduced into another Huh7 cells by retransfection. Total cellular RNA used for the transfection was prepared to contain 1×10^7 copies of replicon RNA based on the number of copies of the above-determined replicon RNA. Transfection was performed as described in (C) above, and then selective culture was performed under G418 concentration conditions of 1 mg/ml. Thus, the colony formation of the replicon-replicating cells was observed (Fig. 7). The colony-forming ability in this case was 1 colony or more per 1×10^6 copies of the replicon RNA used for transfection, when it was calculated from the number of colonies obtained.

[0086]

On the other hand, the number of copies of in vitro synthetic RNA that had been synthesized in vitro using pSGREP-JFH1 as a template and T7 RNA polymerase was approximately 2×10^{11} copies/ μ g-RNA, when calculated based on the weight and the length of the RNA. The colony-forming ability in the case of

using the in vitro synthetic RNA for transfection in a manner similar to the above method was 1 colony per 5×10^7 copies. These results revealed that when RNA derived from cells extracted from replicon-replicating cells and in vitro synthetic RNA were each transfected to Huh7 cells as replicon RNA in the same number of copies, the use of the replicon RNA replicated within Huh7 cells resulted in colony-forming ability approximately 50 times higher than that of the in vitro synthetic RNA.

[0087]

[Example 4]

(F) Detection of replicon RNA

According to (E) above, cell clones [clones Nos. 1 to 11] were established by retransfection of total RNA that had been obtained from the replicon-replicating cell clone established by transfection of rSGREP-JFH1 to Huh7 cells to another Huh7 cells. From the established cell clones and pool clones (prepared by collecting cell clones that had formed colonies from whole one dish and then culturing them), respectively, total RNAs were extracted by an acidic phenol extraction method. Subsequently the total RNAs were analyzed by the Northern blot method using a pSGREP-JFH1-specific probe as a probe. As control, total RNA extracted similarly from untransfected Huh7 cells (in Fig. 8, denoted as "Huh7"), a sample prepared by adding 10^7 copies of replicon RNA synthesized in vitro to the total RNA extracted from Huh7 cells (in Fig. 8, denoted as " 10^7 "), and a sample (in Fig. 8, denoted as " 10^8 ") prepared by adding 10^8 copies of replicon RNA synthesized in vitro to the total RNA extracted from Huh7 cells, were used. In Fig. 8, 1 to 11 represent cell clone Numbers.

[0088]

As a result, RNA of approximately the same size as that of rSGREP-JFH1 was detected using a pSGREP-JFH1-specific probe (Fig. 8). Thus, it was confirmed that the replicon RNA from rSGREP-JFH1 that had been transfected at the beginning replicated and proliferated within the cell clones. In addition, it

was shown that the cell clones differed from each other in the quantity of the replicated replicon RNA. In Fig. 8, for example, clones 2, 6, 9 and 10 contained high quantities of the replicated replicon RNA, and clones 4, 8 and 11 contained low quantities of the replicated replicon RNA.

[0089]

(G) Confirmation of the presence or the absence of the incorporation of a neomycin resistance gene into genomic DNA

For the cell clones that had been obtained by retransfection of replicon RNA as described in Example 3, PCR amplification was performed using neomycin resistance gene-specific primers; sense primer, NEO-S3: 5'-AACAAAGATGGATTGCACGCA-3' (SEQ ID NO: 16) and antisense primer, NEO-R: 5'-CGTCAAGAAGGCGATAGAAG-3' (SEQ ID NO: 17), and the host cellular genomic DNA extracted from each of the cell clones as a template, in order to confirm that the resistance of each of the cell clones against G418 was not due to the incorporation of the neomycin resistance gene into the genome. The cell clones used herein were the cell clones Nos. 1 to 8 obtained by retransfection of rSGREP-JFH1-derived replicated replicon RNA (rSGREP-JFH1-derived cell clones Nos. 1 to 8), and cell clones Nos. 1 to 6 obtained by retransfection of rSGREP-JCH1-derived replicated replicon RNA (rSGREP-JCH1-derived cell clones Nos. 1 to 6). As a result, as shown in Fig. 9, in the eight examined rSGREP-JFH1-derived cell clones, positive clones showing the amplification of the neomycin resistance gene were not observed. For rSGREP-JCH1-derived cell clones, only 1 out of the 6 examined clones was positive (in Fig. 9, lane 3 in the right photograph). It was considered that this positive clone had acquired G418 resistance by the incorporation of the neomycin resistance gene in rSGREP-JCH1-derived replicated replicon RNA into the genomic DNA of the host cells. Thus, in the positive clone, unlike other clones, it was thought that the replicon RNA itself did not autonomously replicate within the cells. This was confirmed by the results of the experiment shown in the next (H) that no HCV proteins were

detected from the positive clone.

[0090]

(H) Detection of HCV protein

Protein was extracted from rSGREP-JFH1- and rSGREP-JCH1-transfected cell clones by a standard procedure, and then analyzed by SDS-PAGE and Western blot method (Fig. 10). The examined cell clones were the same as those used in (G) above: rSGREP-JFH1-derived cell clones Nos. 1 to 8 and rSGREP-JCH1-derived cell clones Nos. 1 to 6. In addition, a cellular extract from the cell obtained by transiently transfecting expression plasmid DNA containing NS3 gene into Huh7 cells was regarded as a positive control (NS3 protein). Furthermore, a protein extracted from the untransfected Huh7 cells was used as a negative control. A protein sample extracted from each cell clone was blotted onto a PVDF membrane (Immobilon-P, Millipore), and then detection of NS3 protein encoded by replicated replicon RNA was performed using anti-NS3-specific antibody (provided by Dr. Moradpour; Wolk B, et al, J. Virology. 2000, 74: 2293-2304). As shown in Fig. 10, in rSGREP-JFH1-derived cell clones Nos. 1 to 8 and rSGREP-JCH1-derived cell clones Nos. 1, 2 and 4 to 6, proteins of the same size as those of the positive control were detected. In rSGREP-JCH1-derived cell clone No. 3 (the clone detected as a positive clone in (G) above), no expression of NS3 protein was detected. That is, in rSGREP-JCH1-derived cell clone No. 3, no replication of replicon RNA was confirmed. NS3 protein was not detected in the untransfected Huh7 cells, revealing that in cell clones wherein NS3 protein was detected, the transfected replicon RNA autonomously replicated so that NS3 protein was expressed.

[0091]

Moreover, by the use of the serum of a hepatitis C patient as an antibody, the expression of NS5a protein from the replicon RNA was also confirmed in each cell clone for which the expression of NS3 protein had been confirmed as described above.

[0092]

Based on the results of (G) and (H) above, it was confirmed that replicon RNAs were replicated in the cell clones established by transfection of the replicon RNA.

[0093]

[Example 5]

(I) Analysis of adaptive mutation

According to descriptions of Example 3, total RNA obtained from the replicon-replicating cell clones established through the transfection of rSGREP-JFH1 into Huh7 cells was re-transfected to another Huh7 cells, thereby establishing 21 cell clones. Total RNA was extracted from each of these cell clones by a standard procedure. cDNA corresponding to the replicon RNA was synthesized using the total RNA as a template, reverse transcriptase Superscript II (Invitrogen) and primer 9641R-IH (5'-GCACTCTCTGCAGTCATGCGGCTCACGGAC-3' (SEQ ID NO: 18)). The composition of a reaction solution for the synthesis of cDNA by reverse transcription reaction is as shown below.

[0094]

<u>Composition of Reaction Solution</u>	<u>Fluid Volume (μl)</u>
5x 1st strand Buffer	4
2 mM dNTP	5
0.1 M DTT	1
9651R-IH primer (100 μM)	1
DW (distilled water)	6.5
Sample RNA (2 mg/mL)	1
RNasin (Promega) (40 U/μL)	0.5
<u>Superscript II RT (Invitrogen)</u>	<u>1</u>
Total	20 μl

[0095]

In cDNA synthesis reaction, the above reagents other than RNasin and Superscript II were mixed to prepare a first reaction solution. The solution was heated at 90°C for 3 minutes, and then cooled on ice. Subsequently, RNasin and Superscript II were added to this reaction solution, and then the solution was allowed to react at 42°C for 1 hour, followed by another reaction at 70°C for 15 minutes.

[0096]

Furthermore, PCR amplification was performed using the thus obtained cDNA together with five primer sets by the following procedures, so that DNA amplification fragments covering almost all the regions of the replicon RNA were obtained. The primer sets used and regions amplified by each primer set are shown in Table 1 and Table 2 below.

[0097]

[Table 1]

Designation of amplified fragment	Primer set		Amplified region
	Primer 1	Primer 2	
A/	42S-IH	433R-neo	41 - 470
B/	C/S17ssp	4680R-IH	28 - 3026
C/	4534S-IH	7279R-IH	2880 - 5625
D/	7198S-IH	9367R-IH	5544 - 7713
E/	9247S-NF	9576R-NF	7597 - 7960

In Table 1, an amplified region is represented by nucleotide numbers in rSGREP-JFH1 (SEQ ID NO: 1) that the region corresponds to.

[Table 2]

Primer designation	Nucleotide sequence (5'→3')	SEQ ID NO:
42S-IH	CCCCTGTGAGGAAGTACTGTCTTCACGC	SEQ ID NO: 19
C/S17ssp	CCGGGAGAGCCATAGTGGTCTGCG	SEQ ID NO: 20
4534S-IH	CCACTCAAAGAAAAAGTGTGACGAGCTCGC	SEQ ID NO: 21
7198S-IH	GGCTTGGGCACGGCCTGA	SEQ ID NO: 22
9247S-NF	GCGGTGAAGACCAAGCTCAAACCTCACTCCA	SEQ ID NO: 23

433R-neo	AGAACCTGCGTGCAATCCATC	SEQ ID NO: 24
4680R-IH	CCCGTCATGAGGGCGTCGGTGGC	SEQ ID NO: 25
7279R-IH	ACCAGCAACGGTGGGCGGTTGGTAATC	SEQ ID NO: 26
9367R-RI	GGCACGCGACACGCTGTG	SEQ ID NO: 27
9576R-NF	AGCTAGCCGTGACTAGGGCTAAGATGGAGC	SEQ ID NO: 28

[0098]

The composition of a reaction solution in this PCR reaction is as follows.

Composition of Reaction Solution	Fluid Volume (μ l)
Primer 1 (10 μ M)	1.0
Primer 2 (10 μ M)	1.0
2.5 mM dNTPs	5.0
10x LA Buffer	5.0
MgCl ₂ (25 mM)	5.0
LA Taq (TAKARA) (5 U/ μ l)	0.3
DW (distilled water)	30.7
Template cDNA	2.0
Total	50 μ l

[0099]

In addition, PCR reaction conditions are as follows: 95°C for 2 minutes; 35 cycles of 98°C for 10 seconds and then 68°C for 8 minutes; and 72°C for 7 minutes; after which the temperature is kept at 4°C.

[0100]

The nucleotide sequence of each PCR product obtained as described above was determined, and then the RNA sequence corresponding to the DNA sequence was compared with the sequence of rSGREP-JFH1. The results are shown in Table 3.

[0101]

[Table 3]

Region	Synonymous substitution	Nonsynonymous substitution	Total number of mutations
NS3	0	5	5
NS4A	0	2	2
NS4B	0	3	3
NS5A	0	7	7
NS5B	3	5	8
Total	3	22	25

[0102]

As shown in Table 3, total number of nucleotide mutations observed in 21 cell clones was 25. 22 of these mutations were nonsynonymous substitutions inducing amino acid mutation. Types of these mutations are as shown in Table 4. In addition, the positions of these mutations in the non-structural region are shown in Fig. 11.

[0103]

[Table 4]

Clone designation	Mutation site			
	Nucleotide No.	Nucleotide mutation	Amino acid mutation	Amino acid No.
C1	7098	A \Rightarrow G	None	
	7157	A \Rightarrow G	Y \Rightarrow C	2824
C2	4955	C \Rightarrow U	A \Rightarrow V	2090
C3	4936	A \Rightarrow G	T \Rightarrow A	2084
	5000	A \Rightarrow G	Y \Rightarrow C	2105
	7287	A \Rightarrow G	None	
	7288	A \Rightarrow G	M \Rightarrow V	2868
C4	5901	G \Rightarrow U	E \Rightarrow D	2405
	6113	A \Rightarrow U	H \Rightarrow L	2476
C5	2890	A \Rightarrow G	K \Rightarrow E	1402
C6	7209	A \Rightarrow G	None	

[0104]

In Table 4 and Fig. 11, "C1 to C6" represent replicon-replicating cell clones C1 to C6 having replicon RNA found to have mutations. "Nucleotide No." shows the corresponding nucleotide numbers within the nucleotide sequence of replicon RNA rSGREP-JFH1 (SEQ ID NO: 1). "Amino acid No." shows the corresponding amino acid numbers within the amino acid sequence encoded by the JFH-1 clone (SEQ ID NO: 4). The types of nucleotides and amino acids at mutation sites are described according to their general notations. As shown in Table 4, in clone C2, a nucleotide corresponding to nucleotide No. 4955 of SEQ ID NO: 1 on the replicon RNA mutated from C (cytosine) to U (uracil), which results in a mutation of an amino acid corresponding to amino acid No. 2090 of SEQ ID NO: 4 from A (alanine) to V (valine).

[0105]

Furthermore, mutation positions shown in Fig. 11 are shown with bar lines with the nucleotide numbers shown in Table 4. A thick bar line represents nonsynonymous substitution, and a thin bar line represents synonymous substitution.

[0106]

There were 2 clones having no nucleotide mutations at all that cause amino acid mutations. When Northern blot analysis was conducted for the 2 clones, it was shown that in these 2 clones, the quantity of replicon RNAs replicated was lower than those in the cell clones that had replicated replicon RNAs having a nucleotide mutation that causes an amino acid mutation. Hence, it was considered that the nucleotide mutation causing an amino acid mutation within the replicon RNA was an adaptive mutation for increasing the replication efficiency of the replicon RNA in Huh7 cells.

[0107]

[Example 6]

(J) Establishment of replicon-replicating cell clone using cells other than Huh7

cells

According to the method described in Example 1, rSGREP-JFH1 was transfected into some hepatic cancer cells other than Huh7 cells and non-liver-derived cells. The transfected cells were seeded into culture dishes and then cultured. Colony formation was observed and the number of colonies was counted. The cells used for transfection are as follows.

[0108]

- (1) HepG2 cells (representative hepatic cancer cells as well as Huh7 cells)
- (2) IMY-N9 cells (established by Ito et al; fusion cells of HepG2 cells and human primary culture hepatic cells (Hepatology 2001, 34: 566-572))
- (3) HeLa cells (human cervical cancer-derived cells (Can Cer Res. 1952, 12: 264-265))
- (4) 293 cells (human fetal kidney-derived cells (Gen. Virol. 1977, 36: 59-72))

[0109]

The results of transfection using HepG2 cells, IMY-N9 cells, HeLa cells or 293 cells, respectively, are shown in Fig. 12a to d. As shown in Fig. 12a to d, all HepG2 cells, IMY-N9 cells, HeLa cells, and 293 cells showed colony formation for rSGREP-JFH1-transfected cells.

[0110]

For the established cell clones, detection of the replicated replicon RNA, confirmation of the presence or the absence of the incorporation of the neomycin resistance gene into host genomic DNA, and confirmation of the expression of HCV protein were performed as described later, (L) and (M). The cell clones, for which the replication of the replicon in the cells had been confirmed, were regarded as replicon-replicating cell clones. Specifically, it was demonstrated that the use of rSGREP-JFH1 also enables the preparation of HCV replicon-replicating cells using hepatic cancer cells other than Huh7 cells and non-hepatic cells with which the production of HCV replicon-replicating cells had previously been unsuccessful (Blight et al., Science, (2000) 290; 1972-1974).

[0111]

(K) Detection of replicon RNA in replicon-replicating cells using cells other than Huh7 cells

Northern blot analysis was conducted according to a description of Molecular Cloning, A laboratory Manual, 2nd edition, J. Sambrook, E. F. Fritsch, T. Maniatis, Cold Spring Harbor Laboratory Press (1989). In accordance with the descriptions of the previous section (J), total RNA was extracted by the acidic phenol extraction method from each of the replicon-replicating cell clones that had been established by transfection of rSGREP-JFH1 into HepG2, IMY or HeLa cells respectively, and from pool clones of the replicon-replicating cells that had been established through transfection of rSGREP-JFH1 into 239 cells (prepared by collecting cell clones that had formed colonies from whole one dish and culturing them). Next, the total RNAs were analyzed by the Northern blot method using a pSGREP-JFH1-specific probe as a probe. As controls, total RNAs (lanes 1 and 17 in Fig. 13) extracted similarly from untransfected Huh7 cells and HepG2 cells, and RNA (lanes 2 and 3 in Fig. 13) prepared by adding 10^7 copies or 10^8 copies of the replicon RNA synthesized in vitro to total RNA extracted from Huh7 cells were used. As a result, RNA of approximately the same size of that of rSGREP-JFH1 was detected using a pSGREP-JFH1-specific probe (Fig. 13). Accordingly, it was confirmed that the replicon RNA derived from rSGREP-JFH1 that had been transfected at the beginning was replicated and proliferated within the cell clone. Furthermore, it was also revealed that the quantities of replicated replicon RNAs differed depending on cell type, and IMY cells were found to replicate the replicon RNA particularly efficiently. Moreover, it was revealed that the clones differed from each other in the quantity of the replicated replicon RNA.

[0112]

(L) Confirmation of the presence or the absence of the incorporation of the neomycin resistance gene into genomic DNA

For the thus established replicon RNA-replicating cell clone, PCR

amplification was performed using neomycin resistance gene-specific primers (sense primer, NEO-S3: 5'-AACAAGATGGATTGCACGCA-3' (SEQ ID NO: 29), antisense primer, NEO-R: 5'-CGTCAAGAAGGCGATAGAAG-3' (SEQ ID NO: 30)) and the host cellular genomic DNA extracted from each of the cell clones as a template, in order to confirm that the resistance of each of the cell clones against G418 was not due to the incorporation of the neomycin resistance gene into the genome. The cell clones used herein were the cell clones Nos. 1, 5, 7, 8, 9, 10, 11, 12 and 13 obtained by retransfection of rSGREP-JFH1-derived replicated replicon RNA into HepG2 cells, and the cell clones Nos. 3, 4, 5, 6, 7, 8, 9, 10 and 11 obtained by retransfection of rSGREP-JFH1-derived replicated replicon RNA into IMY N9 cells. As a result, as shown in Fig. 14, in the nine examined cell clones obtained by introduction of rSGREP-JFH1 into HepG2 cells, a positive clone showing the amplification of the neomycin resistance gene was not observed. In the 9 examined cell clones obtained by introduction of rSGREP-JFH1 into IMY N9 cells, a positive clone showing the amplification of the neomycin resistance gene was not observed.

[0113]

A similar examination was performed for cell clones obtained by retransfection of rSGREP-JFH1-derived replicated replicon RNA into HeLa cells, and cell clones obtained by retransfection of rSGREP-JFH1-derived replicated replicon RNA into 293 cells. Then, a positive clone showing the amplification of the neomycin resistance gene was not observed.

[0114]

(M) Detection of HCV protein

Proteins were extracted from the established cell clones by a standard procedure, and then analyzed by SDS-PAGE and the Western blot method (Fig. 15). The cell clones examined in this case were the same as those used in the above section: the cell clones Nos. 1, 5, 7, 8, 9, 10, 11, 12 and 13 obtained by retransfection of rSGREP-JFH1-derived replicated replicon RNA into HepG2 cells,

and the cell clones Nos. 3, 4, 5, 6, 7, 8, 9, 10 and 11 obtained by retransfection of rSGREP-JFH1-derived replicated replicon RNA into IMY-N9 cells. Furthermore, according to a previous report (Lehmann et. al., Science, (1999)), the HCV RNA replicon-replicating cell clone prepared by introducing rSGREP-JFH1 into HuH7 was regarded as a positive control (Fig. 15, lane 4-1, C6). Moreover, a protein extracted from untransfected cells was used as a negative control (Fig. 15, lane N). Protein samples extracted from each cell clone were blotted onto PVDF membranes (Immobilon-P, Millipore), and then detection of NS3 protein encoded by the replicated replicon RNA was performed using anti-NS3-specific antibody (provided by Dr. Moradpour; Wolk B, et al, J. Virology, 2000, 74: 2293-2304). As shown in the upper section in Fig. 15, a protein of the same size as that of the positive control was detected in the cell clones Nos. 1, 5, 7, 8, 9, 10, 11, 12 and 13 obtained by retransfection of rSGREP-JFH1-derived replicated replicon RNA, and in the cell clones Nos. 3, 4, 5, 6, 7, 8, 9, 10 and 11 obtained by retransfection of rSGREP-JFH1-derived replicated replicon RNA into IMY N9 cells.

[0115]

Moreover, by the use of the serum of a hepatitis C patient as an antibody, the confirmation of the expression of NS5a protein from the replicon RNA was performed for each cell clone that had been confirmed above to express NS3 protein. In this experiment, examination was performed in a manner similar to that in the case of the expression of NS3 protein, but using an antibody instead of the serum of the patient. As a result, as shown in the lower section in Fig. 15, a protein of the same size as that of the positive control was detected in the cell clones Nos. 1, 5, 7, 8, 9, 10, 11, 12 and 13 obtained by retransfection of rSGREP-JFH1-derived replicated replicon RNA, and the cell clones Nos. 3, 4, 5, 6, 7, 8, 9, 10 and 11 obtained by retransfection of rSGREP-JFH1-derived replicated replicon RNA into IMY N9 cells.

[0116]

When similar examination was performed for the cell clones obtained by

retransfection of rSGREP-JFH1-derived replicated replicon RNA into HeLa cells, and the cell clones obtained by retransfection of rSGREP-JFH1-derived replicated replicon RNA into 293 cells, the expression of NS3 and that of NS5a proteins could be confirmed.

[0117]

As described above, it was confirmed that the replicon RNA was replicated in the cell clones that had been established through transfection of the replicon RNA.

[0118]

[Example 7]

(N) Analysis of adaptive mutation

According to the descriptions of Example 3, total RNAs obtained from the replicon-replicating cell clones established through the transfection of rSGREP-JFH1 into HepG2 and HeLa cells were re-transfected into another cells of the each cell line, respectively, so that 14 cell clones were established for HepG2 cells and 8 cell clones were established for HeLa cells. From each of these cell clones, total RNA was extracted by a standard procedure. cDNA corresponding to the replicon RNA was synthesized using the total RNA as a template, reverse transcriptase Superscript II (Invitrogen) and a primer 9641R-IH (5'-GCACTCTCTGCAGTCATGCGGCTCACGGAC-3' (SEQ ID NO: 31)). The composition of a reaction solution for the synthesis of cDNA by reverse transcription reaction is as shown below.

[0119]

<u>Composition of Reaction Solution</u>	<u>Fluid Volume (μl)</u>
5x 1st strand Buffer	4
2 mM dNTP	5
0.1 M DTT	1
9651R-IH primer (100 μ M)	1

DW (distilled water)	6.5
Sample RNA (2 mg/mL)	1
RNAsin (Promega)(40 U/ μ L)	0.5
<u>Superscript II RT (Invitrogen)</u>	<u>1</u>
Total	20 μ l

[0120]

In cDNA synthesis reaction, the above reagents other than RNAsin and Superscript II were mixed to prepare a first reaction solution. The first reaction solution was heated at 90°C for 3 minutes, and then cooled on ice. Subsequently, RNAsin and Superscript II were added to the reaction solution, and then the solution was allowed to react at 42°C for 1 hour, followed by another reaction at 70°C for 15 minutes.

[0121]

Furthermore, PCR amplification was performed using the thus obtained cDNA together with five primer sets by the following procedures, so that DNA amplification fragments covering almost all the regions of replicon RNA were obtained. The primer sets used and regions amplified by each primer set are shown in Table 5 and Table 6 below.

[0122]

[Table 5]

Designation of amplified fragment	Primer set		Amplified region
	Primer 1	Primer 2	
A	42S-IH	433R-neo	41-470
B	C/S17ssp	4680R-IH	28-3026
C	4534S-IH	7279R-IH	2280-5625
D	7198S-IH	9367R-IH	5544-7713
E	9247S-NF	9576R-NF	7597-7966

In this table, an amplified region is represented by nucleotide numbers in

rSGREP-JFH1 (SEQ ID NO: 1) that the region corresponds to.

[0123]

[Table 6]

Primer	Nucleotide Sequence (5' to 3')	SEQ ID NO:
<u>Designation</u>		
43S-IH	CCCCTGTGAGGAAGTACTGTCTTCACGC	SEQ ID NO: 14
C/S17ssp	CCGGGAGAGCCATAGTGGTCTGCG	SEQ ID NO: 15
4534S-IH	CCACTCAAAGAAAAAGTGTGACGAGCTCGC	SEQ ID NO: 16
7198S-IH	GGCTTGGGCACGGCCTGA	SEQ ID NO: 17
9247S-NF	GCGGTGAAGACCAAGCTCAAAGTCACTCCA	SEQ ID NO: 18
433R-neo	AGAACCTGCGTGCAATCCATC	SEQ ID NO: 19
4680R-IH	CCCGTCATGAGGGCGTCCGGTGGC	SEQ ID NO: 20
7279R-IH	ACCAGCAACGGTGGGCGGTTGGTAATC	SEQ ID NO: 21
9367R-IH	GGAACGCGACACGCTGTG	SEQ ID NO: 22
9576R-NF	AGCTAGCCGTGACTAGGGCTAAGATGGAGC	SEQ ID NO: 23

[0124]

The composition of a reaction solution in this PCR reaction is as follows.

<u>Composition of Reaction Solution</u>	<u>Fluid Volume (μl)</u>
Primer 1 (10 μ M)	1.0
Primer 2 (10 μ M)	1.0
2.5 mM dNTPs	5.0
10x LA Buffer	5.0
MgCl ₂ (25 mM)	5.0
LA Taq (TAKARA) (5 U/ μ l)	0.3
DW (distilled water)	30.7
<u>Template cDNA</u>	<u>2.0</u>
Total	50 μ l

[0125]

In addition, PCR reaction conditions are as follows: 95°C for 2 minutes; 35 cycles of 98°C for 10 seconds and then 68°C for 8 minutes; followed by 72°C for 7 minutes, after which the temperature is kept at 4°C.

[0126]

The nucleotide sequence of each PCR product obtained as described above was determined, and then the RNA sequence corresponding to the DNA sequence was compared with the sequence of rSGREP-JFH1. The results are shown in Table 7 and Table 8.

[Table 7]

Analysis of adaptive mutation of JFH-1 replicon in HepG2 cells				
Clone	Mutation site		Mutation	
	Nucleotide No.	Amino acid No.	Nucleotide	Amino acid
HepIH1	6826	2714	C⇒A	Q⇒K
HepIH3	6887	2734	C⇒A	T⇒N
HepIH5	6887		U⇒A	None
HepIH8	6580	2632	U⇒A	S⇒T
	7159	2825	U⇒C	Y⇒H
HepIH9	3342		A⇒G	None
	3594		C⇒A	None
	7230	2848	U⇒A	N⇒K
HepIH10	5052		U⇒C	None
	6943	2753	C⇒A	P⇒T
HepIH12	None			
HepIH13	4302		C⇒U	None
	5687	2334	G⇒A	G⇒D
	6110	2475	A⇒G	Y⇒C

[0127]

As shown in Table 7, in the case of HepG2 cells, a total of 13 nucleotide mutations were observed in 8 cell clones, and 8 of these mutations were nonsynonymous substitutions that cause amino acid mutations. Types of these

mutations are shown in Table 8. On the other hand, in the case of HeLa cells, a total of 7 nucleotide mutations were observed in 3 cell clones, and 5 of these mutations were nonsynonymous substitutions that cause amino acid mutations. Types of these mutations are shown in Table 8.

[0128]

[Table 8]

Analysis of adaptive mutation of JFH-1 replicon in HeLa cells				
Clone	Mutation site		Mutation	
	Nucleotide No.	Amino acid No.	Nucleotide	Amino acid
HeLaH1	None			
HeLaIH2	5550	2272	U⇒C	S⇒P
	6252		A⇒G	None
	7182		U⇒C	None
	7217	2844	A⇒G	H⇒R
HeLaIH5	3643	1653	A⇒G	M⇒V
	5851	2389	G⇒A	A⇒T
	5914	2410	G⇒A	E⇒K

[0129]

In Tables 7 and 8, "HepIH No." represents clone numbers of replicon-replicating cell clones that have replicon RNA and have been cloned using HepG2 cells. "Nucleotide No." shows the corresponding nucleotide number in the nucleotide sequence (SEQ ID NO: 1) of replicon RNA rSGREP-JFH1. "Amino acid No." shows the corresponding amino acid number in the amino acid sequence (SEQ ID NO: 4) encoded by the JFH-1 clone. The types of nucleotides and amino acids at mutation sites are described according to their general notations. As shown in Table 7, for example, in clone HepIH1, a nucleotide corresponding to nucleotide No. 6826 of SEQ ID NO: on the replicon RNA mutated from C to A, so that an amino acid corresponding to amino acid No. 2714 of SEQ ID NO: mutated from Q to E. Similarly, in Table 8, "HeLaIH No." represents numbers of replicon-replicating cell clones that have replicon RNA and have been cloned

using HeLa cells.

[0130]

In addition, when Northern blot analysis was conducted for clones having no nucleotide mutations at all that cause amino acid mutations, it was shown that the quantity of replicon RNA replicated by the clones was lower than that of a cell clone replicating replicon RNA having a nucleotide mutation that causes an amino acid mutation. Hence, it was concluded that the nucleotide mutation in replicon RNA inducing an amino acid mutation was an adaptive mutation for increasing the replication efficiency of replicon RNA in cells.

[Industrial Applicability]

[0131]

The replicon-replicating cells according to the present invention can be utilized as a culture system for the continuous production of HCV genotype 2a-derived RNA and HCV protein. Moreover, the replicon-replicating cells according to the present invention are useful as a test system for screening for various substances affecting the replication of HCV and/or the translation into HCV protein.

[Brief Description of Drawings]

[0132]

[Fig. 1] Fig. 1 is a schematic view showing procedures for constructing a template DNA for preparing the HCV-RNA replicon according to the present invention. The upper section of Fig. 1 shows the structure of the region within pJFH1 or pJCH1, with the viral genome inserted into it. The lower section of Fig. 1 shows the structure of the region within plasmid DNA pSGREP-JFH1 or pSGREP-JCH1, with the viral genome inserted into it, that had been constructed by substituting a part of viral genome-inserted region of pJFH1 or pJCH1 with a DNA fragment containing a neomycin resistance gene and EMCV IRES. Symbols in Fig. 1 are as described below. T7, T7 RNA promoter; G, dGTP that was inserted upstream of the 5' end of the inserted DNA derived from JFH-1 or JCH-1

and downstream of the 3' end of T7 RNA promoter sequence; 5' NTR, 5' untranslated region; Core, core protein; and 3' NTR, 3' untranslated region. E1 and E2 represent envelope proteins. NS2, NS3, NS4A, NS4B, NS5A and NS5B represent non-structural proteins. Age I, Cla I and Xba I represent cleavage sites of restriction enzymes Age I, Cla I and Xba I, respectively. GDD, the position of amino acid motif GDD corresponding to the active center of NS5B protein; neo, neomycin resistance gene; and EMCV IRES, internal ribosome entry site of encephalomyocarditis virus (EMCV IRES).

[Fig. 2A] Fig. 2A shows the nucleotide sequence of rSGREP-JFH1.

[Fig. 2B] Fig. 2B shows the nucleotide sequence of rSGREP-JFH1.

[Fig. 2C] Fig. 2C shows the nucleotide sequence of rSGREP-JFH1.

[Fig. 2D] Fig. 2D shows the nucleotide sequence of rSGREP-JFH1.

[Fig. 2E] Fig. 2E shows the nucleotide sequence of rSGREP-JFH1.

[Fig. 2F] Fig. 2F shows the nucleotide sequence of rSGREP-JFH1.

[Fig. 3A] Fig. 3A shows the nucleotide sequence of rSGREP-JCH1.

[Fig. 3B] Fig. 3B shows the nucleotide sequence of rSGREP-JCH1.

[Fig. 3C] Fig. 3C shows the nucleotide sequence of rSGREP-JCH1.

[Fig. 3D] Fig. 3D shows the nucleotide sequence of rSGREP-JCH1.

[Fig. 3E] Fig. 3E shows the nucleotide sequence of rSGREP-JCH1.

[Fig. 3F] Fig. 3F shows the nucleotide sequence of rSGREP-JCH1.

[Fig. 4] Fig. 4 shows photographs showing the colony formation of Huh7 cells to which rSGREP-JFH1, rSGREP-JFH1/GND and rSGREP-JFH1/dGDD was transfected, respectively. The amount of each of three transfected RNAs in the upper section was 100 ng and that of three transfected RNAs in the lower section was 300 ng.

[Fig. 5] Fig. 5 shows photographs showing colony formation of Huh7 cells to which rSGREP-JFH1 and rSGREP-JCH1 respectively had been transfected when the concentration of G418 was 0.5 mg/ml of the medium. The amount of each of these RNAs transfected was 100 ng.

[Fig. 6] Fig. 6 shows photographs showing the effect of Mung Bean Nuclease treatment conducted on the colony-forming ability of the transfected cells. The amount of rSGREP-JFH1 RNA transfected was 100 ng for both cases. The concentration of G418 was 1.0 mg/ml in both media.

[Fig. 7] Fig. 7 shows photographs showing colony formation when total cellular RNA derived from the replicon-replicating cell clone, which had been established by transfection of rSGREP-JFH1, was retransfected to another Huh7 cells. The photograph on the left shows that the formation of 96 colonies was observed as a result, when using the total cellular RNA derived from the replicon-replicating cell clone No. 6. The photograph on the right shows that the formation of 77 colonies was observed as a result, when using the total cellular RNA derived from the pool clones. In both cases, RNA was retransfected in an amount containing 1×10^7 copies of the replicon RNA.

[Fig. 8] Fig. 8 shows photographs showing the results of detecting by the Northern blot method using an rSGREP-JFH1-specific probe for the total RNA derived from a cell clone that had been obtained by retransfecting the total cellular RNA (derived from the replicon-replicating cell clone established by transfection of rSGREP-JFH1) into another Huh7 cells. Explanation of the lanes is as follows. 10^8 represents sample prepared by adding 10^8 copies of the replicon RNA synthesized in vitro to total RNA extracted from Huh7 cells. 10^7 represents sample prepared by adding 10^7 copies of the replicon RNA synthesized in vitro to total RNA extracted from Huh7 cells. Huh7, total RNA extracted from untransfected Huh7 cells; pool clone, total RNA extracted from the pool clones; and 1-11, total RNA extracted from each of cell clones Nos. 1 to 11. "Replicon RNA" represents the electrophoresed position of a molecular weight marker indicating the size of rSGREP-JFH1, "28S" represents the same of a ribosomal RNA marker indicating the size of molecular weight of 4.5 kb, and "18S" represents the same of a ribosomal RNA marker indicating the size of molecular weight of 1.9 kb.

[Fig. 9] Fig. 9 shows photographs showing the presence or the absence of the incorporation of a neomycin resistance gene into the genomic DNA of a host cell in the cell clone to which rSGREP-JFH1- or rSGREP-JCH1-derived replicated replicon RNA was retransfected. Explanation of the lanes in the photograph on the left is as follows. M, DNA molecular weight marker; 1-8, rSGREP-JFH1-derived cell clones Nos. 1 to 8; N, untransfected Huh7 cells; and P, positive control (PCR amplification product of the neomycin resistance gene). Furthermore, explanation of the lanes in the photograph on the right is as follows. M, DNA molecular weight marker; and 1-6, rSGREP-JCH1-derived cell clones Nos. 1 to 6.

[Fig. 10] Fig. 10 shows photographs showing the results of detecting NS3 protein expressed in the cell clone that was retransfected with rSGREP-JFH1- or rSGREP-JCH1-derived replicated replicon RNA. Lanes 1 to 8 of the photograph on the left represent rSGREP-JFH1-derived cell clones Nos. 1 to 8. Lanes 1-6 of the photograph on the right represent rSGREP-JCH1-derived cell clones Nos. 1 to 6. Lane P of the photograph on the right represents NS3 protein (positive control) and N represents protein extracted from untransfected Huh7 cells (negative control).

[Fig. 11] Fig. 11 shows the positions of nucleotide mutations in replicon RNAs obtained from 21 cell clones that were established through the retransfection of rSGREP-JFH1-derived replicated replicon RNA into Huh7 cells. Mutation positions are indicated using bar lines shown with nucleotide numbers listed in Table 4. A thick bar line denotes nonsynonymous substitution and a thin bar line denotes synonymous substitution.

[Fig. 12] Fig. 12 shows photographs showing the results of transfection with rSGREP-JFH1 using 1) HepG2 cells; 2) IMY-N9 cells; 3) 293 cells; or 4) HeLa cells.

[Fig. 13] Fig. 13 shows photographs showing the results of performing Northern blotting for replicon-replicating cell clones.

[Fig. 14] Fig. 14 shows photographs showing the results of electrophoresis performed for confirming the incorporation of the neomycin resistance gene into genomic DNA.

[Fig. 15] Fig. 15 shows photographs showing the results of analyzing by the Western blot method proteins derived from the replicon-replicating cell clones.

[Sequence Listing Free Text]

[0133]

SEQ ID NO: 1. Explanation of artificial sequence: replicon

SEQ ID NO: 2. Explanation of artificial sequence: replicon

SEQ ID NO: 7. Explanation of artificial sequence: replicon

SEQ ID NOS: 8 to 12. Explanation of artificial sequences: synthetic RNA

SEQ ID NOS: 13 to 41. Explanation of artificial sequences: synthetic DNA

[Sequence Listing]

SEQUENCE LISTING

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Tokyo Metropolitan Organization for Medical Research

Johannes Gutenberg-Universitaet Mainz

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<223> Inventor: Wakita, Takaji

Inventor: Kato, Takanobu

Inventor: Date, Tomoko

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Arg Ala His Ala Leu Ile Arg Val Cys Ala Leu Val Lys Gln Leu Ala			
920	925	930	
ggg ggt agg tat gtt cag gtg gcg cta ttg gcc ctt ggc agg tgg act			3187
Gly Gly Arg Tyr Val Gln Val Ala Leu Leu Ala Leu Gly Arg Trp Thr			
935	940	945	
ggc acc tac atc tat gac cac ctc aca cct atg tcg gac tgg gcc gct			3235
Gly Thr Tyr Ile Tyr Asp His Leu Thr Pro Met Ser Asp Trp Ala Ala			
950	955	960	965
agc ggc ctg cgc gac tta gcg gtc gcc gtg gaa ccc atc atc ttc agt			3283
Ser Gly Leu Arg Asp Leu Ala Val Ala Val Glu Pro Ile Ile Phe Ser			
970	975	980	
ccg atg gag aag aag gtc atc gtc tgg gga gcg gag acg gct gca tgt			3331
Pro Met Glu Lys Lys Val Ile Val Trp Gly Ala Glu Thr Ala Ala Cys			
985	990	995	
ggg gac att cta cat gga ctt ccc gtg tcc gcc cga ctc ggc cag gag			3379
Gly Asp Ile Leu His Gly Leu Pro Val Ser Ala Arg Leu Gly Gln Glu			
1000	1005	1010	
atc ctc ctc ggc cca gct gat ggc tac acc tcc aag ggg tgg aag ctc			3427
Ile Leu Leu Gly Pro Ala Asp Gly Tyr Thr Ser Lys Gly Trp Lys Leu			

1015	1020	1025	
ctt gct ccc atc act gct tat gcc cag caa aca cga ggc ctc ctg ggc			3475
Leu Ala Pro Ile Thr Ala Tyr Ala Gln Gln Thr Arg Gly Leu Leu Gly			
1030	1035	1040	1045
gcc ata gtg gtg agt atg acg ggg cgt gac agg aca gaa cag gcc ggg			3523
Ala Ile Val Val Ser Met Thr Gly Arg Asp Arg Thr Glu Gln Ala Gly			
	1050	1055	1060
gaa gtc caa atc ctg tcc aca gtc tct cag tcc ttc ctc gga aca acc			3571
Glu Val Gln Ile Leu Ser Thr Val Ser Gln Ser Phe Leu Gly Thr Thr			
	1065	1070	1075
atc tcg ggg gtt ttg tgg act gtt tac cac gga gct ggc aac aag act			3619
Ile Ser Gly Val Leu Trp Thr Val Tyr His Gly Ala Gly Asn Lys Thr			
	1080	1085	1090
cta gcc ggc tta cgg ggt ccg gtc acg cag atg tac tcg agt gct gag			3667
Leu Ala Gly Leu Arg Gly Pro Val Thr Gln Met Tyr Ser Ser Ala Glu			
	1095	1100	1105
ggg gac ttg gta ggc tgg ccc agc ccc cct ggg acc aag tct ttg gag			3715
Gly Asp Leu Val Gly Trp Pro Ser Pro Pro Gly Thr Lys Ser Leu Glu			
	1110	1115	1120
ccg tgc aag tgt gga gcc gtc gac cta tat ctg gtc acg cgg aac gct			3763
Pro Cys Lys Cys Gly Ala Val Asp Leu Tyr Leu Val Thr Arg Asn Ala			
	1130	1135	1140
gat gtc atc ccg gct cgg aga cgc ggg gac aag cgg gga gca ttg ctc			3811
Asp Val Ile Pro Ala Arg Arg Arg Gly Asp Lys Arg Gly Ala Leu Leu			

1145	1150	1155	
tcc ccg aga ccc att tcg acc ttg aag ggg tcc tcg ggg ggg ccg gtg	3859		
Ser Pro Arg Pro Ile Ser Thr Leu Lys Gly Ser Ser Gly Gly Pro Val			
1160	1165	1170	
ctc tgc cct agg ggc cac gtc gtt ggg ctc ttc cga gca gct gtg tgc	3907		
Leu Cys Pro Arg Gly His Val Val Gly Leu Phe Arg Ala Ala Val Cys			
1175	1180	1185	
tct cgg ggc gtg gcc aaa tcc atc gat ttc atc ccc gtt gag aca ctc	3955		
Ser Arg Gly Val Ala Lys Ser Ile Asp Phe Ile Pro Val Glu Thr Leu			
1190	1195	1200	1205
gac gtt gtt aca agg tct ccc act ttc agt gac aac agc acg cca ccg	4003		
Asp Val Val Thr Arg Ser Pro Thr Phe Ser Asp Asn Ser Thr Pro Pro			
1210	1215	1220	
gct gtg ccc cag acc tat cag gtc ggg tac ttg cat gct cca act ggc	4051		
Ala Val Pro Gln Thr Tyr Gln Val Gly Tyr Leu His Ala Pro Thr Gly			
1225	1230	1235	
agt gga aag agc acc aag gtc cct gtc gcg tat gcc gcc cag ggg tac	4099		
Ser Gly Lys Ser Thr Lys Val Pro Val Ala Tyr Ala Ala Gln Gly Tyr			
1240	1245	1250	
aaa gta cta gtg ctt aac ccc tcg gta gct gcc acc ctg ggg ttt ggg	4147		
Lys Val Leu Val Leu Asn Pro Ser Val Ala Ala Thr Leu Gly Phe Gly			
1255	1260	1265	
gcg tac cta tcc aag gca cat ggc atc aat ccc aac att agg act gga	4195		
Ala Tyr Leu Ser Lys Ala His Gly Ile Asn Pro Asn Ile Arg Thr Gly			

1270	1275	1280	1285	
gtc agg acc gtg atg acc ggg gag gcc atc acg tac tcc aca tat ggc				4243
Val Arg Thr Val Met Thr Gly Glu Ala Ile Thr Tyr Ser Thr Tyr Gly				
	1290	1295	1300	
aaa ttt ctc gcc gat ggg ggc tgc gct agc ggc gcc tat gac atc atc				4291
Lys Phe Leu Ala Asp Gly Gly Cys Ala Ser Gly Ala Tyr Asp Ile Ile				
	1305	1310	1315	
ata tgc gat gaa tgc cac gct gtg gat gct acc tcc att ctc ggc atc				4339
Ile Cys Asp Glu Cys His Ala Val Asp Ala Thr Ser Ile Leu Gly Ile				
	1320	1325	1330	
gga acg gtc ctt gat caa gca gag aca gcc ggg gtc aga cta act gtg				4387
Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly Val Arg Leu Thr Val				
	1335	1340	1345	
ctg gct acg gcc aca ccc ccc ggg tca gtg aca acc ccc cat ccc gat				4435
Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr Thr Pro His Pro Asp				
1350	1355	1360	1365	
ata gaa gag gta ggc ctc ggg cgg gag ggt gag atc ccc ttc tat ggg				4483
Ile Glu Glu Val Gly Leu Gly Arg Glu Gly Glu Ile Pro Phe Tyr Gly				
	1370	1375	1380	
agg gcg att ccc cta tcc tgc atc aag gga ggg aga cac ctg att ttc				4531
Arg Ala Ile Pro Leu Ser Cys Ile Lys Gly Gly Arg His Leu Ile Phe				
	1385	1390	1395	
tgc cac tca aag aaa aag tgt gac gag ctc gcg gcg gcc ctt cgg ggc				4579
Cys His Ser Lys Lys Lys Cys Asp Glu Leu Ala Ala Ala Leu Arg Gly				

1400	1405	1410	
atg ggc ttg aat gcc gtg gca tac tat aga ggg ttg gac gtc tcc ata 4627			
Met Gly Leu Asn Ala Val Ala Tyr Tyr Arg Gly Leu Asp Val Ser Ile			
1415	1420	1425	
ata cca gct cag gga gat gtg gtg gtc gtc gcc acc gac gcc ctc atg 4675			
Ile Pro Ala Gln Gly Asp Val Val Val Val Ala Thr Asp Ala Leu Met			
1430	1435	1440	1445
acg ggg tac act gga gac ttt gac tcc gtg atc gac tgc aat gta gcg 4723			
Thr Gly Tyr Thr Gly Asp Phe Asp Ser Val Ile Asp Cys Asn Val Ala			
	1450	1455	1460
gtc acc caa gct gtc gac ttc agc ctg gac ccc acc ttc act ata acc 4771			
Val Thr Gln Ala Val Asp Phe Ser Leu Asp Pro Thr Phe Thr Ile Thr			
	1465	1470	1475
aca cag act gtc cca caa gac gct gtc tca cgc agt cag cgc cgc ggg 4819			
Thr Gln Thr Val Pro Gln Asp Ala Val Ser Arg Ser Gln Arg Arg Gly			
1480	1485	1490	
cgc aca ggt aga gga aga cag ggc act tat agg tat gtt tcc act ggt 4867			
Arg Thr Gly Arg Gly Arg Gln Gly Thr Tyr Arg Tyr Val Ser Thr Gly			
1495	1500	1505	
gaa cga gcc tca gga atg ttt gac agt gta gtg ctt tgt gag tgc tac 4915			
Glu Arg Ala Ser Gly Met Phe Asp Ser Val Val Leu Cys Glu Cys Tyr			
1510	1515	1520	1525
gac gca ggg gct gcg tgg tac gat ctc aca cca gcg gag acc acc gtc 4963			
Asp Ala Gly Ala Ala Trp Tyr Asp Leu Thr Pro Ala Glu Thr Thr Val			

1530	1535	1540	
agg ctt aga gcg tat ttc aac acg ccc ggc cta ccc gtg tgt caa gac			5011
Arg Leu Arg Ala Tyr Phe Asn Thr Pro Gly Leu Pro Val Cys Gln Asp			
1545	1550	1555	
cat ctt gaa ttt tgg gag gca gtt ttc acc ggc ctc aca cac ata gac			5059
His Leu Glu Phe Trp Glu Ala Val Phe Thr Gly Leu Thr His Ile Asp			
1560	1565	1570	
gcc cac ttc ctc tcc caa aca aag caa gcg ggg gag aac ttc gcg tac			5107
Ala His Phe Leu Ser Gln Thr Lys Gln Ala Gly Glu Asn Phe Ala Tyr			
1575	1580	1585	
cta gta gcc tac caa gct acg gtg tgc gcc aga gcc aag gcc cct ccc			5155
Leu Val Ala Tyr Gln Ala Thr Val Cys Ala Arg Ala Lys Ala Pro Pro			
1590	1595	1600	1605
ccg tcc tgg gac gcc atg tgg aag tgc ctg gcc cga ctc aag cct acg			5203
Pro Ser Trp Asp Ala Met Trp Lys Cys Leu Ala Arg Leu Lys Pro Thr			
1610	1615	1620	
ctt gcg ggc ccc aca cct ctc ctg tac cgt ttg ggc cct att acc aat			5251
Leu Ala Gly Pro Thr Pro Leu Leu Tyr Arg Leu Gly Pro Ile Thr Asn			
1625	1630	1635	
gag gtc acc ctc aca cac cct ggg acg aag tac atc gcc aca tgc atg			5299
Glu Val Thr Leu Thr His Pro Gly Thr Lys Tyr Ile Ala Thr Cys Met			
1640	1645	1650	
caa gct gac ctt gag gtc atg acc agc acg tgg gtc cta gct gga gga			5347
Gln Ala Asp Leu Glu Val Met Thr Ser Thr Trp Val Leu Ala Gly Gly			

1655	1660	1665	
gtc ctg gca gcc gtc gcc gca tat tgc ctg gcg act gga tgc gtt tcc 5395			
Val Leu Ala Ala Val Ala Ala Tyr Cys Leu Ala Thr Gly Cys Val Ser			
1670	1675	1680	1685
atc atc ggc cgc ttg cac gtc aac cag cga gtc gtc gtt gcg ccg gat 5443			
Ile Ile Gly Arg Leu His Val Asn Gln Arg Val Val Val Ala Pro Asp			
	1690	1695	1700
aag gag gtc ctg tat gag gct ttt gat gag atg gag gaa tgc gcc tct 5491			
Lys Glu Val Leu Tyr Glu Ala Phe Asp Glu Met Glu Glu Cys Ala Ser			
	1705	1710	1715
agg gcg gct ctc atc gaa gag ggg cag cgg ata gcc gag atg ttg aag 5539			
Arg Ala Ala Leu Ile Glu Glu Gly Gln Arg Ile Ala Glu Met Leu Lys			
	1720	1725	1730
tcc aag atc caa ggc ttg ctg cag cag gcc tct aag cag gcc cag gac 5587			
Ser Lys Ile Gln Gly Leu Leu Gln Gln Ala Ser Lys Gln Ala Gln Asp			
	1735	1740	1745
ata caa ccc gct atg cag gct tca tgg ccc aaa gtg gaa caa ttt tgg 5635			
Ile Gln Pro Ala Met Gln Ala Ser Trp Pro Lys Val Glu Gln Phe Trp			
1750	1755	1760	1765
gcc aga cac atg tgg aac ttc att agc ggc atc caa tac ctc gca gga 5683			
Ala Arg His Met Trp Asn Phe Ile Ser Gly Ile Gln Tyr Leu Ala Gly			
	1770	1775	1780
ttg tca aca ctg cca ggg aac ccc gcg gtg gct tcc atg atg gca ttc 5731			
Leu Ser Thr Leu Pro Gly Asn Pro Ala Val Ala Ser Met Met Ala Phe			

1785	1790	1795	
agt gcc gcc ctc acc agt ccg ttg tcg acc agt acc acc atc ctt ctc Ser Ala Ala Leu Thr Ser Pro Leu Ser Thr Ser Thr Thr Ile Leu Leu			5779
1800	1805	1810	
aac atc atg gga ggc tgg tta gcg tcc cag atc gca cca ccc gcg ggg Asn Ile Met Gly Gly Trp Leu Ala Ser Gln Ile Ala Pro Pro Ala Gly			5827
1815	1820	1825	
gcc acc ggc ttt gtc gtc agt ggc ctg gtg ggg gct gcc gtg ggc agc Ala Thr Gly Phe Val Val Ser Gly Leu Val Gly Ala Ala Val Gly Ser			5875
1830	1835	1840	1845
ata ggc ctg ggt aag gtg ctg gtg gac atc ctg gca gga tat ggt gcg Ile Gly Leu Gly Lys Val Leu Val Asp Ile Leu Ala Gly Tyr Gly Ala			5923
1850	1855	1860	
ggc att tcg ggg gcc ctc gtc gca ttc aag atc atg tct ggc gag aag Gly Ile Ser Gly Ala Leu Val Ala Phe Lys Ile Met Ser Gly Glu Lys			5971
1865	1870	1875	
ccc tct atg gaa gat gtc atc aat cta ctg cct ggg atc ctg tct ccg Pro Ser Met Glu Asp Val Ile Asn Leu Leu Pro Gly Ile Leu Ser Pro			6019
1880	1885	1890	
gga gcc ctg gtg gtg ggg gtc atc tgc gcg gcc att ctg cgc cgc cac Gly Ala Leu Val Val Gly Val Ile Cys Ala Ala Ile Leu Arg Arg His			6067
1895	1900	1905	
gtg gga ccg ggg gag ggc gcg gtc caa tgg atg aac agg ctt att gcc Val Gly Pro Gly Glu Gly Ala Val Gln Trp Met Asn Arg Leu Ile Ala			6115

1910	1915	1920	1925	
ttt gct tcc aga gga aac cac gtc gcc cct act cac tac gtg acg gag				6163
Phe Ala Ser Arg Gly Asn His Val Ala Pro Thr His Tyr Val Thr Glu				
	1930	1935	1940	
tcg gat gcg tcg cag cgt gtg acc caa cta ctt ggc tct ctt act ata				6211
Ser Asp Ala Ser Gln Arg Val Thr Gln Leu Leu Gly Ser Leu Thr Ile				
	1945	1950	1955	
acc agc cta ctc aga aga ctc cac aat tgg ata act gag gac tgc ccc				6259
Thr Ser Leu Leu Arg Arg Leu His Asn Trp Ile Thr Glu Asp Cys Pro				
	1960	1965	1970	
atc cca tgc tcc gga tcc tgg ctc cgc gac gtg tgg gac tgg gtt tgc				6307
Ile Pro Cys Ser Gly Ser Trp Leu Arg Asp Val Trp Asp Trp Val Cys				
	1975	1980	1985	
acc atc ttg aca gac ttc aaa aat tgg ctg acc tct aaa ttg ttc ccc				6355
Thr Ile Leu Thr Asp Phe Lys Asn Trp Leu Thr Ser Lys Leu Phe Pro				
1990	1995	2000	2005	
aag ctg ccc ggc ctc ccc ttc atc tct tgt caa aag ggg tac aag ggt				6403
Lys Leu Pro Gly Leu Pro Phe Ile Ser Cys Gln Lys Gly Tyr Lys Gly				
	2010	2015	2020	
gtg tgg gcc ggc act ggc atc atg acc acg cgc tgc cct tgc ggc gcc				6451
Val Trp Ala Gly Thr Gly Ile Met Thr Thr Arg Cys Pro Cys Gly Ala				
	2025	2030	2035	
aac atc tct ggc aat gtc cgc ctg ggc tct atg agg atc aca ggg cct				6499
Asn Ile Ser Gly Asn Val Arg Leu Gly Ser Met Arg Ile Thr Gly Pro				

2040	2045	2050	
aaa acc tgc atg aac acc tgg cag ggg acc ttt cct atc aat tgc tac			6547
Lys Thr Cys Met Asn Thr Trp Gln Gly Thr Phe Pro Ile Asn Cys Tyr			
2055	2060	2065	
acg gag ggc cag tgc gcg ccg aaa ccc ccc acg aac tac aag acc gcc			6595
Thr Glu Gly Gln Cys Ala Pro Lys Pro Pro Thr Asn Tyr Lys Thr Ala			
2070	2075	2080	2085
atc tgg agg gtg gcg gcc tcg gag tac gcg gag gtg acg cag cat ggg			6643
Ile Trp Arg Val Ala Ala Ser Glu Tyr Ala Glu Val Thr Gln His Gly			
2090	2095	2100	
tcg tac tcc tat gta aca gga ctg acc act gac aat ctg aaa att cct			6691
Ser Tyr Ser Tyr Val Thr Gly Leu Thr Thr Asp Asn Leu Lys Ile Pro			
2105	2110	2115	
tgc caa cta cct tct cca gag ttt ttc tcc tgg gtg gac ggt gtg cag			6739
Cys Gln Leu Pro Ser Pro Glu Phe Phe Ser Trp Val Asp Gly Val Gln			
2120	2125	2130	
atc cat agg ttt gca ccc aca cca aag ccg ttt ttc cgg gat gag gtc			6787
Ile His Arg Phe Ala Pro Thr Pro Lys Pro Phe Phe Arg Asp Glu Val			
2135	2140	2145	
tcg ttc tgc gtt ggg ctt aat tcc tat gct gtc ggg tcc cag ctt ccc			6835
Ser Phe Cys Val Gly Leu Asn Ser Tyr Ala Val Gly Ser Gln Leu Pro			
2150	2155	2160	2165
tgt gaa cct gag ccc gac gca gac gta ttg agg tcc atg cta aca gat			6883
Cys Glu Pro Glu Pro Asp Ala Asp Val Leu Arg Ser Met Leu Thr Asp			

2170	2175	2180	
ccg ccc cac atc acg gcg gag act gcg gcg cgg cgc ttg gca cgg gga	6931		
Pro Pro His Ile Thr Ala Glu Thr Ala Ala Arg Arg Leu Ala Arg Gly			
2185	2190	2195	
tca cct cca tct gag gcg agc tcc tca gtg agc cag cta tca gca ccg	6979		
Ser Pro Pro Ser Glu Ala Ser Ser Ser Val Ser Gln Leu Ser Ala Pro			
2200	2205	2210	
tcg ctg cgg gcc acc tgc acc acc cac agc aac acc tat gac gtg gac	7027		
Ser Leu Arg Ala Thr Cys Thr Thr His Ser Asn Thr Tyr Asp Val Asp			
2215	2220	2225	
atg gtc gat gcc aac ctg ctc atg gag ggc ggt gtg gct cag aca gag	7075		
Met Val Asp Ala Asn Leu Leu Met Glu Gly Gly Val Ala Gln Thr Glu			
2230	2235	2240	2245
cct gag tcc agg gtg ccc gtt ctg gac ttt ctc gag cca atg gcc gag	7123		
Pro Glu Ser Arg Val Pro Val Leu Asp Phe Leu Glu Pro Met Ala Glu			
2250	2255	2260	
gaa gag agc gac ctt gag ccc tca ata cca tcg gag tgc atg ctc ccc	7171		
Glu Glu Ser Asp Leu Glu Pro Ser Ile Pro Ser Glu Cys Met Leu Pro			
2265	2270	2275	
agg agc ggg ttt cca cgg gcc tta ccg gct tgg gca cgg cct gac tac	7219		
Arg Ser Gly Phe Pro Arg Ala Leu Pro Ala Trp Ala Arg Pro Asp Tyr			
2280	2285	2290	
aac ccg ccg ctc gtg gaa tcg tgg agg agg cca gat tac caa ccg ccc	7267		
Asn Pro Pro Leu Val Glu Ser Trp Arg Arg Pro Asp Tyr Gln Pro Pro			

2295	2300	2305	
acc gtt gct ggt tgt gct ctc ccc ccc ccc aag aag gcc ccg acg cct			7315
Thr Val Ala Gly Cys Ala Leu Pro Pro Pro Lys Lys Ala Pro Thr Pro			
2310	2315	2320	2325
ccc cca agg aga cgc cgg aca gtg ggt ctg agc gag agc acc ata tca			7363
Pro Pro Arg Arg Arg Arg Thr Val Gly Leu Ser Glu Ser Thr Ile Ser			
	2330	2335	2340
gaa gcc ctc cag caa ctg gcc atc aag acc ttt ggc cag ccc ccc tcg			7411
Glu Ala Leu Gln Gln Leu Ala Ile Lys Thr Phe Gly Gln Pro Pro Ser			
	2345	2350	2355
agc ggt gat gca ggc tcg tcc acg ggg gcg ggc gcc gcc gaa tcc ggc			7459
Ser Gly Asp Ala Gly Ser Ser Thr Gly Ala Gly Ala Ala Glu Ser Gly			
	2360	2365	2370
ggt ccg acg tcc cct ggt gag ccg gcc ccc tca gag aca ggt tcc gcc			7507
Gly Pro Thr Ser Pro Gly Glu Pro Ala Pro Ser Glu Thr Gly Ser Ala			
	2375	2380	2385
tcc tct atg ccc ccc ctc gag ggg gag cct gga gat ccg gac ctg gag			7555
Ser Ser Met Pro Pro Leu Glu Gly Glu Pro Gly Asp Pro Asp Leu Glu			
2390	2395	2400	2405
tct gat cag gta gag ctt caa cct ccc ccc cag ggg ggg ggg gta gct			7603
Ser Asp Gln Val Glu Leu Gln Pro Pro Pro Gln Gly Gly Gly Val Ala			
	2410	2415	2420
ccc ggt tcg ggc tcg ggg tct tgg tct act tgc tcc gag gag gac gat			7651
Pro Gly Ser Gly Ser Gly Ser Trp Ser Thr Cys Ser Glu Glu Asp Asp			

2425	2430	2435	
acc acc gtg tgc tgc tcc atg tca tac tcc tgg acc ggg gct cta ata			7699
Thr Thr Val Cys Cys Ser Met Ser Tyr Ser Trp Thr Gly Ala Leu Ile			
2440	2445	2450	
act ccc tgt agc ccc gaa gag gaa aag ttg cca atc aac cct ttg agt			7747
Thr Pro Cys Ser Pro Glu Glu Glu Lys Leu Pro Ile Asn Pro Leu Ser			
2455	2460	2465	
aac tcg ctg ttg cga tac cat aac aag gtg tac tgt aca aca tca aag			7795
Asn Ser Leu Leu Arg Tyr His Asn Lys Val Tyr Cys Thr Thr Ser Lys			
2470	2475	2480	2485
agc gcc tca cag agg gct aaa aag gta act ttt gac agg acg caa gtg			7843
Ser Ala Ser Gln Arg Ala Lys Lys Val Thr Phe Asp Arg Thr Gln Val			
2490	2495	2500	
ctc gac gcc cat tat gac tca gtc tta aag gac atc aag cta gcg gct			7891
Leu Asp Ala His Tyr Asp Ser Val Leu Lys Asp Ile Lys Leu Ala Ala			
2505	2510	2515	
tcc aag gtc agc gca agg ctc ctc acc ttg gag gag gcg tgc cag ttg			7939
Ser Lys Val Ser Ala Arg Leu Leu Thr Leu Glu Glu Ala Cys Gln Leu			
2520	2525	2530	
act cca ccc cat tct gca aga tcc aag tat gga ttc ggg gcc aag gag			7987
Thr Pro Pro His Ser Ala Arg Ser Lys Tyr Gly Phe Gly Ala Lys Glu			
2535	2540	2545	
gtc cgc agc ttg tcc ggg agg gcc gtt aac cac atc aag tcc gtg tgg			8035
Val Arg Ser Leu Ser Gly Arg Ala Val Asn His Ile Lys Ser Val Trp			

2550	2555	2560	2565	
aag gac ctc ctg gaa gac cca caa aca cca att ccc aca acc atc atg				8083
Lys Asp Leu Leu Glu Asp Pro Gln Thr Pro Ile Pro Thr Thr Ile Met				
	2570	2575	2580	
gcc aaa aat gag gtg ttc tgc gtg gac ccc gcc aag ggg ggt aag aaa				8131
Ala Lys Asn Glu Val Phe Cys Val Asp Pro Ala Lys Gly Gly Lys Lys				
	2585	2590	2595	
cca gct cgc ctc atc gtt tac cct gac ctc ggc gtc cgg gtc tgc gag				8179
Pro Ala Arg Leu Ile Val Tyr Pro Asp Leu Gly Val Arg Val Cys Glu				
	2600	2605	2610	
aaa atg gcc ctc tat gac att aca caa aag ctt cct cag gcg gta atg				8227
Lys Met Ala Leu Tyr Asp Ile Thr Gln Lys Leu Pro Gln Ala Val Met				
	2615	2620	2625	
gga gct tcc tat ggc ttc cag tac tcc cct gcc caa cgg gtg gag tat				8275
Gly Ala Ser Tyr Gly Phe Gln Tyr Ser Pro Ala Gln Arg Val Glu Tyr				
	2630	2635	2640	2645
ctc ttg aaa gca tgg gcg gaa aag aag gac ccc atg ggt ttt tcg tat				8323
Leu Leu Lys Ala Trp Ala Glu Lys Lys Asp Pro Met Gly Phe Ser Tyr				
	2650	2655	2660	
gat acc cga tgc ttc gac tca acc gtc act gag aga gac atc agg acc				8371
Asp Thr Arg Cys Phe Asp Ser Thr Val Thr Glu Arg Asp Ile Arg Thr				
	2665	2670	2675	
gag gag tcc ata tac cag gcc tgc tcc ctg ccc gag gag gcc cgc act				8419
Glu Glu Ser Ile Tyr Gln Ala Cys Ser Leu Pro Glu Glu Ala Arg Thr				

2680	2685	2690	
gcc ata cac tcg ctg act gag aga ctt tac gta gga ggg ccc atg ttc			8467
Ala Ile His Ser Leu Thr Glu Arg Leu Tyr Val Gly Gly Pro Met Phe			
2695	2700	2705	
aac agc aag ggt caa acc tgc ggt tac aga cgt tgc cgc gcc agc ggg			8515
Asn Ser Lys Gly Gln Thr Cys Gly Tyr Arg Arg Cys Arg Ala Ser Gly			
2710	2715	2720	2725
gtg cta acc act agc atg ggt aac acc atc aca tgc tat gtg aaa gcc			8563
Val Leu Thr Thr Ser Met Gly Asn Thr Ile Thr Cys Tyr Val Lys Ala			
2730	2735	2740	
cta gcg gcc tgc aag gct gcg ggg ata gtt gcg ccc aca atg ctg gta			8611
Leu Ala Ala Cys Lys Ala Ala Gly Ile Val Ala Pro Thr Met Leu Val			
2745	2750	2755	
tgc ggc gat gac cta gta gtc atc tca gaa agc cag ggg act gag gag			8659
Cys Gly Asp Asp Leu Val Val Ile Ser Glu Ser Gln Gly Thr Glu Glu			
2760	2765	2770	
gac gag cgg aac ctg aga gcc ttc acg gag gcc atg acc agg tac tct			8707
Asp Glu Arg Asn Leu Arg Ala Phe Thr Glu Ala Met Thr Arg Tyr Ser			
2775	2780	2785	
gcc cct cct ggt gat ccc ccc aga ccg gaa tat gac ctg gag cta ata			8755
Ala Pro Pro Gly Asp Pro Pro Arg Pro Glu Tyr Asp Leu Glu Leu Ile			
2790	2795	2800	2805
aca tcc tgt tcc tca aat gtg tct gtg gcg ttg ggc ccg cgg ggc cgc			8803
Thr Ser Cys Ser Ser Asn Val Ser Val Ala Leu Gly Pro Arg Gly Arg			

2810	2815	2820	
cgc aga tac tac ctg acc aga gac cca acc act cca ctc gcc cgg gct	8851		
Arg Arg Tyr Tyr Leu Thr Arg Asp Pro Thr Thr Pro Leu Ala Arg Ala			
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gcc tgg gaa aca gtt aga cac tcc cct atc aat tca tgg ctg gga aac	8899		
Ala Trp Glu Thr Val Arg His Ser Pro Ile Asn Ser Trp Leu Gly Asn			
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atc atc cag tat gct cca acc ata tgg gtt cgc atg gtc cta atg aca	8947		
Ile Ile Gln Tyr Ala Pro Thr Ile Trp Val Arg Met Val Leu Met Thr			
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cac ttc ttc tcc att ctc atg gtc caa gac acc ctg gac cag aac ctc	8995		
His Phe Phe Ser Ile Leu Met Val Gln Asp Thr Leu Asp Gln Asn Leu			
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Asn Phe Glu Met Tyr Gly Ser Val Tyr Ser Val Asn Pro Leu Asp Leu			
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cca gcc ata att gag agg tta cac ggg ctt gac gcc ttt tct atg cac	9091		
Pro Ala Ile Ile Glu Arg Leu His Gly Leu Asp Ala Phe Ser Met His			
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Thr Tyr Ser His His Glu Leu Thr Arg Val Ala Ser Ala Leu Arg Lys			
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ctt ggg gcg cca ccc ctc agg gtg tgg aag agt cgg gct cgc gca gtc	9187		
Leu Gly Ala Pro Pro Leu Arg Val Trp Lys Ser Arg Ala Arg Ala Val			

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 agg gcg tcc ctc atc tcc cgt gga ggg aaa gcg gcc gtt tgc ggc cga 9235
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 Tyr Leu Phe Asn Trp Ala Val Lys Thr Lys Leu Lys Leu Thr Pro Leu
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 Gly Gly Gly Asp Ile Phe His Ser Val Ser Arg Ala Arg Pro Arg Ser
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 tta ctc ttc ggc cta ctc cta ctt ttc gta ggg gta ggc ctc ttc cta 9427
 Leu Leu Phe Gly Leu Leu Leu Leu Phe Val Gly Val Gly Leu Phe Leu
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 ctc ccc gct cgg tag agcggcacac actaggtaca ctccatagct aactgttct 9482
 Leu Pro Ala Arg
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 tctttcttcc ctctcatct tattctactt tctttcttgg tggctccatc tttagccctag 9602
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9678

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<211> 3033

<212> PRT

<213> Hepatitis C virus

<400> 4

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Gly Val Tyr Leu Leu Pro Arg Arg Gly Pro Arg Leu Gly Val Arg Thr

35 40 45

Thr Arg Lys Thr Ser Glu Arg Ser Gln Pro Arg Gly Arg Arg Gln Pro

50 55 60

Ile Pro Lys Asp Arg Arg Ser Thr Gly Lys Ala Trp Gly Lys Pro Gly

65 70 75 80

Arg Pro Trp Pro Leu Tyr Gly Asn Glu Gly Leu Gly Trp Ala Gly Trp

85 90 95

Leu Leu Ser Pro Arg Gly Ser Arg Pro Ser Trp Gly Pro Thr Asp Pro

100 105 110

Arg His Arg Ser Arg Asn Val Gly Lys Val Ile Asp Thr Leu Thr Cys

115 120 125

Gly Phe Ala Asp Leu Met Gly Tyr Ile Pro Val Val Gly Ala Pro Leu

130 135 140

Ser Gly Ala Ala Arg Ala Val Ala His Gly Val Arg Val Leu Glu Asp

145 150 155 160

Gly Val Asn Tyr Ala Thr Gly Asn Leu Pro Gly Phe Pro Phe Ser Ile

165 170 175

Phe Leu Leu Ala Leu Leu Ser Cys Ile Thr Val Pro Val Ser Ala Ala

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Gln Val Lys Asn Thr Ser Ser Ser Tyr Met Val Thr Asn Asp Cys Ser			
195	200	205	
Asn Asp Ser Ile Thr Trp Gln Leu Glu Ala Ala Val Leu His Val Pro			
210	215	220	
Gly Cys Val Pro Cys Glu Arg Val Gly Asn Thr Ser Arg Cys Trp Val			
225	230	235	240
Pro Val Ser Pro Asn Met Ala Val Arg Gln Pro Gly Ala Leu Thr Gln			
245	250	255	
Gly Leu Arg Thr His Ile Asp Met Val Val Met Ser Ala Thr Phe Cys			
260	265	270	
Ser Ala Leu Tyr Val Gly Asp Leu Cys Gly Gly Val Met Leu Ala Ala			
275	280	285	
Gln Val Phe Ile Val Ser Pro Gln Tyr His Trp Phe Val Gln Glu Cys			
290	295	300	
Asn Cys Ser Ile Tyr Pro Gly Thr Ile Thr Gly His Arg Met Ala Trp			
305	310	315	320
Asp Met Met Met Asn Trp Ser Pro Thr Ala Thr Met Ile Leu Ala Tyr			
325	330	335	
Val Met Arg Val Pro Glu Val Ile Ile Asp Ile Val Ser Gly Ala His			
340	345	350	
Trp Gly Val Met Phe Gly Leu Ala Tyr Phe Ser Met Gln Gly Ala Trp			
355	360	365	
Ala Lys Val Ile Val Ile Leu Leu Leu Ala Ala Gly Val Asp Ala Gly			
370	375	380	
Thr Thr Thr Val Gly Gly Ala Val Ala Arg Ser Thr Asn Val Ile Ala			
385	390	395	400
Gly Val Phe Ser His Gly Pro Gln Gln Asn Ile Gln Leu Ile Asn Thr			
405	410	415	
Asn Gly Ser Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser			
420	425	430	
Leu Asn Thr Gly Phe Leu Ala Ala Leu Phe Tyr Thr Asn Arg Phe Asn			

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 Ser Ser Gly Cys Pro Gly Arg Leu Ser Ala Cys Arg Asn Ile Glu Ala
 450 455 460
 Phe Arg Ile Gly Trp Gly Thr Leu Gln Tyr Glu Asp Asn Val Thr Asn
 465 470 475 480
 Pro Glu Asp Met Arg Pro Tyr Cys Trp His Tyr Pro Pro Lys Pro Cys
 485 490 495
 Gly Val Val Pro Ala Arg Ser Val Cys Gly Pro Val Tyr Cys Phe Thr
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 Pro Ser Pro Val Val Val Gly Thr Thr Asp Arg Arg Gly Val Pro Thr
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 Tyr Thr Trp Gly Glu Asn Glu Thr Asp Val Phe Leu Leu Asn Ser Thr
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 Arg Pro Pro Gln Gly Ser Trp Phe Gly Cys Thr Trp Met Asn Ser Thr
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 Gly Phe Thr Lys Thr Cys Gly Ala Pro Pro Cys Arg Thr Arg Ala Asp
 565 570 575
 Phe Asn Ala Ser Thr Asp Leu Leu Cys Pro Thr Asp Cys Phe Arg Lys
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 His Pro Asp Ala Thr Tyr Ile Lys Cys Gly Ser Gly Pro Trp Leu Thr
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 Pro Lys Cys Leu Val His Tyr Pro Tyr Arg Leu Trp His Tyr Pro Cys
 610 615 620
 Thr Val Asn Phe Thr Ile Phe Lys Ile Arg Met Tyr Val Gly Gly Val
 625 630 635 640
 Glu His Arg Leu Thr Ala Ala Cys Asn Phe Thr Arg Gly Asp Arg Cys
 645 650 655
 Asp Leu Glu Asp Arg Asp Arg Ser Gln Leu Ser Pro Leu Leu His Ser
 660 665 670
 Thr Thr Glu Trp Ala Ile Leu Pro Cys Thr Tyr Ser Asp Leu Pro Ala
 675 680 685
 Leu Ser Thr Gly Leu Leu His Leu His Gln Asn Ile Val Asp Val Gln

690 695 700
 Tyr Met Tyr Gly Leu Ser Pro Ala Ile Thr Lys Tyr Val Val Arg Trp
 705 710 715 720
 Glu Trp Val Val Leu Leu Phe Leu Leu Leu Ala Asp Ala Arg Val Cys
 725 730 735
 Ala Cys Leu Trp Met Leu Ile Leu Leu Gly Gln Ala Glu Ala Ala Leu
 740 745 750
 Glu Lys Leu Val Val Leu His Ala Ala Ser Ala Ala Asn Cys His Gly
 755 760 765
 Leu Leu Tyr Phe Ala Ile Phe Phe Val Ala Ala Trp His Ile Arg Gly
 770 775 780
 Arg Val Val Pro Leu Thr Thr Tyr Cys Leu Thr Gly Leu Trp Pro Phe
 785 790 795 800
 Cys Leu Leu Leu Met Ala Leu Pro Arg Gln Ala Tyr Ala Tyr Asp Ala
 805 810 815
 Pro Val His Gly Gln Ile Gly Val Gly Leu Leu Ile Leu Ile Thr Leu
 820 825 830
 Phe Thr Leu Thr Pro Gly Tyr Lys Thr Leu Leu Gly Gln Cys Leu Trp
 835 840 845
 Trp Leu Cys Tyr Leu Leu Thr Leu Gly Glu Ala Met Ile Gln Glu Trp
 850 855 860
 Val Pro Pro Met Gln Val Arg Gly Gly Arg Asp Gly Ile Ala Trp Ala
 865 870 875 880
 Val Thr Ile Phe Cys Pro Gly Val Val Phe Asp Ile Thr Lys Trp Leu
 885 890 895
 Leu Ala Leu Leu Gly Pro Ala Tyr Leu Leu Arg Ala Ala Leu Thr His
 900 905 910
 Val Pro Tyr Phe Val Arg Ala His Ala Leu Ile Arg Val Cys Ala Leu
 915 920 925
 Val Lys Gln Leu Ala Gly Gly Arg Tyr Val Gln Val Ala Leu Leu Ala
 930 935 940
 Leu Gly Arg Trp Thr Gly Thr Tyr Ile Tyr Asp His Leu Thr Pro Met

945 950 955 960
 Ser Asp Trp Ala Ala Ser Gly Leu Arg Asp Leu Ala Val Ala Val Glu
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 Pro Ile Ile Phe Ser Pro Met Glu Lys Lys Val Ile Val Trp Gly Ala
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 Glu Thr Ala Ala Cys Gly Asp Ile Leu His Gly Leu Pro Val Ser Ala
 995 1000 1005
 Arg Leu Gly Gln Glu Ile Leu Leu Gly Pro Ala Asp Gly Tyr Thr Ser
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 Lys Gly Trp Lys Leu Leu Ala Pro Ile Thr Ala Tyr Ala Gln Gln Thr
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 1045 1050 1055
 Thr Glu Gln Ala Gly Glu Val Gln Ile Leu Ser Thr Val Ser Gln Ser
 1060 1065 1070
 Phe Leu Gly Thr Thr Ile Ser Gly Val Leu Trp Thr Val Tyr His Gly
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 Ala Gly Asn Lys Thr Leu Ala Gly Leu Arg Gly Pro Val Thr Gln Met
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 Tyr Ser Ser Ala Glu Gly Asp Leu Val Gly Trp Pro Ser Pro Pro Gly
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 Thr Lys Ser Leu Glu Pro Cys Lys Cys Gly Ala Val Asp Leu Tyr Leu
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 Val Thr Arg Asn Ala Asp Val Ile Pro Ala Arg Arg Arg Gly Asp Lys
 1140 1145 1150
 Arg Gly Ala Leu Leu Ser Pro Arg Pro Ile Ser Thr Leu Lys Gly Ser
 1155 1160 1165
 Ser Gly Gly Pro Val Leu Cys Pro Arg Gly His Val Val Gly Leu Phe
 1170 1175 1180
 Arg Ala Ala Val Cys Ser Arg Gly Val Ala Lys Ser Ile Asp Phe Ile
 1185 1190 1195 1200
 Pro Val Glu Thr Leu Asp Val Val Thr Arg Ser Pro Thr Phe Ser Asp

1205	1210	1215	
Asn Ser Thr Pro Pro Ala Val Pro Gln Thr Tyr Gln Val Gly Tyr Leu			
1220	1225	1230	
His Ala Pro Thr Gly Ser Gly Lys Ser Thr Lys Val Pro Val Ala Tyr			
1235	1240	1245	
Ala Ala Gln Gly Tyr Lys Val Leu Val Leu Asn Pro Ser Val Ala Ala			
1250	1255	1260	
Thr Leu Gly Phe Gly Ala Tyr Leu Ser Lys Ala His Gly Ile Asn Pro			
1265	1270	1275	1280
Asn Ile Arg Thr Gly Val Arg Thr Val Met Thr Gly Glu Ala Ile Thr			
1285	1290	1295	
Tyr Ser Thr Tyr Gly Lys Phe Leu Ala Asp Gly Gly Cys Ala Ser Gly			
1300	1305	1310	
Ala Tyr Asp Ile Ile Ile Cys Asp Glu Cys His Ala Val Asp Ala Thr			
1315	1320	1325	
Ser Ile Leu Gly Ile Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly			
1330	1335	1340	
Val Arg Leu Thr Val Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr			
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Thr Pro His Pro Asp Ile Glu Glu Val Gly Leu Gly Arg Glu Gly Glu			
1365	1370	1375	
Ile Pro Phe Tyr Gly Arg Ala Ile Pro Leu Ser Cys Ile Lys Gly Gly			
1380	1385	1390	
Arg His Leu Ile Phe Cys His Ser Lys Lys Lys Cys Asp Glu Leu Ala			
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Ala Ala Leu Arg Gly Met Gly Leu Asn Ala Val Ala Tyr Tyr Arg Gly			
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Leu Asp Val Ser Ile Ile Pro Ala Gln Gly Asp Val Val Val Val Ala			
1425	1430	1435	1440
Thr Asp Ala Leu Met Thr Gly Tyr Thr Gly Asp Phe Asp Ser Val Ile			
1445	1450	1455	
Asp Cys Asn Val Ala Val Thr Gln Ala Val Asp Phe Ser Leu Asp Pro			

1460	1465	1470	
Thr Phe Thr Ile Thr Thr Gln Thr Val Pro Gln Asp Ala Val Ser Arg			
1475	1480	1485	
Ser Gln Arg Arg Gly Arg Thr Gly Arg Gly Arg Gln Gly Thr Tyr Arg			
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Tyr Val Ser Thr Gly Glu Arg Ala Ser Gly Met Phe Asp Ser Val Val			
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Leu Cys Glu Cys Tyr Asp Ala Gly Ala Ala Trp Tyr Asp Leu Thr Pro			
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Ala Glu Thr Thr Val Arg Leu Arg Ala Tyr Phe Asn Thr Pro Gly Leu			
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Pro Val Cys Gln Asp His Leu Glu Phe Trp Glu Ala Val Phe Thr Gly			
1555	1560	1565	
Leu Thr His Ile Asp Ala His Phe Leu Ser Gln Thr Lys Gln Ala Gly			
1570	1575	1580	
Glu Asn Phe Ala Tyr Leu Val Ala Tyr Gln Ala Thr Val Cys Ala Arg			
1585	1590	1595	1600
Ala Lys Ala Pro Pro Pro Ser Trp Asp Ala Met Trp Lys Cys Leu Ala			
1605	1610	1615	
Arg Leu Lys Pro Thr Leu Ala Gly Pro Thr Pro Leu Leu Tyr Arg Leu			
1620	1625	1630	
Gly Pro Ile Thr Asn Glu Val Thr Leu Thr His Pro Gly Thr Lys Tyr			
1635	1640	1645	
Ile Ala Thr Cys Met Gln Ala Asp Leu Glu Val Met Thr Ser Thr Trp			
1650	1655	1660	
Val Leu Ala Gly Gly Val Leu Ala Ala Val Ala Ala Tyr Cys Leu Ala			
1665	1670	1675	1680
Thr Gly Cys Val Ser Ile Ile Gly Arg Leu His Val Asn Gln Arg Val			
1685	1690	1695	
Val Val Ala Pro Asp Lys Glu Val Leu Tyr Glu Ala Phe Asp Glu Met			
1700	1705	1710	
Glu Glu Cys Ala Ser Arg Ala Ala Leu Ile Glu Glu Gly Gln Arg Ile			

1715	1720	1725	
Ala Glu Met Leu Lys Ser Lys Ile Gln Gly Leu Leu Gln Gln Ala Ser			
1730	1735	1740	
Lys Gln Ala Gln Asp Ile Gln Pro Ala Met Gln Ala Ser Trp Pro Lys			
1745	1750	1755	1760
Val Glu Gln Phe Trp Ala Arg His Met Trp Asn Phe Ile Ser Gly Ile			
1765	1770	1775	
Gln Tyr Leu Ala Gly Leu Ser Thr Leu Pro Gly Asn Pro Ala Val Ala			
1780	1785	1790	
Ser Met Met Ala Phe Ser Ala Ala Leu Thr Ser Pro Leu Ser Thr Ser			
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Thr Thr Ile Leu Leu Asn Ile Met Gly Gly Trp Leu Ala Ser Gln Ile			
1810	1815	1820	
Ala Pro Pro Ala Gly Ala Thr Gly Phe Val Val Ser Gly Leu Val Gly			
1825	1830	1835	1840
Ala Ala Val Gly Ser Ile Gly Leu Gly Lys Val Leu Val Asp Ile Leu			
1845	1850	1855	
Ala Gly Tyr Gly Ala Gly Ile Ser Gly Ala Leu Val Ala Phe Lys Ile			
1860	1865	1870	
Met Ser Gly Glu Lys Pro Ser Met Glu Asp Val Ile Asn Leu Leu Pro			
1875	1880	1885	
Gly Ile Leu Ser Pro Gly Ala Leu Val Val Gly Val Ile Cys Ala Ala			
1890	1895	1900	
Ile Leu Arg Arg His Val Gly Pro Gly Glu Gly Ala Val Gln Trp Met			
1905	1910	1915	1920
Asn Arg Leu Ile Ala Phe Ala Ser Arg Gly Asn His Val Ala Pro Thr			
1925	1930	1935	
His Tyr Val Thr Glu Ser Asp Ala Ser Gln Arg Val Thr Gln Leu Leu			
1940	1945	1950	
Gly Ser Leu Thr Ile Thr Ser Leu Leu Arg Arg Leu His Asn Trp Ile			
1955	1960	1965	
Thr Glu Asp Cys Pro Ile Pro Cys Ser Gly Ser Trp Leu Arg Asp Val			

1970	1975	1980	
Trp Asp Trp Val Cys Thr Ile Leu Thr Asp Phe Lys Asn Trp Leu Thr			
1985	1990	1995	2000
Ser Lys Leu Phe Pro Lys Leu Pro Gly Leu Pro Phe Ile Ser Cys Gln			
	2005	2010	2015
Lys Gly Tyr Lys Gly Val Trp Ala Gly Thr Gly Ile Met Thr Thr Arg			
	2020	2025	2030
Cys Pro Cys Gly Ala Asn Ile Ser Gly Asn Val Arg Leu Gly Ser Met			
	2035	2040	2045
Arg Ile Thr Gly Pro Lys Thr Cys Met Asn Thr Trp Gln Gly Thr Phe			
	2050	2055	2060
Pro Ile Asn Cys Tyr Thr Glu Gly Gln Cys Ala Pro Lys Pro Pro Thr			
2065	2070	2075	2080
Asn Tyr Lys Thr Ala Ile Trp Arg Val Ala Ala Ser Glu Tyr Ala Glu			
	2085	2090	2095
Val Thr Gln His Gly Ser Tyr Ser Tyr Val Thr Gly Leu Thr Thr Asp			
	2100	2105	2110
Asn Leu Lys Ile Pro Cys Gln Leu Pro Ser Pro Glu Phe Phe Ser Trp			
	2115	2120	2125
Val Asp Gly Val Gln Ile His Arg Phe Ala Pro Thr Pro Lys Pro Phe			
	2130	2135	2140
Phe Arg Asp Glu Val Ser Phe Cys Val Gly Leu Asn Ser Tyr Ala Val			
2145	2150	2155	2160
Gly Ser Gln Leu Pro Cys Glu Pro Glu Pro Asp Ala Asp Val Leu Arg			
	2165	2170	2175
Ser Met Leu Thr Asp Pro Pro His Ile Thr Ala Glu Thr Ala Ala Arg			
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Arg Leu Ala Arg Gly Ser Pro Pro Ser Glu Ala Ser Ser Ser Val Ser			
	2195	2200	2205
Gln Leu Ser Ala Pro Ser Leu Arg Ala Thr Cys Thr Thr His Ser Asn			
	2210	2215	2220
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 Val Ala Gln Thr Glu Pro Glu Ser Arg Val Pro Val Leu Asp Phe Leu
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 Glu Pro Met Ala Glu Glu Glu Ser Asp Leu Glu Pro Ser Ile Pro Ser
 2260 2265 2270
 Glu Cys Met Leu Pro Arg Ser Gly Phe Pro Arg Ala Leu Pro Ala Trp
 2275 2280 2285
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 Asp Tyr Gln Pro Pro Thr Val Ala Gly Cys Ala Leu Pro Pro Pro Lys
 2305 2310 2315 2320
 Lys Ala Pro Thr Pro Pro Pro Arg Arg Arg Arg Thr Val Gly Leu Ser
 2325 2330 2335
 Glu Ser Thr Ile Ser Glu Ala Leu Gln Gln Leu Ala Ile Lys Thr Phe
 2340 2345 2350
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 2370 2375 2380
 Glu Thr Gly Ser Ala Ser Ser Met Pro Pro Leu Glu Gly Glu Pro Gly
 2385 2390 2395 2400
 Asp Pro Asp Leu Glu Ser Asp Gln Val Glu Leu Gln Pro Pro Pro Gln
 2405 2410 2415
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 2420 2425 2430
 Ser Glu Glu Asp Asp Thr Thr Val Cys Cys Ser Met Ser Tyr Ser Trp
 2435 2440 2445
 Thr Gly Ala Leu Ile Thr Pro Cys Ser Pro Glu Glu Glu Lys Leu Pro
 2450 2455 2460
 Ile Asn Pro Leu Ser Asn Ser Leu Leu Arg Tyr His Asn Lys Val Tyr
 2465 2470 2475 2480
 Cys Thr Thr Ser Lys Ser Ala Ser Gln Arg Ala Lys Lys Val Thr Phe

2485	2490	2495
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2515	2520	2525
Glu Ala Cys Gln Leu Thr Pro Pro His Ser Ala Arg Ser Lys Tyr Gly		
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Phe Gly Ala Lys Glu Val Arg Ser Leu Ser Gly Arg Ala Val Asn His		
2545	2550	2555
Ile Lys Ser Val Trp Lys Asp Leu Leu Glu Asp Pro Gln Thr Pro Ile		
2565	2570	2575
Pro Thr Thr Ile Met Ala Lys Asn Glu Val Phe Cys Val Asp Pro Ala		
2580	2585	2590
Lys Gly Gly Lys Lys Pro Ala Arg Leu Ile Val Tyr Pro Asp Leu Gly		
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Val Arg Val Cys Glu Lys Met Ala Leu Tyr Asp Ile Thr Gln Lys Leu		
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Pro Gln Ala Val Met Gly Ala Ser Tyr Gly Phe Gln Tyr Ser Pro Ala		
2625	2630	2635
Gln Arg Val Glu Tyr Leu Leu Lys Ala Trp Ala Glu Lys Lys Asp Pro		
2645	2650	2655
Met Gly Phe Ser Tyr Asp Thr Arg Cys Phe Asp Ser Thr Val Thr Glu		
2660	2665	2670
Arg Asp Ile Arg Thr Glu Glu Ser Ile Tyr Gln Ala Cys Ser Leu Pro		
2675	2680	2685
Glu Glu Ala Arg Thr Ala Ile His Ser Leu Thr Glu Arg Leu Tyr Val		
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Gly Gly Pro Met Phe Asn Ser Lys Gly Gln Thr Cys Gly Tyr Arg Arg		
2705	2710	2715
Cys Arg Ala Ser Gly Val Leu Thr Thr Ser Met Gly Asn Thr Ile Thr		
2725	2730	2735
Cys Tyr Val Lys Ala Leu Ala Ala Cys Lys Ala Ala Gly Ile Val Ala		

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Pro Thr Met Leu Val Cys Gly Asp Asp Leu Val Val Ile Ser Glu Ser			
2755	2760	2765	
Gln Gly Thr Glu Glu Asp Glu Arg Asn Leu Arg Ala Phe Thr Glu Ala			
2770	2775	2780	
Met Thr Arg Tyr Ser Ala Pro Pro Gly Asp Pro Pro Arg Pro Glu Tyr			
2785	2790	2795	2800
Asp Leu Glu Leu Ile Thr Ser Cys Ser Ser Asn Val Ser Val Ala Leu			
2805	2810	2815	
Gly Pro Arg Gly Arg Arg Arg Tyr Tyr Leu Thr Arg Asp Pro Thr Thr			
2820	2825	2830	
Pro Leu Ala Arg Ala Ala Trp Glu Thr Val Arg His Ser Pro Ile Asn			
2835	2840	2845	
Ser Trp Leu Gly Asn Ile Ile Gln Tyr Ala Pro Thr Ile Trp Val Arg			
2850	2855	2860	
Met Val Leu Met Thr His Phe Phe Ser Ile Leu Met Val Gln Asp Thr			
2865	2870	2875	2880
Leu Asp Gln Asn Leu Asn Phe Glu Met Tyr Gly Ser Val Tyr Ser Val			
2885	2890	2895	
Asn Pro Leu Asp Leu Pro Ala Ile Ile Glu Arg Leu His Gly Leu Asp			
2900	2905	2910	
Ala Phe Ser Met His Thr Tyr Ser His His Glu Leu Thr Arg Val Ala			
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Ser Ala Leu Arg Lys Leu Gly Ala Pro Pro Leu Arg Val Trp Lys Ser			
2930	2935	2940	
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Ala Val Cys Gly Arg Tyr Leu Phe Asn Trp Ala Val Lys Thr Lys Leu			
2965	2970	2975	
Lys Leu Thr Pro Leu Pro Glu Ala Arg Leu Leu Asp Leu Ser Ser Trp			
2980	2985	2990	
Phe Thr Val Gly Ala Gly Gly Gly Asp Ile Phe His Ser Val Ser Arg			

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Ala Arg Pro Arg Ser Leu Leu Phe Gly Leu Leu Leu Leu Phe Val Gly
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Val Gly Leu Phe Leu Leu Pro Ala Arg
      3025              3030

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<212> DNA

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aagactgggt cctttcttgg ataaaccac tctatgcccg gccatttggg cgtgcccccg 240

caagactgct agccgagtag cgttgggttg cgaaaggcct tgtgggtactg cctgataggg 300

tgcttgccag tgccccggga ggtctcgtag accgtgcacc atg agc aca aat ccc 355
      Met Ser Thr Asn Pro
              1              5

aaa cct caa aga aaa acc aaa aga aac act aac cgt cgc cca caa gac 403

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 Pro Arg Arg Gly Pro Arg Leu Gly Val Arg Ala Thr Arg Lys Ala Ser
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 cgc tcc act ggc aag tcc tgg ggg aag cca gga tac ccc tgg ccc ctg 595
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 Tyr Gly Asn Glu Gly Leu Gly Trp Ala Gly Trp Leu Leu Ser Pro Arg
 90 95 100
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 Gly Ser Arg Pro Ser Trp Gly Pro Asn Asp Pro Arg His Arg Ser Arg
 105 110 115
 aat gtg ggt aag gtc atc gat acc cta acg tgc ggc ttt gcc gac ctc 739
 Asn Val Gly Lys Val Ile Asp Thr Leu Thr Cys Gly Phe Ala Asp Leu
 120 125 130
 ttg ggg tac gtc ccc gtc gta ggc gcc ccg ctt agt ggc gtt gcc agt 787

Leu Gly Tyr Val Pro Val Val Gly Ala Pro Leu Ser Gly Val Ala Ser
 135 140 145

 gct ctc gcg cac gcc gtg aga gtc ctg gag gac ggg gtt aat ttt gca 835
 Ala Leu Ala His Gly Val Arg Val Leu Glu Asp Gly Val Asn Phe Ala
 150 155 160 165

 aca ggg aac tta cct ggt tgc tcc ttt tct atc ttc ttg ctg gcc cta 883
 Thr Gly Asn Leu Pro Gly Cys Ser Phe Ser Ile Phe Leu Leu Ala Leu
 170 175 180

 ctg tcc tgc atc act act ccg gtc tct gct gtc caa gtg aag aac acc 931
 Leu Ser Cys Ile Thr Thr Pro Val Ser Ala Val Gln Val Lys Asn Thr
 185 190 195

 agc aac gcc tat atg gcg act aac gac tgt tcc aat gac agc atc act 979
 Ser Asn Ala Tyr Met Ala Thr Asn Asp Cys Ser Asn Asp Ser Ile Thr
 200 205 210

 tgg cag ctt gag gcc gca gtc ctc cat gtc ccc ggg tgc gtc ccg tgc 1027
 Trp Gln Leu Glu Ala Ala Val Leu His Val Pro Gly Cys Val Pro Cys
 215 220 225

 gag aaa atg ggg aac aca tca cgg tgc tgg ata cca gtc tca cca aac 1075
 Glu Lys Met Gly Asn Thr Ser Arg Cys Trp Ile Pro Val Ser Pro Asn
 230 235 240 245

 gtg gct gtg cgg cag cct gcc gcc ctc acg cgg gcc ttg cgg acg cac 1123
 Val Ala Val Arg Gln Pro Gly Ala Leu Thr Arg Gly Leu Arg Thr His
 250 255 260

 atc gac atg gtc gtg ttg tcc gcc acg ctc tgc tcc gct ctc tac gtg 1171

Ile Asp Met Val Val Leu Ser Ala Thr Leu Cys Ser Ala Leu Tyr Val
 265 270 275
 ggg gac ctc tgt ggc ggg gtg atg ctc gcg tcc cag atg ttc att gtc 1219
 Gly Asp Leu Cys Gly Gly Val Met Leu Ala Ser Gln Met Phe Ile Val
 280 285 290
 tcg ccg cag cac cac tgg ttc gtg cag gaa tgc aat tgc tcc atc tac 1267
 Ser Pro Gln His His Trp Phe Val Gln Glu Cys Asn Cys Ser Ile Tyr
 295 300 305
 cct ggc gcc atc act ggg cac cgt atg gca tgg gac atg atg atg aac 1315
 Pro Gly Ala Ile Thr Gly His Arg Met Ala Trp Asp Met Met Met Asn
 310 315 320 325
 tgg tcg ccc acg acc acc atg atc ctg gcg tac gtg atg cgc gtt ccc 1363
 Trp Ser Pro Thr Thr Thr Met Ile Leu Ala Tyr Val Met Arg Val Pro
 330 335 340
 gag gtc atc ata gac atc att agc gga gct cac tgg ggc gtc atg ttt 1411
 Glu Val Ile Ile Asp Ile Ile Ser Gly Ala His Trp Gly Val Met Phe
 345 350 355
 ggc ctg gcc tac ttc tct atg cag gga gcg tgg gcg aag gtc gtt gtc 1459
 Gly Leu Ala Tyr Phe Ser Met Gln Gly Ala Trp Ala Lys Val Val Val
 360 365 370
 atc ctc ctg ctg gcc tct ggg gtg gac gcg tac acc acc acg act ggg 1507
 Ile Leu Leu Leu Ala Ser Gly Val Asp Ala Tyr Thr Thr Thr Thr Gly
 375 380 385
 agc gct gct ggg cgc act acc agt agc ctg gcc agc gcc ttc tcc cct 1555

Ser Ala Ala Gly Arg Thr Thr Ser Ser Leu Ala Ser Ala Phe Ser Pro
 390 395 400 405

 ggc gct cgg cag aac att cag ctc att aat acc aat ggt agc tgg cac 1603
 Gly Ala Arg Gln Asn Ile Gln Leu Ile Asn Thr Asn Gly Ser Trp His
 410 415 420

 atc aac cgc acc gcc ctg aat tgc aac gat tcc ttg cac acc ggc ttc 1651
 Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser Leu His Thr Gly Phe
 425 430 435

 ttc acg gcc ctg ttc tac atc cat aag ttc aac tcg tcg gga tgt ccc 1699
 Phe Thr Ala Leu Phe Tyr Ile His Lys Phe Asn Ser Ser Gly Cys Pro
 440 445 450

 gag cgc ctg tcc gcc tgt cgc aac atc gag gac ttc cgg ata gga tgg 1747
 Glu Arg Leu Ser Ala Cys Arg Asn Ile Glu Asp Phe Arg Ile Gly Trp
 455 460 465

 ggc gcc ctg caa tac gac gac aat gtc acc aat cca gaa gat atg agg 1795
 Gly Ala Leu Gln Tyr Asp Asp Asn Val Thr Asn Pro Glu Asp Met Arg
 470 475 480 485

 cca tat tgc tgg cac tac cca cca aaa cag tgt ggc gta gtc ccc gca 1843
 Pro Tyr Cys Trp His Tyr Pro Pro Lys Gln Cys Gly Val Val Pro Ala
 490 495 500

 ggg acc gtg tgc ggc cca gtg tac tgt ttc acc cct agc ccg gtg gta 1891
 Gly Thr Val Cys Gly Pro Val Tyr Cys Phe Thr Pro Ser Pro Val Val
 505 510 515

 gtg ggc acg acc gat aga ctt gga gtg cct act tac acg tgg gga gag 1939

Val Gly Thr Thr Asp Arg Leu Gly Val Pro Thr Tyr Thr Trp Gly Glu
 520 525 530
 aat gag aca gat gtc ttc cta ttg aac agc acc cga cca ccg tcg ggg 1987
 Asn Glu Thr Asp Val Phe Leu Leu Asn Ser Thr Arg Pro Pro Ser Gly
 535 540 545
 tca tgg ttt ggc tgc acg tgg atg aac tcc act ggc ttc acc aag acc 2035
 Ser Trp Phe Gly Cys Thr Trp Met Asn Ser Thr Gly Phe Thr Lys Thr
 550 555 560 565
 tgc ggc gca cca ccc tgc cgc act aga gct gac ttc aat acc agc aca 2083
 Cys Gly Ala Pro Pro Cys Arg Thr Arg Ala Asp Phe Asn Thr Ser Thr
 570 575 580
 gat ctg ttg tgc ccc acg gac tgt ttt aga aaa cat cct gaa gcc act 2131
 Asp Leu Leu Cys Pro Thr Asp Cys Phe Arg Lys His Pro Glu Ala Thr
 585 590 595
 tac atc aaa tgt ggt tcc ggg cct tgg ctc acg cca aag tgt ctg gtt 2179
 Tyr Ile Lys Cys Gly Ser Gly Pro Trp Leu Thr Pro Lys Cys Leu Val
 600 605 610
 gac tac ccc tac agg ctc tgg cat tac cct tgc aca gtc aat tac tcc 2227
 Asp Tyr Pro Tyr Arg Leu Trp His Tyr Pro Cys Thr Val Asn Tyr Ser
 615 620 625
 acc ttc aag atc agg atg tat gtg ggg gga gtt gag cac agg ctc atg 2275
 Thr Phe Lys Ile Arg Met Tyr Val Gly Gly Val Glu His Arg Leu Met
 630 635 640 645
 gcc ggc tgc aat ttc act cgt ggg gat cgc tgc aac ttg gag gat agg 2323

Ala Ala Cys Asn Phe Thr Arg Gly Asp Arg Cys Asn Leu Glu Asp Arg
650 655 660

gac aga agt caa cag act cct ctg ttg cac tcc acc acg gaa tgg gcc 2371
Asp Arg Ser Gln Gln Thr Pro Leu Leu His Ser Thr Thr Glu Trp Ala
665 670 675

att ttg ccc tgc tct ttc tca gac ttg ccc gct ttg tcc act ggt ctt 2419
Ile Leu Pro Cys Ser Phe Ser Asp Leu Pro Ala Leu Ser Thr Gly Leu
680 685 690

ctc cac ctc cac caa aat atc gtg gac gta caa tat atg tat ggc ctg 2467
Leu His Leu His Gln Asn Ile Val Asp Val Gln Tyr Met Tyr Gly Leu
695 700 705

tca cct gcc ctc aca caa tat atc gtt cga tgg gag tgg gta gta ctc 2515
Ser Pro Ala Leu Thr Gln Tyr Ile Val Arg Trp Glu Trp Val Val Leu
710 715 720 725

tta ttc ctg ctc cta gcg gac gcc agg gtc tgc gcc tgc ttg tgg atg 2563
Leu Phe Leu Leu Leu Ala Asp Ala Arg Val Cys Ala Cys Leu Trp Met
730 735 740

ctc atc ttg ctg ggc caa gcc gaa gca gca ctg gag aag ctg gtc gtc 2611
Leu Ile Leu Leu Gly Gln Ala Glu Ala Ala Leu Glu Lys Leu Val Val
745 750 755

ttg cac gct gcg agc gca gct agc tgc aat ggc ttc ctg tat ttt gtc 2659
Leu His Ala Ala Ser Ala Ala Ser Cys Asn Gly Phe Leu Tyr Phe Val
760 765 770

atc ttt ctc gtg gct gct tgg cac atc aag ggt agg gtg gtc ccc ttg 2707

Ile Phe Leu Val Ala Ala Trp His Ile Lys Gly Arg Val Val Pro Leu
 775 780 785

gct gct tat tcc ctt act ggc ctg tgg ccg ttc tgc cta ctg ctc cta 2755
 Ala Ala Tyr Ser Leu Thr Gly Leu Trp Pro Phe Cys Leu Leu Leu Leu
 790 795 800 805

gca ctg ccc cag cag gct tac gcc tat gat gca tct gtg cac gga cag 2803
 Ala Leu Pro Gln Gln Ala Tyr Ala Tyr Asp Ala Ser Val His Gly Gln
 810 815 820

gtg ggc gcg gct ttg cta gta ctg att acc ctc ttt aca ctc acc ccg 2851
 Val Gly Ala Ala Leu Leu Val Leu Ile Thr Leu Phe Thr Leu Thr Pro
 825 830 835

ggg tat aag acc ctt ctc agc cag tcc ctg tgg tgg ttg tgc tat ctc 2899
 Gly Tyr Lys Thr Leu Leu Ser Gln Ser Leu Trp Trp Leu Cys Tyr Leu
 840 845 850

ctg acc ctg gcg gaa acc atg gtc cag gag tgg gca cca tcc atg cag 2947
 Leu Thr Leu Ala Glu Thr Met Val Gln Glu Trp Ala Pro Ser Met Gln
 855 860 865

gcg cgc ggc ggc cgt gat ggc atc ata tgg gcc gcc acc ata ttt tgc 2995
 Ala Arg Gly Gly Arg Asp Gly Ile Ile Trp Ala Ala Thr Ile Phe Cys
 870 875 880 885

ccg ggc gta gtg ttt gac ata acc aag tgg ctc tta gcg gtg ctt ggg 3043
 Pro Gly Val Val Phe Asp Ile Thr Lys Trp Leu Leu Ala Val Leu Gly
 890 895 900

cct ggt tac ctc cta aga ggt gct ttg acg cgc gtg cca tat ttc gtc 3091

Pro Gly Tyr Leu Leu Arg Gly Ala Leu Thr Arg Val Pro Tyr Phe Val
 905 910 915
 aga gcc cac gct ctg ctg aga atg tgc act atg gtg agg cac ctc gcg 3139
 Arg Ala His Ala Leu Leu Arg Met Cys Thr Met Val Arg His Leu Ala
 920 925 930
 ggg ggt agg tac gtc cag atg gcg cta tta gcc ctt ggc agg tgg act 3187
 Gly Gly Arg Tyr Val Gln Met Ala Leu Leu Ala Leu Gly Arg Trp Thr
 935 940 945
 ggc act tac atc tat gac cac ctc acc cct atg tcg gat tgg gct gct 3235
 Gly Thr Tyr Ile Tyr Asp His Leu Thr Pro Met Ser Asp Trp Ala Ala
 950 955 960 965
 agc gcc ctg cgg gac ttg gcg gtc gct gtg gag cct atc atc ttc agt 3283
 Ser Gly Leu Arg Asp Leu Ala Val Ala Val Glu Pro Ile Ile Phe Ser
 970 975 980
 ccg atg gag aag aaa gtc atc gtt tgg gga gcg gag acg gct gcg tgc 3331
 Pro Met Glu Lys Lys Val Ile Val Trp Gly Ala Glu Thr Ala Ala Cys
 985 990 995
 ggg gac atc ttg cac gga ctt ccc gtg tcc gcc cga ctc ggt cgg gag 3379
 Gly Asp Ile Leu His Gly Leu Pro Val Ser Ala Arg Leu Gly Arg Glu
 1000 1005 1010
 atc ctc ctt ggc cca gct gat ggc tac acc tcc aag ggg tgg aag ctt 3427
 Ile Leu Leu Gly Pro Ala Asp Gly Tyr Thr Ser Lys Gly Trp Lys Leu
 1015 1020 1025
 ctc gcc ccc atc acc gct tac gcc cag cag aca cga ggt ctc ttg gcc 3475

Leu Ala Pro Ile Thr Ala Tyr Ala Gln Gln Thr Arg Gly Leu Leu Gly
 1030 1035 1040 1045

 tct ata gtg gtg agc atg acg ggg cgt gac aag aca gaa cag gcc ggg 3523
 Ser Ile Val Val Ser Met Thr Gly Arg Asp Lys Thr Glu Gln Ala Gly
 1050 1055 1060

 gag gtc caa gtc ctg tcc aca gtc act cag tcc ttc ctc gga aca tcc 3571
 Glu Val Gln Val Leu Ser Thr Val Thr Gln Ser Phe Leu Gly Thr Ser
 1065 1070 1075

 att tcg ggg gtc tta tgg act gtt tac cac gga gct ggc aac aag aca 3619
 Ile Ser Gly Val Leu Trp Thr Val Tyr His Gly Ala Gly Asn Lys Thr
 1080 1085 1090

 cta gcc ggc tcg cgg ggc ccg gtc acg cag atg tac tcg agc gcc gag 3667
 Leu Ala Gly Ser Arg Gly Pro Val Thr Gln Met Tyr Ser Ser Ala Glu
 1095 1100 1105

 ggg gac ttg gtc ggg tgg ccc agc cct cct ggg acc aaa tct ttg gag 3715
 Gly Asp Leu Val Gly Trp Pro Ser Pro Pro Gly Thr Lys Ser Leu Glu
 1110 1115 1120 1125

 ccg tgt acg tgt gga gcg gtc gac ctg tat ttg gtc acg cgg aac gct 3763
 Pro Cys Thr Cys Gly Ala Val Asp Leu Tyr Leu Val Thr Arg Asn Ala
 1130 1135 1140

 gat gtc atc ccg gct cga aga cgc ggg gac aag cgg gga gcg ctg ctc 3811
 Asp Val Ile Pro Ala Arg Arg Arg Gly Asp Lys Arg Gly Ala Leu Leu
 1145 1150 1155

 tcc ccg aga ccc ctt tcg acc ttg aag ggg tcc tcg ggg gga cct gtg 3859

Ser Pro Arg Pro Leu Ser Thr Leu Lys Gly Ser Ser Gly Gly Pro Val
 1160 1165 1170
 ctt tgc cct agg ggc cac gct gtc gga atc ttc cgg gca gct gtg tgc 3907
 Leu Cys Pro Arg Gly His Ala Val Gly Ile Phe Arg Ala Ala Val Cys
 1175 1180 1185
 tct cgg ggt gtg gct aag tcc ata gat ttc atc ccc gtt gag acg ctc 3955
 Ser Arg Gly Val Ala Lys Ser Ile Asp Phe Ile Pro Val Glu Thr Leu
 1190 1195 1200 1205
 gac atc gtc acg cgg tct ccc acc ttt agt gac aac agc aca cca cca 4003
 Asp Ile Val Thr Arg Ser Pro Thr Phe Ser Asp Asn Ser Thr Pro Pro
 1210 1215 1220
 gct gtg ccc cag acc tat cag gtg ggg tac ttg cac gcc ccc act ggc 4051
 Ala Val Pro Gln Thr Tyr Gln Val Gly Tyr Leu His Ala Pro Thr Gly
 1225 1230 1235
 agt gga aaa agc acc aag gtc ccc gtc gcg tac gcc gcc cag ggg tat 4099
 Ser Gly Lys Ser Thr Lys Val Pro Val Ala Tyr Ala Ala Gln Gly Tyr
 1240 1245 1250
 aaa gtg ctg gtg ctc aat ccc tcg gtg gct gcc acc ctg gga ttt ggg 4147
 Lys Val Leu Val Leu Asn Pro Ser Val Ala Ala Thr Leu Gly Phe Gly
 1255 1260 1265
 gcg tac ttg tcc aag gca cat ggc atc aac ccc aac att agg act gga 4195
 Ala Tyr Leu Ser Lys Ala His Gly Ile Asn Pro Asn Ile Arg Thr Gly
 1270 1275 1280 1285
 gtc aga act gtg acg acc ggg gag ccc att aca tac tcc acg tat ggt 4243

Val Arg Thr Val Thr Thr Gly Glu Pro Ile Thr Tyr Ser Thr Tyr Gly
 1290 1295 1300

 aaa ttc ctc gcc gat ggg ggc tgc gca ggc ggc gcc tat gac atc atc 4291
 Lys Phe Leu Ala Asp Gly Gly Cys Ala Gly Gly Ala Tyr Asp Ile Ile
 1305 1310 1315

 ata tgc gat gaa tgc cac tct gtg gat gct acc act att ctc ggc atc 4339
 Ile Cys Asp Glu Cys His Ser Val Asp Ala Thr Thr Ile Leu Gly Ile
 1320 1325 1330

 ggg aca gtc ctt gac caa gca gag aca gcc ggg gtc agg cta act gta 4387
 Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly Val Arg Leu Thr Val
 1335 1340 1345

 ctg gcc acg gcc acg ccc ccc ggg tcg gtg aca acc ccc cat ccc aat 4435
 Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr Thr Pro His Pro Asn
 1350 1355 1360 1365

 ata gag gag gta gcc ctc gga cag gag ggt gag atc ccc ttc tat ggg 4483
 Ile Glu Glu Val Ala Leu Gly Gln Glu Gly Glu Ile Pro Phe Tyr Gly
 1370 1375 1380

 agg gcg ttt ccc ctg tct tac atc aag gga ggg agg cac ttg att ttc 4531
 Arg Ala Phe Pro Leu Ser Tyr Ile Lys Gly Gly Arg His Leu Ile Phe
 1385 1390 1395

 tgc cac tca aag aaa aag tgt gac gag ctc gca acg gcc ctt cgg ggc 4579
 Cys His Ser Lys Lys Lys Cys Asp Glu Leu Ala Thr Ala Leu Arg Gly
 1400 1405 1410

 atg ggc ttg aac gct gtg gca tat tac aga ggg ttg gac gtc tcc ata 4627

Met Gly Leu Asn Ala Val Ala Tyr Tyr Arg Gly Leu Asp Val Ser Ile
1415 1420 1425

ata cca act caa gga gat gtg gtg gtc gtt gcc acc gac gcc ctc atg 4675
Ile Pro Thr Gln Gly Asp Val Val Val Val Ala Thr Asp Ala Leu Met
1430 1435 1440 1445

acg ggg tat act gga gac ttt gac tcc gtg atc gac tgc aac gta gcg 4723
Thr Gly Tyr Thr Gly Asp Phe Asp Ser Val Ile Asp Cys Asn Val Ala
1450 1455 1460

gtc acc cag gcc gta gac ttc agc ctg gac ccc acc ttc act ata acc 4771
Val Thr Gln Ala Val Asp Phe Ser Leu Asp Pro Thr Phe Thr Ile Thr
1465 1470 1475

aca cag act gtc ccg caa gac gct gtc tca cgt agt cag cgc cga ggg 4819
Thr Gln Thr Val Pro Gln Asp Ala Val Ser Arg Ser Gln Arg Arg Gly
1480 1485 1490

cgc acg ggt aga gga aga ctg ggc att tat agg tat gtt tcc act ggt 4867
Arg Thr Gly Arg Gly Arg Leu Gly Ile Tyr Arg Tyr Val Ser Thr Gly
1495 1500 1505

gag cga gcc tca gga atg ttt gac agt gta gta ctc tgt gag tgc tac 4915
Glu Arg Ala Ser Gly Met Phe Asp Ser Val Val Leu Cys Glu Cys Tyr
1510 1515 1520 1525

gac gca gga gct gct tgg tat gag ctc tca cca gtg gag acg acc gtc 4963
Asp Ala Gly Ala Ala Trp Tyr Glu Leu Ser Pro Val Glu Thr Thr Val
1530 1535 1540

agg ctc agg gcg tat ttc aac acg cct ggc ttg cct gtg tgc cag gac 5011

Arg Leu Arg Ala Tyr Phe Asn Thr Pro Gly Leu Pro Val Cys Gln Asp
 1545 1550 1555

cac ctt gag ttt tgg gag gca gtt ttc acc ggc ctc aca cac ata gac 5059
 His Leu Glu Phe Trp Glu Ala Val Phe Thr Gly Leu Thr His Ile Asp
 1560 1565 1570

gct cat ttc ctt tcc cag aca aag cag tcg ggg gaa aat ttc gca tac 5107
 Ala His Phe Leu Ser Gln Thr Lys Gln Ser Gly Glu Asn Phe Ala Tyr
 1575 1580 1585

tta gta gcc tat cag gcc aca gtg tgc gcc agg gcc aaa gcg ccc ccc 5155
 Leu Val Ala Tyr Gln Ala Thr Val Cys Ala Arg Ala Lys Ala Pro Pro
 1590 1595 1600 1605

ccg tcc tgg gac gtc atg tgg aag tgc ttg act cga ctc aag ccc acg 5203
 Pro Ser Trp Asp Val Met Trp Lys Cys Leu Thr Arg Leu Lys Pro Thr
 1610 1615 1620

ctt gtg ggc cct aca cct ctc ctg tac cgt ttg ggc tct gtt acc aac 5251
 Leu Val Gly Pro Thr Pro Leu Leu Tyr Arg Leu Gly Ser Val Thr Asn
 1625 1630 1635

gag gtc acc ctt aca cac ccc gtg aca aaa tac atc gcc aca tgc atg 5299
 Glu Val Thr Leu Thr His Pro Val Thr Lys Tyr Ile Ala Thr Cys Met
 1640 1645 1650

caa gct gac ctc gag gtc atg acc agc acg tgg gtc ctg gct ggg gga 5347
 Gln Ala Asp Leu Glu Val Met Thr Ser Thr Trp Val Leu Ala Gly Gly
 1655 1660 1665

gtc tta gca gcc gtc gcc gcg tat tgc tta gcg acc ggg tgt gtt tcc 5395

Val Leu Ala Ala Val Ala Ala Tyr Cys Leu Ala Thr Gly Cys Val Ser
 1670 1675 1680 1685

 atc att ggc cgt tta cac atc aac cag cga gct gtc gtc gct ccg gac 5443
 Ile Ile Gly Arg Leu His Ile Asn Gln Arg Ala Val Val Ala Pro Asp
 1690 1695 1700

 aag gag gtc ctc tat gag gct ttt gat gag atg gag gaa tgt gcc tcc 5491
 Lys Glu Val Leu Tyr Glu Ala Phe Asp Glu Met Glu Glu Cys Ala Ser
 1705 1710 1715

 aga gcg gct ctc ctt gaa gag ggg cag cgg ata gcc gag atg ctg aag 5539
 Arg Ala Ala Leu Leu Glu Glu Gly Gln Arg Ile Ala Glu Met Leu Lys
 1720 1725 1730

 tcc aag atc caa ggc tta ttg cag caa gcc tct aaa cag gcc cag gac 5587
 Ser Lys Ile Gln Gly Leu Leu Gln Gln Ala Ser Lys Gln Ala Gln Asp
 1735 1740 1745

 ata caa ccc gct gtg caa gct tcg tgg ccc aag atg gag caa ttc tgg 5635
 Ile Gln Pro Ala Val Gln Ala Ser Trp Pro Lys Met Glu Gln Phe Trp
 1750 1755 1760 1765

 gcc aaa cat atg tgg aac ttc ata agc ggc att cag tac ctc gca gga 5683
 Ala Lys His Met Trp Asn Phe Ile Ser Gly Ile Gln Tyr Leu Ala Gly
 1770 1775 1780

 ctg tca aca ctg cca ggg aac cct gct gtg gct tcc atg atg gca ttc 5731
 Leu Ser Thr Leu Pro Gly Asn Pro Ala Val Ala Ser Met Met Ala Phe
 1785 1790 1795

 agc gcc gcc ctc acc agt ccg ttg tca act agc acc acc atc ctt ctt 5779

Ser Ala Ala Leu Thr Ser Pro Leu Ser Thr Ser Thr Thr Ile Leu Leu
 1800 1805 1810

aac att ctg ggg ggc tgg ctg gcg tcc caa att gcg cca ccc gcg ggg 5827
 Asn Ile Leu Gly Gly Trp Leu Ala Ser Gln Ile Ala Pro Pro Ala Gly
 1815 1820 1825

gcc act ggc ttt gtt gtc agt ggc ctg gtg gga gct gct gtt ggc agc 5875
 Ala Thr Gly Phe Val Val Ser Gly Leu Val Gly Ala Ala Val Gly Ser
 1830 1835 1840 1845

ata ggc ttg ggt aaa gtg ctg gtg gac atc ctg gca ggg tat ggt gcg 5923
 Ile Gly Leu Gly Lys Val Leu Val Asp Ile Leu Ala Gly Tyr Gly Ala
 1850 1855 1860

ggc att tcg ggg gcc ctg gtc gcg ttt aag atc atg tct ggc gag aag 5971
 Gly Ile Ser Gly Ala Leu Val Ala Phe Lys Ile Met Ser Gly Glu Lys
 1865 1870 1875

ccc tcc atg gag gat gtc atc aac ttg ctg cct ggg att ctg tct cca 6019
 Pro Ser Met Glu Asp Val Ile Asn Leu Leu Pro Gly Ile Leu Ser Pro
 1880 1885 1890

ggt gct ctg gtg gtg gga gtc atc tgc gcg gcc att ctg cgc cgc cat 6067
 Gly Ala Leu Val Val Gly Val Ile Cys Ala Ala Ile Leu Arg Arg His
 1895 1900 1905

gtg gga ccg ggg gaa ggc gcg gtc caa tgg atg aac agg ctt atc gcc 6115
 Val Gly Pro Gly Glu Gly Ala Val Gln Trp Met Asn Arg Leu Ile Ala
 1910 1915 1920 1925

ttc gct tcc aga gga aac cac gtc gcc cct act cac tac gtg acg gag 6163

Phe Ala Ser Arg Gly Asn His Val Ala Pro Thr His Tyr Val Thr Glu
 1930 1935 1940
 tcg gat gcg tcg cag cgt gtc acc caa ctg ctt ggc tct ctc act ata 6211
 Ser Asp Ala Ser Gln Arg Val Thr Gln Leu Leu Gly Ser Leu Thr Ile
 1945 1950 1955
 act agt cta ctc agg aga ctt cac aac tgg atc act gag gat tgc ccc 6259
 Thr Ser Leu Leu Arg Arg Leu His Asn Trp Ile Thr Glu Asp Cys Pro
 1960 1965 1970
 atc cca tgc gcc ggc tcg tgg ctc cgc gat gtg tgg gac tgg gtc tgt 6307
 Ile Pro Cys Ala Gly Ser Trp Leu Arg Asp Val Trp Asp Trp Val Cys
 1975 1980 1985
 acc atc cta aca gac ttt aag aac tgg ctg acc tcc aag ctg ttc cca 6355
 Thr Ile Leu Thr Asp Phe Lys Asn Trp Leu Thr Ser Lys Leu Phe Pro
 1990 1995 2000 2005
 aag atg cct ggc ctc ccc ttt atc tct tgc caa aag ggg tac aag ggc 6403
 Lys Met Pro Gly Leu Pro Phe Ile Ser Cys Gln Lys Gly Tyr Lys Gly
 2010 2015 2020
 gtg tgg gcc ggc act ggc atc atg acc aca cga tgc ccc tgc ggc gcc 6451
 Val Trp Ala Gly Thr Gly Ile Met Thr Thr Arg Cys Pro Cys Gly Ala
 2025 2030 2035
 aac atc tct ggc aac gtc cgc ttg ggc tct atg aga atc aca gga ccc 6499
 Asn Ile Ser Gly Asn Val Arg Leu Gly Ser Met Arg Ile Thr Gly Pro
 2040 2045 2050
 aaa acc tgc atg aac acc tgg cag ggg acc ttt cct atc aat tgt tat 6547

Lys Thr Cys Met Asn Thr Trp Gln Gly Thr Phe Pro Ile Asn Cys Tyr
 2055 2060 2065

aca gaa ggc cag tgc ttg ccg aaa ccc gcg tta aac ttc aag acc gcc 6595
 Thr Glu Gly Gln Cys Leu Pro Lys Pro Ala Leu Asn Phe Lys Thr Ala
 2070 2075 2080 2085

atc tgg aga gtg gcg gcc tca gag tac gcg gaa gtg acg cag cac gga 6643
 Ile Trp Arg Val Ala Ala Ser Glu Tyr Ala Glu Val Thr Gln His Gly
 2090 2095 2100

tca tat gcc tat ata aca ggg ctg acc act gac aac tta aaa gtc cct 6691
 Ser Tyr Ala Tyr Ile Thr Gly Leu Thr Thr Asp Asn Leu Lys Val Pro
 2105 2110 2115

tgc caa ctc ccc tct cca gag ttt ttc tct tgg gtg gac gga gta caa 6739
 Cys Gln Leu Pro Ser Pro Glu Phe Phe Ser Trp Val Asp Gly Val Gln
 2120 2125 2130

atc cat agg tcc gcc ccc aca cca aag ccg ttt ttc cgg gat gag gtc 6787
 Ile His Arg Ser Ala Pro Thr Pro Lys Pro Phe Phe Arg Asp Glu Val
 2135 2140 2145

tcg ttc agc gtt ggg ctc aat tca ttt gtc gtc ggg tct cag ctt ccc 6835
 Ser Phe Ser Val Gly Leu Asn Ser Phe Val Val Gly Ser Gln Leu Pro
 2150 2155 2160 2165

tgt gac cct gag ccc gac act gag gta gtg atg tcc atg cta aca gac 6883
 Cys Asp Pro Glu Pro Asp Thr Glu Val Val Met Ser Met Leu Thr Asp
 2170 2175 2180

cca tcc cat atc acg gcg gag gct gca gcg cgg cgt tta gcg cgg ggg 6931

Pro Ser His Ile Thr Ala Glu Ala Ala Ala Arg Arg Leu Ala Arg Gly
 2185 2190 2195
 tca ccc cca tct gag gca agc tcc tca gcg agc cag ctg tcg gcg cca 6979
 Ser Pro Pro Ser Glu Ala Ser Ser Ser Ala Ser Gln Leu Ser Ala Pro
 2200 2205 2210
 tcg ctg cga gcc acc tgc acc acc cac ggt agg acc tat gat gtg gac 7027
 Ser Leu Arg Ala Thr Cys Thr Thr His Gly Arg Thr Tyr Asp Val Asp
 2215 2220 2225
 atg gtg gat gcc aac ctg ttc atg ggg ggc ggc gtg att cgg ata gag 7075
 Met Val Asp Ala Asn Leu Phe Met Gly Gly Gly Val Ile Arg Ile Glu
 2230 2235 2240 2245
 tct gag tcc aaa gtg gtc gtt ctg gac tcc ctc gac tca atg acc gag 7123
 Ser Glu Ser Lys Val Val Val Leu Asp Ser Leu Asp Ser Met Thr Glu
 2250 2255 2260
 gaa gag ggc gac ctt gag cct tca gta cca tcg gag tat atg ctc ccc 7171
 Glu Glu Gly Asp Leu Glu Pro Ser Val Pro Ser Glu Tyr Met Leu Pro
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tgcagatcat gt 9674

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<211> 3033

<212> PRT

<213> Hepatitis C virus

<400> 6

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Gly Val Tyr Leu Leu Pro Arg Arg Gly Pro Arg Leu Gly Val Arg Ala
      35           40           45
Thr Arg Lys Ala Ser Glu Arg Ser Gln Pro Arg Gly Arg Arg Gln Pro
      50           55           60
Ile Pro Lys His Arg Arg Ser Thr Gly Lys Ser Trp Gly Lys Pro Gly
      65           70           75           80
Tyr Pro Trp Pro Leu Tyr Gly Asn Glu Gly Leu Gly Trp Ala Gly Trp
      85           90           95
Leu Leu Ser Pro Arg Gly Ser Arg Pro Ser Trp Gly Pro Asn Asp Pro
      100          105          110
Arg His Arg Ser Arg Asn Val Gly Lys Val Ile Asp Thr Leu Thr Cys
      115          120          125
Gly Phe Ala Asp Leu Leu Gly Tyr Val Pro Val Val Gly Ala Pro Leu
      130          135          140
Ser Gly Val Ala Ser Ala Leu Ala His Gly Val Arg Val Leu Glu Asp
      145          150          155          160
Gly Val Asn Phe Ala Thr Gly Asn Leu Pro Gly Cys Ser Phe Ser Ile
      165          170          175
Phe Leu Leu Ala Leu Leu Ser Cys Ile Thr Thr Pro Val Ser Ala Val
      180          185          190
Gln Val Lys Asn Thr Ser Asn Ala Tyr Met Ala Thr Asn Asp Cys Ser
      195          200          205

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      245                250                255
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      260                265                270
Ser Ala Leu Tyr Val Gly Asp Leu Cys Gly Gly Val Met Leu Ala Ser
      275                280                285
Gln Met Phe Ile Val Ser Pro Gln His His Trp Phe Val Gln Glu Cys
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 Gly Phe Thr Lys Thr Cys Gly Ala Pro Pro Cys Arg Thr Arg Ala Asp
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 Phe Asn Thr Ser Thr Asp Leu Leu Cys Pro Thr Asp Cys Phe Arg Lys
 580 585 590
 His Pro Glu Ala Thr Tyr Ile Lys Cys Gly Ser Gly Pro Trp Leu Thr
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 Pro Lys Cys Leu Val Asp Tyr Pro Tyr Arg Leu Trp His Tyr Pro Cys
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 Glu His Arg Leu Met Ala Ala Cys Asn Phe Thr Arg Gly Asp Arg Cys
 645 650 655
 Asn Leu Glu Asp Arg Asp Arg Ser Gln Gln Thr Pro Leu Leu His Ser
 660 665 670
 Thr Thr Glu Trp Ala Ile Leu Pro Cys Ser Phe Ser Asp Leu Pro Ala
 675 680 685
 Leu Ser Thr Gly Leu Leu His Leu His Gln Asn Ile Val Asp Val Gln
 690 695 700
 Tyr Met Tyr Gly Leu Ser Pro Ala Leu Thr Gln Tyr Ile Val Arg Trp
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 Phe Thr Leu Thr Pro Gly Tyr Lys Thr Leu Leu Ser Gln Ser Leu Trp
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 Trp Leu Cys Tyr Leu Leu Thr Leu Ala Glu Thr Met Val Gln Glu Trp
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 Ala Pro Ser Met Gln Ala Arg Gly Gly Arg Asp Gly Ile Ile Trp Ala
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 Ala Thr Ile Phe Cys Pro Gly Val Val Phe Asp Ile Thr Lys Trp Leu
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 Leu Ala Val Leu Gly Pro Gly Tyr Leu Leu Arg Gly Ala Leu Thr Arg
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 Val Arg His Leu Ala Gly Gly Arg Tyr Val Gln Met Ala Leu Leu Ala
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 Leu Gly Arg Trp Thr Gly Thr Tyr Ile Tyr Asp His Leu Thr Pro Met
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 Ser Asp Trp Ala Ala Ser Gly Leu Arg Asp Leu Ala Val Ala Val Glu
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<220>

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ccccuccccg ggagagccau aguggucugc ggaaccggug aguacaccgg aaungccggg 180
aagacugggu cccuucugg auaaaccac ucuaugcccg gccauuuggg cgugcccccg 240
caagacugcu agccgaguag cguuggguug cgaaaggccu ugugguacug ccugauaggg 300
cgcuugcgag ugccccggga ggucucguag accgugcacc 340

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<211> 340

<212> RNA

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ccccucccg ggagagccau aguggucugc ggaaccggug aguacaccgg aaugccggg 180
aagacugggu ccuuucugg auaaaccac ucuangccc gccauuugg cgugcccccg 240
caagacugcu agccgaguag cguuggguug cgaaaggccu ugugguacug ccugauagg 300
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<210> 11

<211> 236

<212> RNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic RNA

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uauucuacuu ucuuucugg ugguccauc uuagccuag ucacggcuag cugugaaagg 180
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<212> RNA

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 cuacuuucuu ucuugguggc uccaucuuag ccuaggucac ggcuaagcugu gaaagguccg 180
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<211> 17

<212> DNA

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<223> Description of Artificial Sequence: synthetic DNA

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<210> 14

<211> 19

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: synthetic DNA

<400> 14

agtaccacaa ggcctttcg 19

<210> 15

<211> 21

<212> DNA

<213> Artificial Sequence

<220>

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21

<210> 16

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<210> 17

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<400> 17

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<210> 18

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<223> Description of Artificial Sequence: synthetic DNA

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28

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24

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<210> 22

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18

<210> 23

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<210> 24

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<212> DNA

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<223> Description of Artificial Sequence: synthetic DNA

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21

<210> 25

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<210> 26

<211> 27

<212> DNA

<213> Artificial Sequence

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18

<210> 28

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<223> Description of Artificial Sequence: synthetic DNA

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DNA(primer)

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cgtcaagaag gcgatagaag

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<210> 31

<211> 30

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:synthetic DNA

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<210> 32

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<212> DNA

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<223> Description of Artificial Sequence:synthetic DNA

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<210> 33

<211> 24

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence::synthetic DNA

<400> 33

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24

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<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence::synthetic DNA

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<210> 35

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<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence:synthetic

DNA(primer)

<400> 35

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18

<210> 36

<211> 30

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence::synthetic DNA

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<212> DNA

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<212> DNA

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<223> Description of Artificial Sequence::synthetic DNA

<400> 38

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23

<210> 39

<211> 27

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence::synthetic DNA

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<210> 40

<211> 18

<212> DNA

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<223> Description of Artificial Sequence::synthetic DNA

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18

<210> 41

<211> 30

<212> DNA

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<223> Description of Artificial Sequence::synthetic DNA

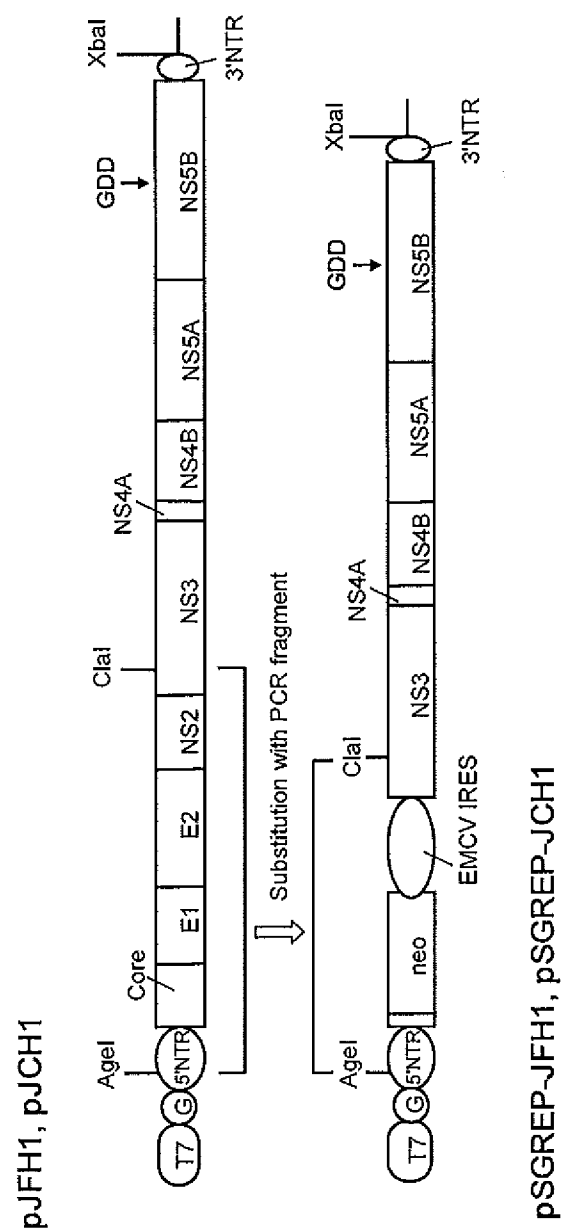
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[Title of Document] Drawings

[Figure 1]



[Figure 2A]

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ACCUGCCCCU AAUAGGGGCG ACACUOCGCC AUGAAUCACU CCCCUGUGAG GAACUACUGU

      70      80      90      100     110     120
CUUCACGCAG AAAGCGCCUA GCCAUGGCGU UAGUAUGAGU GUCCUACAGC CUCCAGGCCC

      130     140     150     160     170     180
CCCCCUGCCG GGAAGGCCAU AGUGGUCUGC GGAACCGGUG AGUACACCGG AAUUGCCGGG

      190     200     210     220     230     240
AAGACUGGGU CCUUCUUGG AUAAACCCAC UCUAUGCCCG GCCAUUUGGG CGUGCCCCCG

      250     260     270     280     290     300
CAAGACUGCU AGCCGAGTAG CGUUGCGUUG GSHAAGGCGU UGUGGUACUG CCUGAUAGGG

      310     320     330     340     350     360
CGCUGGCGAG UGCCCCGGGA GGUUCGUGAG ACCGUGCACC AUGAGGACAA AUCCUAAACC

      370     380     390     400     410     420
UCAAGAGAAA ACCAAAGAA ACACCAACCG UCGCCCAUUG AUUGAACAG AUGGAUUGCA

      430     440     450     460     470     480
CGCAGGUUCU CCGCCCGCUU GGGUGGAGAG GCUAUUCGGC UAUAGUUGGG CACAACAGAC

      490     500     510     520     530     540
AAGCGGUGCG UCUAGUGCGG CCGUGUUCGG GCUUGCAGCG CAGGGGCGCC CGGUUCUUGU

      550     560     570     580     590     600
UGUCAGAGCC GACUGUGCG GUGCCUGAA UGAACUGCAG GACGAGCGAG CCGGGCUAUC

      610     620     630     640     650     660
GUGGUGGGCC ACACGGGGCG UUCUUGGCGC AGCUUGGCUU GACGUUGUCA CUGAAGGGGG

      670     680     690     700     710     720
AAGGGACUGG CUGCUAUUGG GCGAAGUGCC GGGGAGGAGU CUCCUGUCAU CUCACCUUGC

      730     740     750     760     770     780
UCCUGCCGAG AAAGUAUCCA UCAUGGCUGA UGCAAGCGG CCGCUGCAUA CGCUUGAUCC

      790     800     810     820     830     840
GGCUACGUCC CCAUUCGACC ACCAAGCGAA ACAUOGCAUC GAGCGAGCAC GUACUGGGAU

      850     860     870     880     890     900
GGAAGCGGU CUUGUGAUC AGGAUGAUCU GAGCGAAGAG CAUCAGGGGC UCGCGCCAGC

      910     920     930     940     950     960
CGAACUGUUC GCCAGGCUCA AGCGCGGCAU GCGGACGGC GAGGAUCUG UCGUGACCCA

      970     980     990     1000    1010    1020
UGGCGAUGCC UGCUUGCCGA AUUUCAUUGU GGAUAAUGGC CCGUUUUCUG GAUUCAUUGA

      1030    1040    1050    1060    1070    1080
CGUGGCCCG CUGGGUGUGG CGGACCGCUA UCAGGACAU GGGUUGGCUA CCGGUGAUUU

      1090    1100    1110    1120    1130    1140
UGCUAGAGAG CUUGCGGGCG AAUGGCGUGA CGCUCUCUC GUGCUUUAAG GAUUCGCGGC

      1150    1160    1170    1180    1190    1200
UCCCGAUUCG CAGCGCAUGG CCUUCUUCG CCUUCUUGAC GAGUUCUUCU GAGUUUAAAC

      1210    1220    1230    1240    1250    1260
CCUUCGCCGC CCCCCCCC AUUGUACUG GCCGAGCGG CUUGGHAUAA GGGCGUGUG

      1270    1280    1290    1300    1310    1320
CGUUGUCUA UAGGUUUUU UCCACCAUU UGCGGUCUU UGCGAAGUG AGGGCCGGA

      1330    1340    1350    1360    1370    1380
AACCUGGCC UGUUCUUG ACAGCNUK CUGGGGUCU UCCCCUCUC GCCAAGGAA

```

[Figure 2B]

1390	1400	1410	1420	1430	1440
UGCAAGGUCU	GUUGAAGUC	GUUAGGAGG	CAGUCCUCU	GGAAAGCUCU	UGAAGACAA
1450	1460	1470	1480	1490	1500
CAACGUCUGU	AGCGACCCUU	UGCAGGCAGC	GGAAACCCCC	ACCUAGGCGAC	AGGUGCCUCU
1510	1520	1530	1540	1550	1560
GGGGCCAAAA	GCCAGGUGUA	UAGGAUACAC	CUGCAAGGGG	GGCACAACCG	CAGUGCCACG
1570	1580	1590	1600	1610	1620
UUGUGAGUUG	GAUAGUUGUG	GAAAGAGUCA	AAUGGCUCUC	CUCAGGCGUA	UUCACCAAGG
1630	1640	1650	1660	1670	1680
GGCUGAAGGA	UGCCCAAGAG	GUAACCCAUU	GUUAGGGAUC	UGAUCUGGGG	CCCGGGUGCA
1690	1700	1710	1720	1730	1740
CAUGCUUUAC	AUGUGUUUAG	UUGAGGUUAA	AAAAAGGUCU	AGCCCCCCCG	AAACAGGGGG
1750	1760	1770	1780	1790	1800
ACGUGGUAUU	CCUUGAAGAA	ACACGAUGAU	ACCAUGGCUC	CCAUCACUGC	UUUAGCCACG
1810	1820	1830	1840	1850	1860
CAAAACAGAG	GGCUCGCGG	CGCCAUAGUG	GUUAGUAUGA	CGGGGCGUGA	CAGGACAGAA
1870	1880	1890	1900	1910	1920
CAGGCGGGGG	AAUUCCAAAU	CCUGUCCACA	GUCUCUCAGU	CCUUCUCUGG	AACAACCAUC
1930	1940	1950	1960	1970	1980
UUGGGGUAUU	UGGAGGAGUU	UUACCAAGGA	GCUGGCHACA	AGACUCUAGC	CGGCUUACGG
1990	2000	2010	2020	2030	2040
GUUCCGUAUA	CGCAGAUUA	CUCAGGUGCU	GAGGGGAGCU	UGGUAAGGCU	GGCCAGGCCC
2050	2060	2070	2080	2090	2100
CUUGGAGCCA	AGUCUUUGGA	GCCGUGGAGG	UGUGGAGCCG	UUGAAGCUUA	UCUGGUAACG
2110	2120	2130	2140	2150	2160
GGGAAGGCUG	AUGUCAUCCC	GGCUUGGAGA	CGGGGGAGCA	AGCGGGGAGC	AUUGGCUUCC
2170	2180	2190	2200	2210	2220
CGGAGACCCA	UUUGGAGCUU	GAAGGGGUAU	UUGGGGGGAG	CGGUGGCUUG	CCCUAGGGGC
2230	2240	2250	2260	2270	2280
CAUGGUGUUG	GGCUCUUCGG	AGCAGGUGUG	UGCUUCUGGG	GGGUGGCGAA	AUCCAUUGAU
2290	2300	2310	2320	2330	2340
UUCAUCCCCG	UUGAGACACU	CGAGGUGUUG	ACAGGGGUCU	CCAUCUUCAG	UGACACAGGC
2350	2360	2370	2380	2390	2400
ACGCCACCCG	CUGUGCCCCA	GACCUAUCAG	GUUGGGUAU	UGCAUGGUCU	AACUGGCGAU
2410	2420	2430	2440	2450	2460
GGAAAGAGCA	CCAGGUGCCC	UGUCCGUAUU	GCCGCCAGAG	GGACAAAGU	ACUAGUGCUU
2470	2480	2490	2500	2510	2520
AAACCCUCCG	UAGCUGCCAC	CCUGGGUUUU	GGGGGCUACC	UAUCCAGGC	ACAUGGCAUC
2530	2540	2550	2560	2570	2580
AAUCCACACA	UUGAGGAGUG	AGUCAGGACC	GUGAUGACCG	GGGAGGCCAU	CAGGUACUCC
2590	2600	2610	2620	2630	2640
ACAUUAGGCA	AAUUCUUCGU	CGAUGGGGGC	UGCGCUAGCG	GGCCUAUGA	CAUCAUUAU
2650	2660	2670	2680	2690	2700
UGCGAUGAAU	GCCACGCGUU	GGAGGUAACC	UCCAUUCUCC	GCAUGGGAAC	GGUCCUUGAU
2710	2720	2730	2740	2750	2760
CAAGCAGAGG	CAGCGGGGGU	CAGACUAACU	GUGCUUGGUA	CGGCCACACC	CCCGGGGUCA

[Figure 2C]

```

2770      2780      2790      2800      2810      2820
GUUACAACCC CCCAUCCCGA UUAAGAAGAG GUAGGCCUUG GCGGGGAGGG UGAGAUCCCC

2830      2840      2850      2860      2870      2880
UUCUUAUGGA GGGCGAUUCC CCUAUCCUGC AUCAAGGGAG GGAGACACCU GAUUUUCUGC

2890      2900      2910      2920      2930      2940
CACUCAAAGA AAAGUGUGA CGAGCUUGCG GCGGCCUUC GGGCGAUGGG CUUAAUUGCC

2950      2960      2970      2980      2990      3000
GUGGCACACU AUAGAGGGUU GGACGUCUCC AUAAUACCAU CUCAGGGAGA UGUGGUGGUC

3010      3020      3030      3040      3050      3060
GUCGCCACCG ACGCCUCCAU GACGGGGUAC ACUGGAGACU UUGACUCCGU GAUCCGACUGC

3070      3080      3090      3100      3110      3120
AAUGGAGCGG UCAACCAAGC UGUCCACUUC AGCCUUGGAC CCACCUUAC UAUAAACACA

3130      3140      3150      3160      3170      3180
CAGACUUGCC CACAAGAGCG UGUUUCACCG AGUCAGCGCC GCGGGCGCAC AGGUAGAGGA

3190      3200      3210      3220      3230      3240
AGACAGGGCA CUUAUAGGUA UGUUCCACU GUUGAACGAG CCUCAGGAU GUUUGACAGU

3250      3260      3270      3280      3290      3300
GUAGUGCUUU GUAGAGUCUA CGACGCAGGG GCUUGGUGGU ACCAUUCAC ACCAGCGGAG

3310      3320      3330      3340      3350      3360
AGCACCGUCA GGCUTAGAGC GAUUUCCAGC ACGCCCGGCC UACCCGUGUG UCAAGACCAU

3370      3380      3390      3400      3410      3420
CUUGAAUUUU GGGAGGAGU UUUACCGGC CUCACACACA UAGAGGCCCA CUUCCUUCUC

3430      3440      3450      3460      3470      3480
CAAAACAAGC AAGCGGGGGA GAAUUGCGG UACCUAGUAG CUUACCAAGC UACGGUGGCG

3490      3500      3510      3520      3530      3540
GCGAGAGCCA AGGCCCUUC CCGUCCUGG GAGGCCAUGU GGAAGUGCCU GGGCCGACUC

3550      3560      3570      3580      3590      3600
AAGCCUACGC UUGCGGGGCC CACACCUUC CUGUACCGUU UGGGCCUUAU UACCAUUGAG

3610      3620      3630      3640      3650      3660
GUACCCUCA CACACCUUG GACGAAGUAC AUCCGCCAU GCAUGCAAGC UGACCUUGAG

3670      3680      3690      3700      3710      3720
GUCAUAGCCA GCGUGUGGU CUAAGCUGA GAGUCCUGG CAGCGGUGC GCAUUAUUGC

3730      3740      3750      3760      3770      3780
CUGGCGACUG GAUGUGUUUC CAUCAUUGG CCGUUGCAG UCAACCAAGC AGUCGUGGUU

3790      3800      3810      3820      3830      3840
GCGCCGGUUA AGGAGGUCCU GUUAGAGGCU UUGAUGAGA UGAGGAAAG CGCCUUAUGC

3850      3860      3870      3880      3890      3900
GCGGCUUCA UCGAAGAGG GCGCGGAUA GCGAGAUUG UGAAGUCCA GAUCCAAAGC

3910      3920      3930      3940      3950      3960
UUGCUUGAGC AGGCCUUA GAGGCGCCG GACAUACAC CCGCUAUGA GGCUUAUGG

3970      3980      3990      4000      4010      4020
CCCAAGGGG AACAAUUUG GCGCAGACAC AUGUGGACU UCAUAGCGG CAUCCAAUAC

4030      4040      4050      4060      4070      4080
CUCGCGAGAU UGUACACU GCGAGGGAC CCGCGGUGG CUUCCAUGAU GCAUUCAGU

4090      4100      4110      4120      4130      4140
GCGCCCUCA CCAGUCCUU GUAGACCAU ACCACCAUC UUCUACAACU AUGGGAGGC

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[Figure 2D]

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4150      4160      4170      4180      4190      4200
UGGUUAGCGU CCCAGAUCGC ACCACCCGCG GGGGCCACCG GCUUUGUGGU CAGUGGCCUG

4210      4220      4230      4240      4250      4260
GUGGGGUGUG CCGUGGGCAG CAUAGGCCUG GGUAGGGUGC UGGUGGACAU CCUGGCAGGA

4270      4280      4290      4300      4310      4320
UAUGGGGCGG GCAUUUGGGG GGCCCUUGUC GCAUUCAGAG UCAUGUCUGG GGNAGGCCG

4330      4340      4350      4360      4370      4380
UCUAUGGAGG AUGUCUACAA UCUAUCUGCU GGGAUUCUUG CUCCGGAGGC CCUGGUGGUG

4390      4400      4410      4420      4430      4440
GGGUCUAGUG GCGCGGCCAU UCUGCGCCGC CACGUGGGAC CGGGGGAGGG CGCGGUCCAA

4450      4460      4470      4480      4490      4500
UGGAUGAACA GGUUUAUUGG CUUUGCUUCC AGAGGAAACC ACGUCGCCCC UACUACUAC

4510      4520      4530      4540      4550      4560
GGGACGGAGU CGGAGCGGUC GCAGCGUGUG ACCCAACUAC UGGGCUUCUU UACUAAUACC

4570      4580      4590      4600      4610      4620
AGCCUUCUCA GAGACUCCA CAUUGGGAUA ACUAGGGACU GCGCCAUCCC AUGCUCCGGA

4630      4640      4650      4660      4670      4680
UCCUGGCUCC GCGACGUGUG GGAUUGGGUU UGCACCAUCU UGACAGACUU CAAAUAUUGG

4690      4700      4710      4720      4730      4740
CUGACUUGAA AAUUGUCCC CAAGCUGCCC GCGCCUCCCU UCAUCUCUUG UCAAAAGGGG

4750      4760      4770      4780      4790      4800
UACAAGGUGG UGGGGGCCGG CACUGGCAUC AUGACCAAGC GCUGCCUUGG CGGCGCCAAC

4810      4820      4830      4840      4850      4860
AUCUCUGGCA AUGCCGCGCU GGGCUUAUG AGGAUCACAG GGCUAADAC CUGCAUGAAC

4870      4880      4890      4900      4910      4920
ACCGGCGAGG GAGCUUUCU UAUCAAUUGC UACACGGAGG GCGAGUGCGC GCGGAACCC

4930      4940      4950      4960      4970      4980
CCCACGAACU ACAGAACCGC CAUCUGGAGG GUGGCGGCGU CGGAGUACGC GGAAGUAGCG

4990      5000      5010      5020      5030      5040
CAGCAUGGGU CGUACUCCUA UGUAACAGGA CUGACCAUG ACANUCGAA AAUUCUUGG

5050      5060      5070      5080      5090      5100
CAACUACCUU CUCCAGAGUU UUUCUCCUGG GUGGACGGUG UGCAGAUCCA UAGGUUUGCA

5110      5120      5130      5140      5150      5160
CCCACACCAA AGCCGUUUU CCGGGAUGAG GUUUCGUUCU GCGUUGGGCU UAAUUCUUAU

5170      5180      5190      5200      5210      5220
GCUUCGGGU OCCAGCUUC CUUGAAGCU GAGCCCGACG CAGACGUUUU GAGGUCCUUG

5230      5240      5250      5260      5270      5280
CUAACAGAUU CGCCCGACAU CAGGGCGGAG ACUGCGGCGC GCGCUUGGC ACGGGGAUCA

5290      5300      5310      5320      5330      5340
CCUCCAUUGG AGGCGAGCUC CUCAGUGAGC CAGCUAUCAG CACCGUCGCU GCGGGCCACC

5350      5360      5370      5380      5390      5400
UGCAACACCC ACAGCAACAC CUAUGACGUG GACAUGGUUG AUGCCAACU GCUCUUGGAG

5410      5420      5430      5440      5450      5460
GGCGUGUGGG CUCAGACAGA GCCUGAGUCC AGGUUGCCCG UUCUGGACUU UCUCAGCCA

5470      5480      5490      5500      5510      5520
AUGGCGAGGG AAGAGAGCGA CCUUGAGCCC UCAUUAACAU CGAGUGCAU GCUCCCCAGG

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[Figure 2E]

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5530      5540      5550      5560      5570      5580
AGCGGGUUUC CACGGGCCUU ACCGGGCUUGG GCACGGCCUG ACUACAACCC GCGGCUUGUG

5590      5600      5610      5620      5630      5640
GAAUUGUGGA GGAGGGCAGA UUAACCAACC CCCACCGUUG CUGGUUGUGC UCUCCCCCG

5650      5660      5670      5680      5690      5700
CCCAAGAGAG CCCCGACGCC UCCCCCAAGG AGACGCCGGA CAGUGGGUCU GAGGAGAGGC

5710      5720      5730      5740      5750      5760
ACCAUUAUCG AAGCCCCCA GCACUUGGCC AUCAAGACCU UUGGCCAGCC CCCUUCGAGC

5770      5780      5790      5800      5810      5820
GGUGAUGCAG GCUUGUCCAC GGGGGCGGGC GCGGCGGAAU CCGGCGGUCC GACGUCCCCU

5830      5840      5850      5860      5870      5880
GGUGAGCGGG CCCCCUCAGA GACAGGCUCC GCCUCCUUA UGCCCCCCU CAGGGGGAGG

5890      5900      5910      5920      5930      5940
CCUGAGAUUC CGGACCGGA GUCUGAUCAG GUAGAGCUUC AACCCUCCCC CCAGGGGGGG

5950      5960      5970      5980      5990      6000
GGGUAAGCUC CCGUUCGGG CUGGGGGUCU UGGUCUACUU GCUCGAGGA GGACCAUACC

6010      6020      6030      6040      6050      6060
ACCGUGUCU GCGCAUGUC AUACUCCUGG ACCGGGGCUC UAUUAACUCC CUGUAGCCCC

6070      6080      6090      6100      6110      6120
GAAGAGGAAA AGUUGGCAU CAACCCUUUG AGUAACUCGC UGUUGCGAUA CCAUACAAG

6130      6140      6150      6160      6170      6180
UGUUAUCGUA CAAUUAUAA GAGCGCCUCA CAGAGGGCUA AAAAGGUAAC UUUUGACAGG

6190      6200      6210      6220      6230      6240
ACGCAAGUUC UCGACGCCA UUAUGACUA GUCUUAAGG ACUUAAGCU AGCGGCUUC

6250      6260      6270      6280      6290      6300
AAAGUACAGC CAGGCUCCU CACCUUGAG GAGCGGUGCC AGUUGACUCC ACCCAUUCU

6310      6320      6330      6340      6350      6360
GCAAGAUCCA AGUAUGAUA CCGGCGCAG GAGUCCGCA GCUUGUCGG GAGGCGCGU

6370      6380      6390      6400      6410      6420
AACCAUCA AGUCCUGUG GAGGACCCU CUGGAAGACC CACAAACACC AAUUCACCA

6430      6440      6450      6460      6470      6480
ACCAUUAUG CCAAAAUUA GUGGUUCUGU GUGGACCCG CCAAGGGGG UAAGAAACCA

6490      6500      6510      6520      6530      6540
GUCGCGUCA UCGUUUACC UGACCUCCG GUCGCGGUCU GCGAGAAAU GCGCCUUAU

6550      6560      6570      6580      6590      6600
GACAUACAC AAAAGCUCC UGAGGCGUA AUGGAGCUU CCUAUGGCU CCUGUACUC

6610      6620      6630      6640      6650      6660
CCUGCCAC GGUUGAGUA UCUCUUGAA GCAUGGGCG AAAAGAGGA CCGAUUGGU

6670      6680      6690      6700      6710      6720
UUUUGUAUG AUACCGAUG CUUCGACUA ACCGUCUUG AGAGAGACAU CAGGACCGAG

6730      6740      6750      6760      6770      6780
GAGUCCAUU ACCAGCCU CUCUUGCCC GAGGAGGCC GCACUGCCAU ACACCGCUG

6790      6800      6810      6820      6830      6840
ACUGAGACU UUAUGUAG AGGGCCCAUG UUAACAGCA AGGGUCABAC CUGCGGUUAC

6850      6860      6870      6880      6890      6900
AGACUUGCC GCGCAGUG GUGCUAAC ACUAGCAUG GUAAACCAU CCAUGCUAU

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[Figure 2F]

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6910      6920      6930      6940      6950      6960
GUGAAAGCCC UAGCGGCCUG CAAGGCUGCG GGGUAGGUG CGCCACAAU GCUGGUADGC

6970      6980      6990      7000      7010      7020
GCCGAGUAGC UAGUAGUCAU CUCAGAAAGC CAGGGGACUG AGGAGGACGA GCGGAACTCG

7030      7040      7050      7060      7070      7080
AGAGCCUACA CGGAGGCCAU GACGAGGURC UCUGCCCCUC CUGGUGAUCC CCCCAGACCG

7090      7100      7110      7120      7130      7140
GAAUAGAACG UGGAGCUAAU AACAUCCUAG UCCUCAAAG UGUUGUGGC GUGGGCCCG

7150      7160      7170      7180      7190      7200
CGGGCCGCC GCAAGUACUA CCUGACCAAG GACCAACCA CUCCACUCC CCGGCTUGCC

7210      7220      7230      7240      7250      7260
UGGGAACAG UUAGACACUC CCCUACAAU UCAUGGCGUG GAAACAUCA CCAGUUGCU

7270      7280      7290      7300      7310      7320
CCAACCAAU GGUUCGCAU GGUCCUAAU ACACACUUC UCUCAUUCU CAUGGUCCAA

7330      7340      7350      7360      7370      7380
GACGCCCGG ACCAGAACCU CAACUUGAG AUGUAGGAU CAGUUAUCU CGUGAAUCCU

7390      7400      7410      7420      7430      7440
UUGGACCUCC CAGCCUAAU UGAGAGGUUA CACGGGCUUG AGCCUUCUC UAGGCACACA

7450      7460      7470      7480      7490      7500
UUCUCUACC ACGAACUGAC GCGGGUGGU UCAGCCCUCA GAAACUUGG GCGGCCACC

7510      7520      7530      7540      7550      7560
CUCAGGCGU GGAAGAGUC GGCUGGCGA GUCAGGCGU CCCCACUUC CCGUGGAGG

7570      7580      7590      7600      7610      7620
AAAGCGCGG UUGGCGGCC AUUUCUUC AUUUGGCGG UGAAGACCA GCUCAAACUC

7630      7640      7650      7660      7670      7680
ACUCCAUUC CCGAGGCGCG CUUACUGAC UUAUCCAGU GGUUACCGU CCGGCGCGG

7690      7700      7710      7720      7730      7740
GGGGCGACA UUUUCACAG CGUGUGGUC GCGCGGCCG GCUCAUUACU CUUCGGCUA

7750      7760      7770      7780      7790      7800
CUCCUACUU UCGUAGGGU AGGCCUUC CUACUCCCG CUUGUAGAG CGGCACACAC

7810      7820      7830      7840      7850      7860
UAGGUAACU CCUAGCUAA CUGUCCUUU UUUUUUUUU UUUUUUUUU UUUUUUUUU

7870      7880      7890      7900      7910      7920
UUUUUUUUU CUUUUUUUU UUUUUUUU UUUUUUUU UCUUACUUA UUUAUUCUU

7930      7940      7950      7960      7970      7980
UUUUUUUUU GUUCCAUUU AGCCUAGUC ACGGUAGCU GUGAAAGGU CGUAGGCCG

7990      8000      8010      8020      8030      8040
AUGACUGAG AGAUCCCGU AACUGUUC UCUAGAUU AUGU

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[Figure 3A]

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      10      20      30      40      50      60
ACCGGCCCCU AAUAGGGGCG ACACUCCGCC AUGAUAACU CCCCUGUGAG GAAUAUAUGU

      70      80      90     100     110     120
CUUCACGCAG AAGCGUCUA GCCAUGGCGU UAGUAUAAGU GUUGUACAGC CUCCAGGCCC

      130     140     150     160     170     180
CCCCUCCCCG GGAGAGCCAU AGUGGUCUGC GGAAACGGUG AGUACACCGG AAUUGCCCGG

      190     200     210     220     230     240
AAGACUGGCU CCUUUCUUUG AUAACCCAG UCUAUGCCCG GCUUUUGGG CGUGCCCCCG

      250     260     270     280     290     300
CAGACUGGCU AGCGAGUAG CGUUGGCUUG CGAAAGGCCU UGUGUAUCUG CCUGAUAGGG

      310     320     330     340     350     360
UGCUUGCGAG UGGCCCGGGA GGUCUCGUAG ACCUGGCACC AUGAGCACA AUCCCAABACC

      370     380     390     400     410     420
UCAAAGAAAL ACCAABAGAA ACACUAACCG UGCCCCARUG AUUGAACAG AUGGUAUACA

      430     440     450     460     470     480
CGCAGGUUCU CCGCCCGCUU GGGUGGAGAG GCUUUUCGGC UAUGACUGGG CACAAACAGC

      490     500     510     520     530     540
AAUCGGCUGC UCUGAUGCCG CCGUGUCCG GCUGUCAGCG CAGGGCGCGC CGGUCUUUUU

      550     560     570     580     590     600
UGUCAAGACC GACCUUGCCG GUGCCUGABA UGAACUGCGG GACGAGGCGG CCGGCUAUC

      610     620     630     640     650     660
GGGGCUGGCC ACGAGCGGCG UUCUUUGGGC AGCUUGGCUU GACGUUGUCA CUAAAGCGGG

      670     680     690     700     710     720
AAGGGACUGG CUCUAUUUG GCGAAGUGCC GGGGAGGAGU CUCCUUAU CUACCCUUGC

      730     740     750     760     770     780
UCCCGCGAG AAAGUAUCCA UCAUGGCUGA UGCAAGCGCG CGGUGCAUA CGCUUGAUCC

      790     800     810     820     830     840
GGCUACCGGC CCAUUCGACC ACCAAGCGAA ACAUUGCAUC GAGCGAGCAG GUACUUGGAU

      850     860     870     880     890     900
GGAAGCCGCU CUUGUCGAGC AGGAUAUCU GGAAGAGAG CAUCAGGGGC UGCGCGCAGC

      910     920     930     940     950     960
CGAACUGUUC GCGAGGCUCA AGGCGCGCAU GCGGAGGGC GAGGAUUCG UCGUGACCCA

      970     980     990    1000    1010    1020
UGGCGAUGCC UGCUUGCCGA AUAUCUUGU GGAUAAUGGC CGCUUUUCUG GAUUAUUGA

      1030    1040    1050    1060    1070    1080
CUUGGGCGCG CUUGGUGUGG CGGACCGCUA UCAGGACUAU GCGUUGGCUA CCGUGAUUAU

      1090    1100    1110    1120    1130    1140
UGCUAAGAG CUUGGGCGCG AAUGGGCGGA CCGCUUCUUC GUGCUUACG GAUUCGCCGC

      1150    1160    1170    1180    1190    1200
UCCCGAUUCG CAGCGCAUCG CCUUCUAUCG CCUUCUAGC GAGUUCUUCU GAGUUAAAGC

      1210    1220    1230    1240    1250    1260
CCUUCUCCUC CCCCCCCC AACCUUACUG GCGGAAGCCG CUUGGAUUA GCGCGGUGUG

      1270    1280    1290    1300    1310    1320
CGUUGUCUA UAUGUUAUUU UCCACCAUUA UGCGGUCUUU UGGCAUUGG AGGGCCCGGA

      1330    1340    1350    1360    1370    1380
AACCUGGCCC UGUUUCUUG ACGAGCUUC CUAGGGGUCU UCCCCUCUC GCGAAGGAA

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[Figure 3B]

1390	1400	1410	1420	1430	1440
UGCAAGGUCU	GUUGAAUGUC	GUGAAGGAAG	CAGUCCUCU	GGAAGCUCU	UGAAGACAAA
1450	1460	1470	1480	1490	1500
CAACGUCUGU	AGCGACCCUU	UGCAGGCAGC	GGAACCCCC	ACCUGGCAGC	AGGUGCCUCU
1510	1520	1530	1540	1550	1560
GCGGCCAATA	GCCACGUGUA	UARGAUACAC	CUCCAAAGGC	GCCACACCC	CAGUGCCAGC
1570	1580	1590	1600	1610	1620
UUGUGAGUUG	GAUGGUGUG	GAAAGAGUCA	AUUGGCUCUC	CUCAGCGUA	UUCARCAAGG
1630	1640	1650	1660	1670	1680
GCGCAGAGGA	UGCCACAGAG	GUAACCCAUU	GUUUGGUAUC	UGAUCUGGG	CCUGCGUGCA
1690	1700	1710	1720	1730	1740
CAUGGCUUAC	AUGUGUUUAG	UCAGGCUUAA	AAAAACGUCU	AGGCCCCCG	AACACCGGG
1750	1760	1770	1780	1790	1800
AGUGGCUUUU	CCUUGAGAA	ACACGUAUAA	ACCAUGGCC	CCAUCACGC	UUACGCCAG
1810	1820	1830	1840	1850	1860
CAGACACGAG	GUCUCUGGG	CUCUAUUGG	GUGAGCAUGA	CGGGCGUGA	CAAGACAGAA
1870	1880	1890	1900	1910	1920
CAGGCCGGGG	AGGUCCAGU	CCUGGCCACA	GUACUCAGU	CCUUGCCUG	AACHUCCAUU
1930	1940	1950	1960	1970	1980
UGGGGGGUCU	UNUGGACUGU	UUAACACGGA	GCUGGCACCA	AGACACUAGC	CGGCUCGCG
1990	2000	2010	2020	2030	2040
GCCCCGUGCA	CGCAGUGUA	CUCCAGCGCC	GAGGGGACU	UGGUCCGGG	GCDCACCCCU
2050	2060	2070	2080	2090	2100
CCUGGACCA	AUUCUUGGA	GCUGUGUACG	UGUGGAGCG	UCGACCGUA	UUUGGUCACG
2110	2120	2130	2140	2150	2160
CGAAGCGCG	AUUCUUGCC	GCUCGAGAA	CGCGGGGACA	AGCGGGGAGC	GCUGCUCUC
2170	2180	2190	2200	2210	2220
CGAGAGCCCC	UUUGGACCUU	GAGGGGUGCC	UGGGGGGAGC	CUGUGCUUG	CCUAGGGGG
2230	2240	2250	2260	2270	2280
CAGCGUGUG	GAUUCUCCG	GCGAGCUGUG	UGCUCUGGG	GUGUGGCUAA	GUCCAUNGAU
2290	2300	2310	2320	2330	2340
UUCAUCCCC	UUGAGACGCU	CGACAUCCUC	ACGCGGUCUC	CCACCUUUG	UGACAAACAG
2350	2360	2370	2380	2390	2400
ACACCAACAG	CUGUGCCCCA	GACCUACAG	GUGGGGUAU	UGCAGCCCC	CACUGGCAGU
2410	2420	2430	2440	2450	2460
GGAAGAGCA	CAAGGUCCC	CGUCGCGUAC	GCCGCCAGG	GGUAUAAAGU	GCUGGUGCUC
2470	2480	2490	2500	2510	2520
AAUCCUCCG	UGGUGCCAC	CCUGGGAUUU	GCGCGUAU	UGUCCAGGC	ACAUGGCNUC
2530	2540	2550	2560	2570	2580
AACCCACAA	UAGGACUGG	AGUCAGAAU	GUGAGACCG	GCGAGCCCU	UACUACUCC
2590	2600	2610	2620	2630	2640
ACGUAGGUA	AAUUCUUGC	CGAUGGGGC	UGCGAGGCG	GCGCCUAUA	CAUCUACAUA
2650	2660	2670	2680	2690	2700
UGGAGUAAU	GCCACUCUG	GGAUGCUAC	ACUUAUCUG	GCAUCGGAC	AGUCCUAGC
2710	2720	2730	2740	2750	2760
CAAGCAGAA	CAGCCGGGU	CAGGCUAACU	GUACUGGCCA	CGGCCACCCU	CCCGGGUGG

[Figure 3C]

2770	2780	2790	2800	2810	2820
GUGACAAACC	CCCAACCCAA	UAUAGAGGAG	GUAGCCCUCC	GACAGGAGGG	UGAGAUCACC
2830	2840	2850	2860	2870	2880
UUCUAUGGGA	GGGCGUUAAC	CCUGUCUUAU	AUCAAGGGAG	GGAGGACUUA	GAUUUUCUGC
2890	2900	2910	2920	2930	2940
CACUCAAGA	AAAGUGUGA	CGAGCUCGCA	ACGGCCCUUC	GGGGCUGGG	CUUGAACGCU
2950	2960	2970	2980	2990	3000
GUGGCAUAUU	ACAGAGGGUU	GGACGUCUCC	AUAUAACCAA	CUCAAGGAGA	UGUGGUGGUC
3010	3020	3030	3040	3050	3060
GUUGCCACCG	ACGCCCUCAU	GACGGGGUAU	ACUGGAGACU	UUAGACUCCG	GAUCGACUGC
3070	3080	3090	3100	3110	3120
AAAGUAGCGG	UACCCAGGCG	CGUAGACUUC	AGCCCGGAGC	GCACCUUCAC	UAUAACACAA
3130	3140	3150	3160	3170	3180
CAGACUGUCC	CGCAAGACGC	UGUCUCAGGU	AGUCAGCGCC	GAGGGCGCAC	CGSUMGAGGA
3190	3200	3210	3220	3230	3240
AGACUGGGCA	UUUAUAGGUA	UGUUUCCACU	GGUGAGCGAG	CCACAGGAUU	GUUUGACAGU
3250	3260	3270	3280	3290	3300
GUAGUACUCU	GUGAGUGCUA	CAACGACAGG	CCUGCUUGGU	AUGAGCGUCC	ACCAGUGGAG
3310	3320	3330	3340	3350	3360
ACGACCGUCA	GGCTCAGGGC	GUUUUUAUAC	ACGCCUGGCU	UGCCCGUGUG	CAAGGACAC
3370	3380	3390	3400	3410	3420
CUUGAGUUUU	GGAGGCGAGU	UUUCACCGGC	CUACACACAA	UAGACGCUCA	UUUCUUUUCC
3430	3440	3450	3460	3470	3480
CAAGACAAAC	AGUCGCGGGA	AAAUUUUGCA	UACUUAGUAG	CCUAUCAGGC	CACAGUGUGC
3490	3500	3510	3520	3530	3540
CCCAGGGCCA	AAGCGCCCCC	CCCGUCCUGG	GACGUAUGUU	GGAAAGUCCU	GACUCGACUC
3550	3560	3570	3580	3590	3600
AAACCCACGC	UUGUGGGCCC	UACACGUCUC	CUGUACCGUU	UGGGCUCUGU	UACCAACGAG
3610	3620	3630	3640	3650	3660
GUACCCCUUA	CACACCCCGU	GACAAAUUAC	AUGCCCAUUA	GCAUGCAAGC	UGACCCUGAG
3670	3680	3690	3700	3710	3720
GUCAUGACCA	GCAAGUGGGU	CCUGGCTUGG	GGAGUCUUAG	CAGCCGUGGC	CGCGUAUUGC
3730	3740	3750	3760	3770	3780
UUAGGACACG	GGUGGUUUUC	CAUUAUUGGC	CGUUUACACA	UCAACCAAGC	AGCUGUCGUU
3790	3800	3810	3820	3830	3840
GCUCGAGACA	AGGAGGUCCU	CUAUGAGGCU	UUUGAUGAGA	UGGAGGAUUG	UGCCUCCAGA
3850	3860	3870	3880	3890	3900
GGCGUCUCC	UUGAAGAGGG	GCAGCGGAUA	GCCGAGUUGC	UGAAGUCCAA	GAUCCAGGCG
3910	3920	3930	3940	3950	3960
UUUUUGACGC	AAACCCUUAU	ACAGGCGCAG	GACAUAUAC	CCGCUUGUCA	AGCUCUGUGG
3970	3980	3990	4000	4010	4020
CCCAGUAGG	AGCAAUUCCG	GGCCAAACAU	AUGUGGAAAU	UCAUAAGCGG	CAUUCAGUAC
4030	4040	4050	4060	4070	4080
CCCGCAGGAC	UGUACACAU	GCCAGGGAGC	CCUGCUGUGG	CUUCCAUAGU	GCACUACAC
4090	4100	4110	4120	4130	4140
CCCGCCCUCA	CCAGUCCGUU	GUCAACUAGC	ACCAUCCUCC	UUUUUAACAU	UCUGGGGGGC

[Figure 3D]

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4150      4160      4170      4180      4190      4200
UGGCUGGCGU CCCAAAUUGC GCCACCCXCG GGGGCCACUG GCUTUUGUUGU CAGUGGCCUG

4210      4220      4230      4240      4250      4260
GUUGGAGGUG CUGUUGGCAG CAUAGGCUUG GUUAAAGUGC UGGUGGACAU CCUGGCAGGG

4270      4280      4290      4300      4310      4320
UAUGGUGCGG GCAUUUCCGG GGGCCUCGUC GCGUUUAGA UCAUGUCUGG CGAGAAAGCC

4330      4340      4350      4360      4370      4380
UCCAUUGGAG AUUUAUCAA CUGGCGCCU GGAUUUCUGU CUCCAGGUGC UCUGGUGGUG

4390      4400      4410      4420      4430      4440
GGAGUUAUCU GGGGCGCCAU UCUGGCGCGC CAUGUGGGAC GGGGGGAGG CGCGGUCCAA

4450      4460      4470      4480      4490      4500
UGGUAUGACA GCGUUUCCG CUGGCGUUC AGAGGAAACC ACUGGCGGCC UACUACAUAU

4510      4520      4530      4540      4550      4560
GUGAGGAGAU CGGAGGCGUC GCAGCGUGUC ACCCAACUGC UUGGCUUCU CACUAUUAU

4570      4580      4590      4600      4610      4620
AGUCUAUUA GGAUUAUUA CAACUGGAG ACUGAGGAGU GCGCCAUCC AUGGCGCGGC

4630      4640      4650      4660      4670      4680
UCUGGCGUCC GCGAUGUGU GGAUGGGUC UGUAGGUCU UAACAGACU UAAGAACUGG

4690      4700      4710      4720      4730      4740
CUGACCUCCA AGGUGUUCG AAAGAGGCGU GCGCCCGCU UUAUCUCUG CCAAAGGGG

4750      4760      4770      4780      4790      4800
UACAGGGCG UGUGGGCGG CACUGGCAU AUGACACAC GAUGCCCGU CGGCGGCAAC

4810      4820      4830      4840      4850      4860
AUCUCUGGA ACCUCCGCU GGGUCUAGG AGAUAACAG GAUCCAAAC CUGCAUGAAC

4870      4880      4890      4900      4910      4920
ACCUCCGAG GAGCUUUC UAUCAUUGU UAUAAGAGG GCGAGGCUU GCGAAAGCC

4930      4940      4950      4960      4970      4980
GCGUUAACU UCAAGACCG CAUCUGGAG GUGGCGGCU CAGAGGACG GGAAGUGAGG

4990      5000      5010      5020      5030      5040
CAGCAGGAG CAUAGGCGA UAUAAAGGG CUGACACUG ACAACUAA AGUCCCUUGC

5050      5060      5070      5080      5090      5100
CAUCUCCCU CUCAGAGGU UUCUCUUGG UUGGAGGAG UACAAUCCA UAGGUCCGC

5110      5120      5130      5140      5150      5160
CCCAACCAA AGCGUUUUU CCGGAGAGG GUCUGUUA GCGUUGGCU CAUUAUAU

5170      5180      5190      5200      5210      5220
GUGGUUGGG CUCAGCUUC CUGAGACCU GAGCCGACU CUGAGGUAG GAUUGGCUU

5230      5240      5250      5260      5270      5280
CUAACGAGC CAUCCCAUA CAGGCGGAG GCUCAAGCG GCGUUUAGC GCGGGGUA

5290      5300      5310      5320      5330      5340
CCCCAUCG AGCAAGCUC CUCAGCGAG CAGCUGUGG CGCCAUGCU GCGAGGACG

5350      5360      5370      5380      5390      5400
USCACCACC ACGUAAGAC CUAUGAUGG GACAUGGUG AUGCCAACU GUUCAUGGG

5410      5420      5430      5440      5450      5460
GGGCGGUGA UUGGGAUAG GUCUGAGUC AAAGUGGUG UUCUGGACU CAAGACUA

5470      5480      5490      5500      5510      5520
AUGACGAGG AGAGGGCGA CUGAGGCUU UCAGUACAU GGAUUAU GCUCCGAGG

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[Figure 3E]

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5530      5540      5550      5560      5570      5580
AAGAGGURUC CACCGGCCUU ACCGGCUUGG GCGCGGCCUG AUUACAAACC ACCGCUUUGU

5590      5600      5610      5620      5630      5640
GUAUCUGUGA AGAGGCCGGA UUAACCAACA CCCACUGUUG CGGCGUGUGC UCUCGCCGCC

5650      5660      5670      5680      5690      5700
CCCAAAAGA CCCCGACGCC UCCUCCAAAG AGACGCCGSA CAGUGGGUCU GAGCGAGAGC

5710      5720      5730      5740      5750      5760
ACCAUAGGAG AUGCCGCCCA ACAGCUGGCC AUCAGAUCCU UUGGCCAGCC CCCCCCAGC

5770      5780      5790      5800      5810      5820
GGCGAUUCAG GCUUUUCCBC GCGGGCGGAC GCGCGCGACU CCGCGAUUG GACAACCCCU

5830      5840      5850      5860      5870      5880
GACGAGUUGG CUUUUUUGGA GACAGGUUCU ACCUCUCCA UGCCCCCUU CGAGGGGAGG

5890      5900      5910      5920      5930      5940
CCCGGGGACC CAGACCUUGA GCUAGAGCAG GUAGAGCUUC AACUCCUCC CAGGGGGGG

5950      5960      5970      5980      5990      6000
GAGGCAGCUC CCGGCUUGGA CUUGGGGUC UGGUCUACU GCUCCGAGGA GGAUGACUCC

6010      6020      6030      6040      6050      6060
GUCCUGUGCU GCUCCAGUC UAUAUCCCG ACCCGGCCUC UAUAACUCC UUGUAGCCCC

6070      6080      6090      6100      6110      6120
GAGAGGAAU AGUUGCCAU UAACUCCUUG AGCAACUCCG UGUUGCGAU CCAUACAGG

6130      6140      6150      6160      6170      6180
GUUAUCUGUA CUACUCAAU GAGUGCCUCA CUAGGGCUA AAAAGGUAC UUUUUAUAG

6190      6200      6210      6220      6230      6240
AUGCAGGUC UGAGGCCUA UUAUUAUCA GUUUAAAGG ACAUACAGU AGCGGCCUCC

6250      6260      6270      6280      6290      6300
AAGGUCAGCG CAGGCUCCU CACCUUAGAG GAGCGUGCC AAUUGACCC ACCCAGCUCU

6310      6320      6330      6340      6350      6360
GCAAGAUCCA AGUAUGGUGU UCGGCUAAG GAGGUCCGA GCUUUCUCCG GAGGGCCGUC

6370      6380      6390      6400      6410      6420
AACCCAUCA AGUCGUGUG GAGGAGCCU UUGGAGACU CACAACACC AAUUCUACA

6430      6440      6450      6460      6470      6480
ACCAUCUUG CCAAAAUGA GUUGUUCUG GUGGACCCG CCAGGGGGG UAAAAACCA

6490      6500      6510      6520      6530      6540
GCUCGCUUA UCUUUUAGC UGACCUCCG GUCAGGUCU GCGAGAGAU GGCCTUUAU

6550      6560      6570      6580      6590      6600
GAUGUCACAC AAAAGCUCC UAGGGCGUG AUGGGGCUU CUUAUGCUU CCAGUACUCC

6610      6620      6630      6640      6650      6660
CCCGUCAGC GCGUGAGUU UCUCUUGAG GCUAGGCGG AAAAGAGGA CCCTAUGGU

6670      6680      6690      6700      6710      6720
UUUUCGUAU AUACCCGUG CUUAGACUA ACCGUCACU AGAGAGACU CAGGACUGG

6730      6740      6750      6760      6770      6780
GAGUCCAUU ACCAGGCCG CUCCUACCC GAGGAGGCC GAACUGCCAU ACACUGCCG

6790      6800      6810      6820      6830      6840
ACUGAGAGC UCUAUGUGG AGGUCGAGG UCAACAGCA AGGCGCAGU CUGCGGGUC

6850      6860      6870      6880      6890      6900
AGGCUUAGC GCGCAGCGG GUUGCUUAC ACUAGUAUG GUAAACCAU CACAUCCAU

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[Figure 3F]

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6910      6920      6930      6940      6950      6960
GGAAGAAGCCC UAGCGCGCUGG CAAAGGCUUGG GGGAAUAAUUG GGGCCACGAAU GCUUGUAUUGC

6970      6980      6990      7000      7010      7020
GGCGACGACU UGUUCGUUCU CUCAGAAAGC CAGGGGACUG AGGAGGACGA GCGGAACCUG

7030      7040      7050      7060      7070      7080
AGAGCCUUC ACGAGGCUUU GACCAGGUUU UCUGCCCCUC CUGGUGACCC CCCGAGACCG

7090      7100      7110      7120      7130      7140
GAUAUAGACC UGGAGCGAAU AACAGCUUGU UCCUCAAAAG UGUUCUGUGC ACUUGGCCCA

7150      7160      7170      7180      7190      7200
CAGGGCCGCG GCGAGACUA CCUGACGAGA GAGCCACCA CUUCAAUUGC CCGGCGUGCC

7210      7220      7230      7240      7250      7260
UGGGAACAG UUAGACACUC CCGUGUCAU UCAUGGCUUG GAAACAUCAU CCAGUACGCU

7270      7280      7290      7300      7310      7320
CCAGCCAUUU GGUUCGCAU GGUCCUGUG ACACACUUCU UCUCCAUUCU CAUGGCCGAG

7330      7340      7350      7360      7370      7380
GACACCCUAG ACCAGAACCU UAAAUUGAA AUGUACGGAU CCGUGUACUC CGUGAGUCCU

7390      7400      7410      7420      7430      7440
CUGGACCUCC CAGCCAUAAU UGAAAGGUUA CAGGGGCUUG AGCCCUUCC UCUGCACACA

7450      7460      7470      7480      7490      7500
UACACUCCCG ACGAACUGAC GCGGGUGGCU UCAAGCCUCA GAAACUUGG GGGCCACCC

7510      7520      7530      7540      7550      7560
CUCAGAGGU GGAAGAGUG GCGCGGUGCA GUUAGGCGGU CCCUACUUC CCGUGGGGG

7570      7580      7590      7600      7610      7620
AGGGCGGCG UUGCGGUGG GUACCUUUC AACUGGGCG UGAAGACCA GCUCAAACUC

7630      7640      7650      7660      7670      7680
ACUCUUUUG CCGAGGCACG CUCUCCGGAU UAGUCCAGU GGUUUAACCG CCGCGCCGC

7690      7700      7710      7720      7730      7740
GGGGGAGCA UUAUACAGG CGUGUGGCU GCGGACCCC GCGUAUACU CCUAGCCUA

7750      7760      7770      7780      7790      7800
CUCCUACUU CUGUAGGGU AGGCCUUCU CUAUCCCGG CUUGAUAGAG CGGCACAGU

7810      7820      7830      7840      7850      7860
UAGCUACAU CCAUAGCUAA CUGUCCUUU UUUUUUUUU UUUUUUUUU UUUUUUUUU

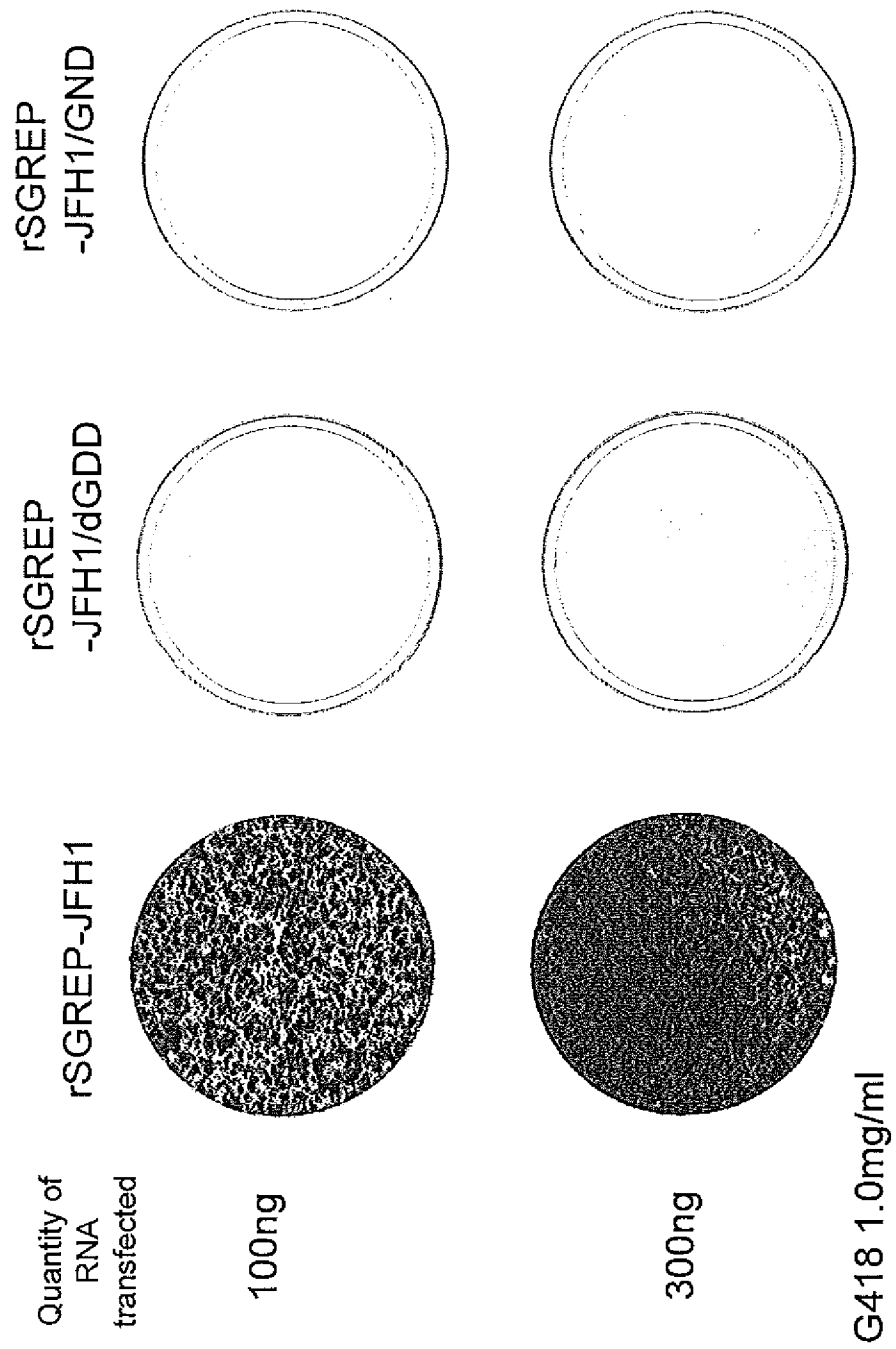
7870      7880      7890      7900      7910      7920
UUUUUUUUU CUUUUUUUU UUUUUUUU UUUUUUUU UCUCAUUGA UUCUACUUC

7930      7940      7950      7960      7970      7980
UUUUUUUGG GCUCAUCU AGCCUAGUC AGGCUAGCU GUGAAAGGC CGUGAGCCGC

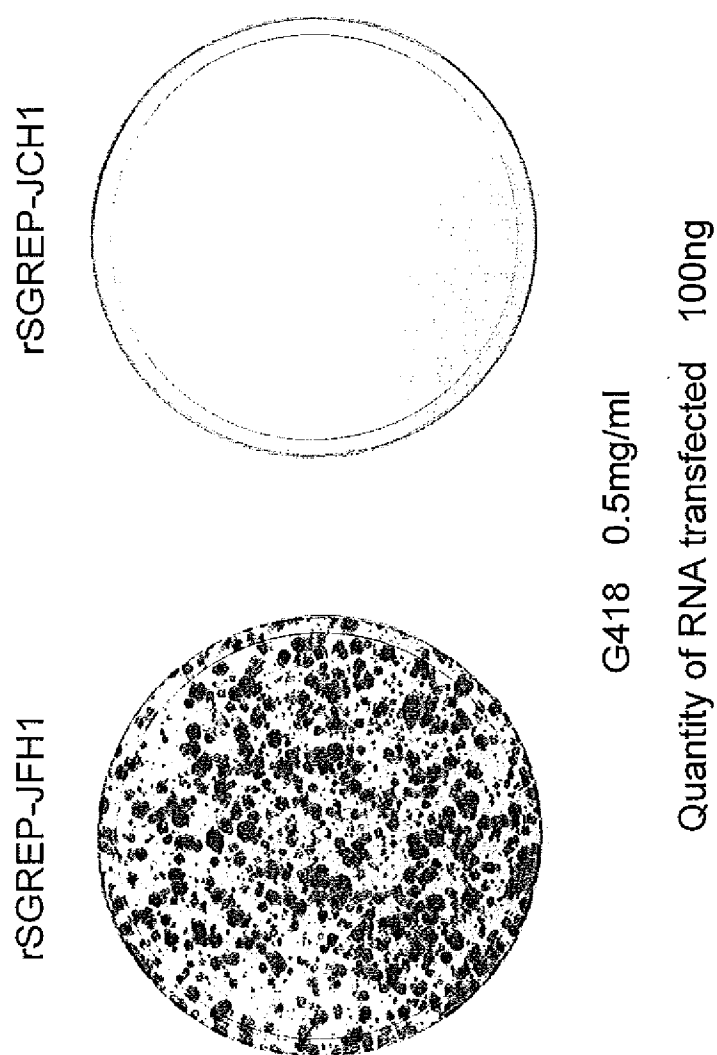
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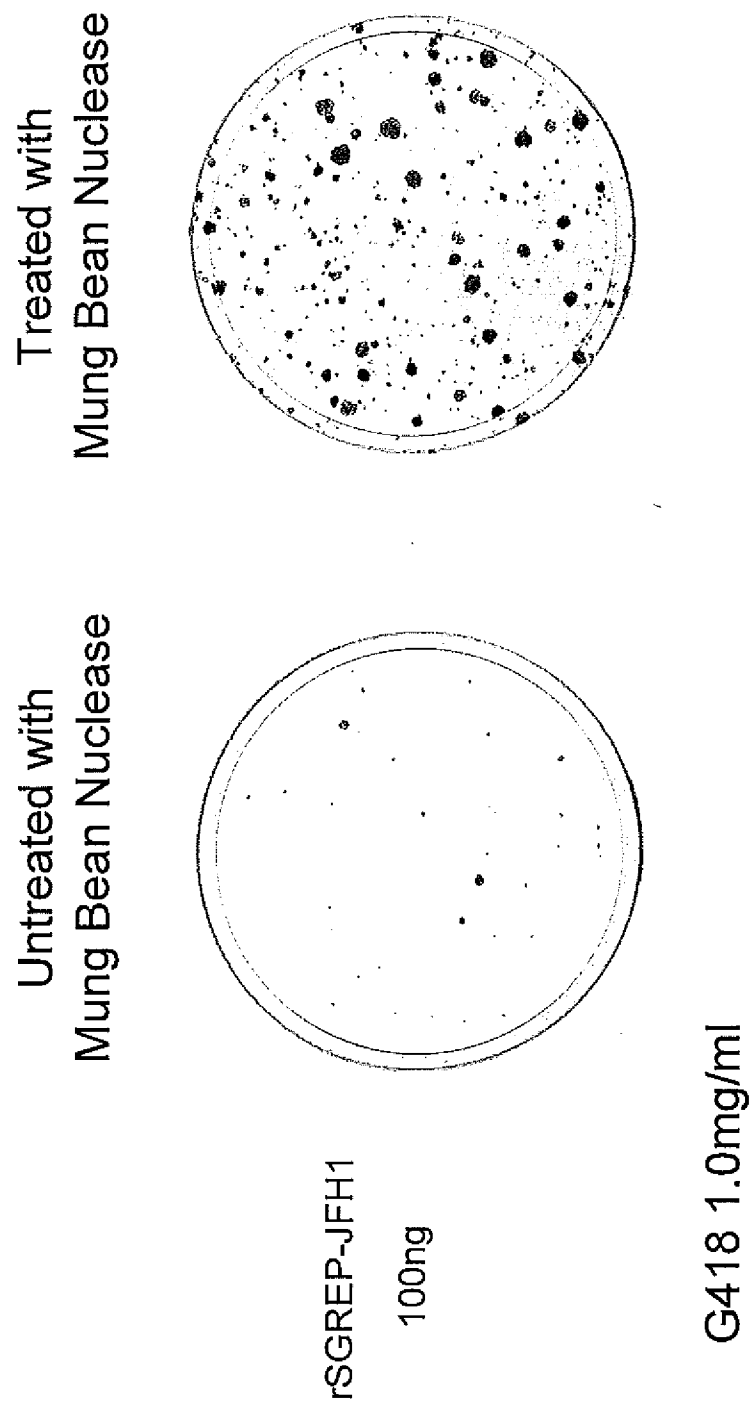
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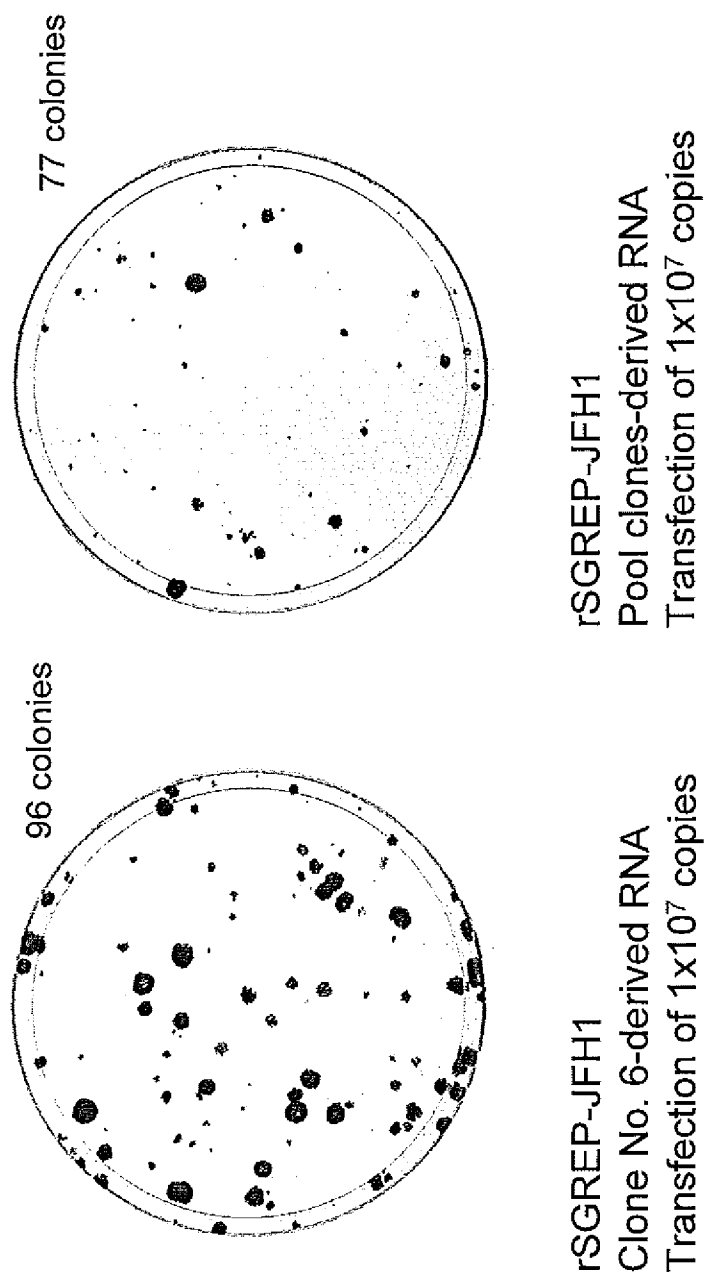
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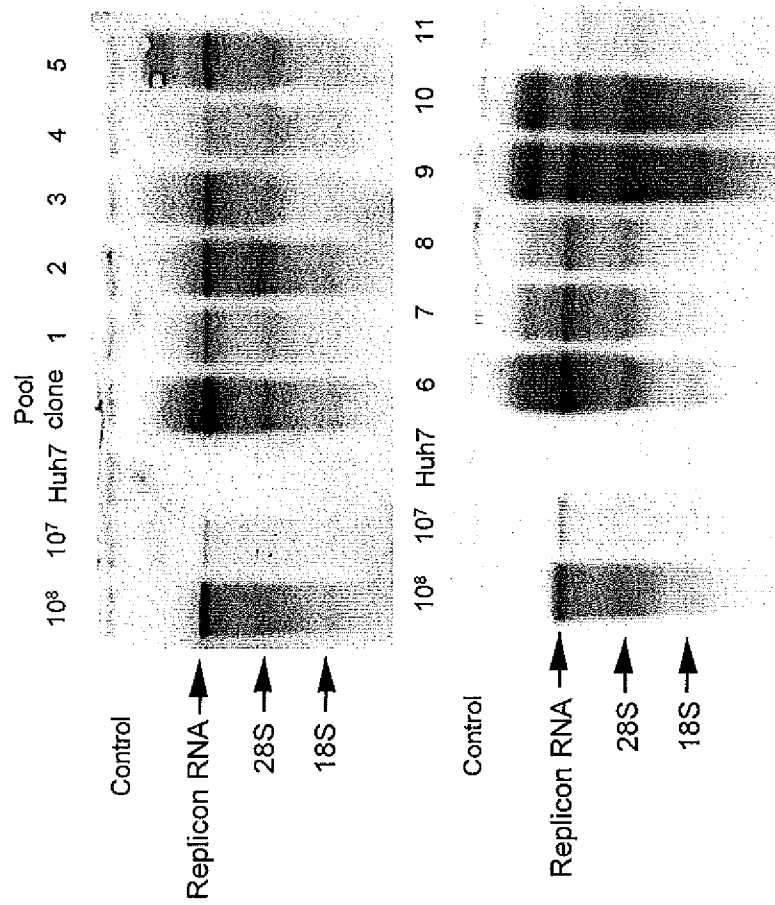
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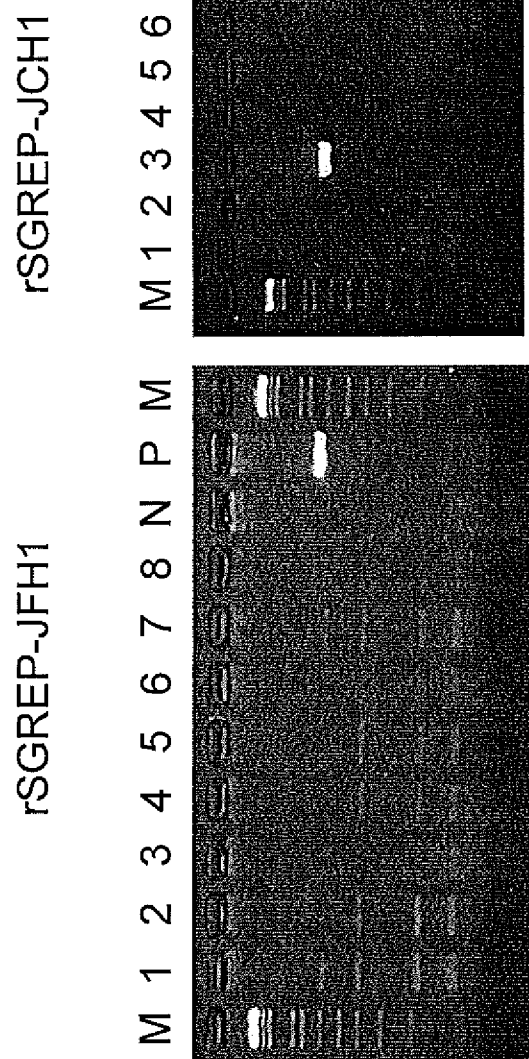
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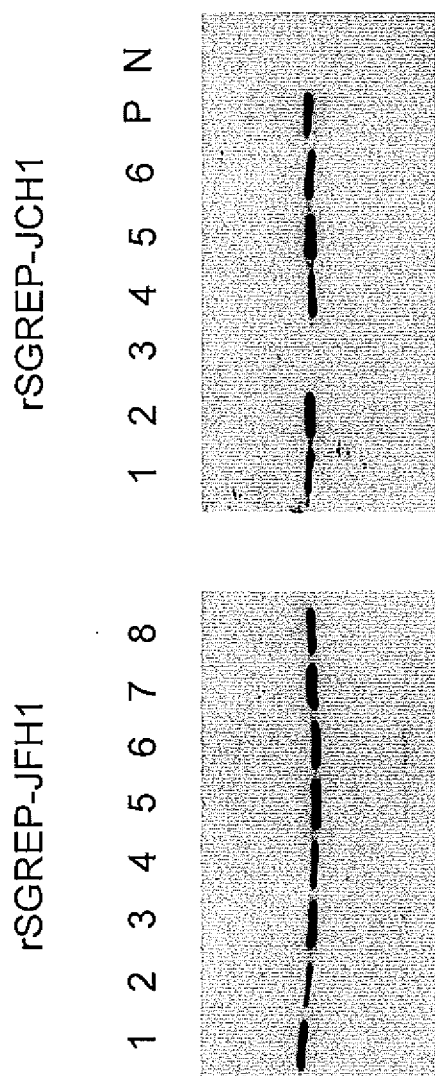
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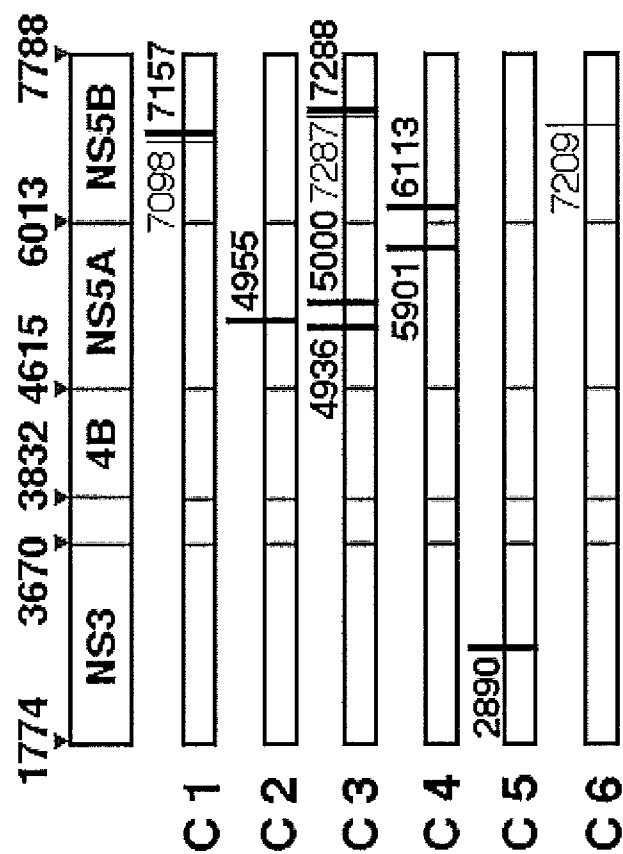
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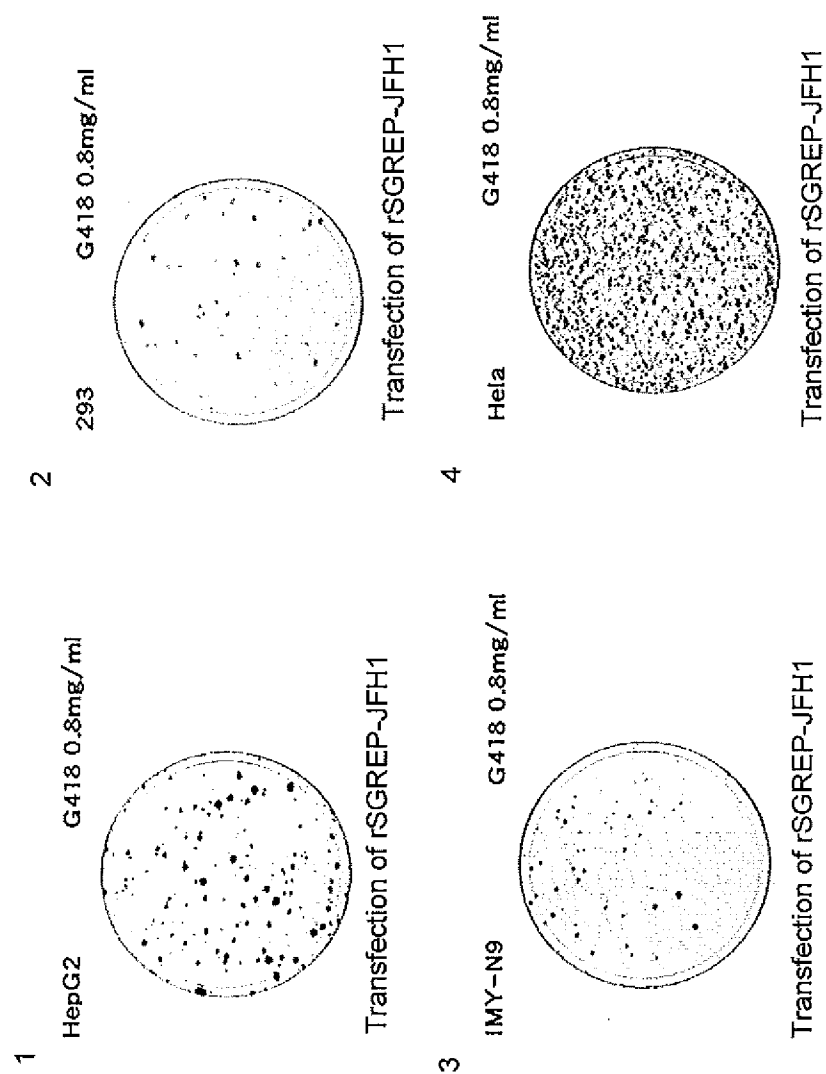
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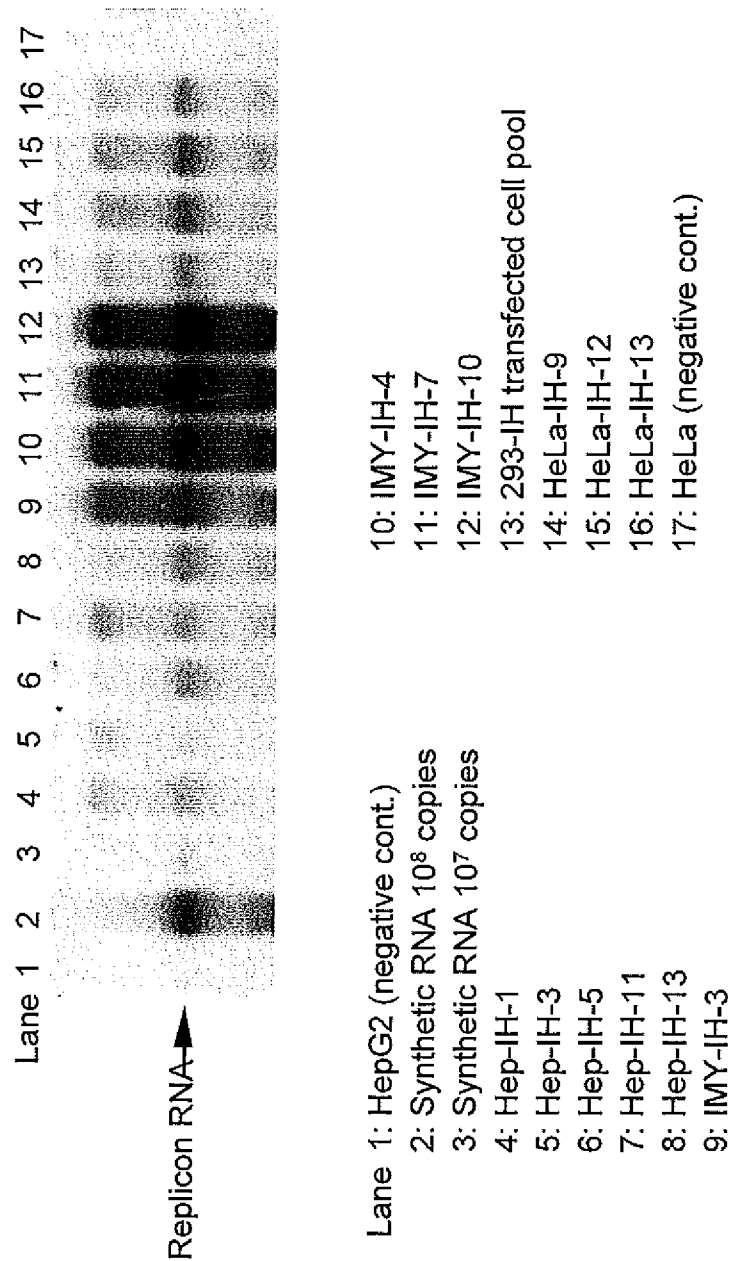
[Figure 11]



[Figure 12]

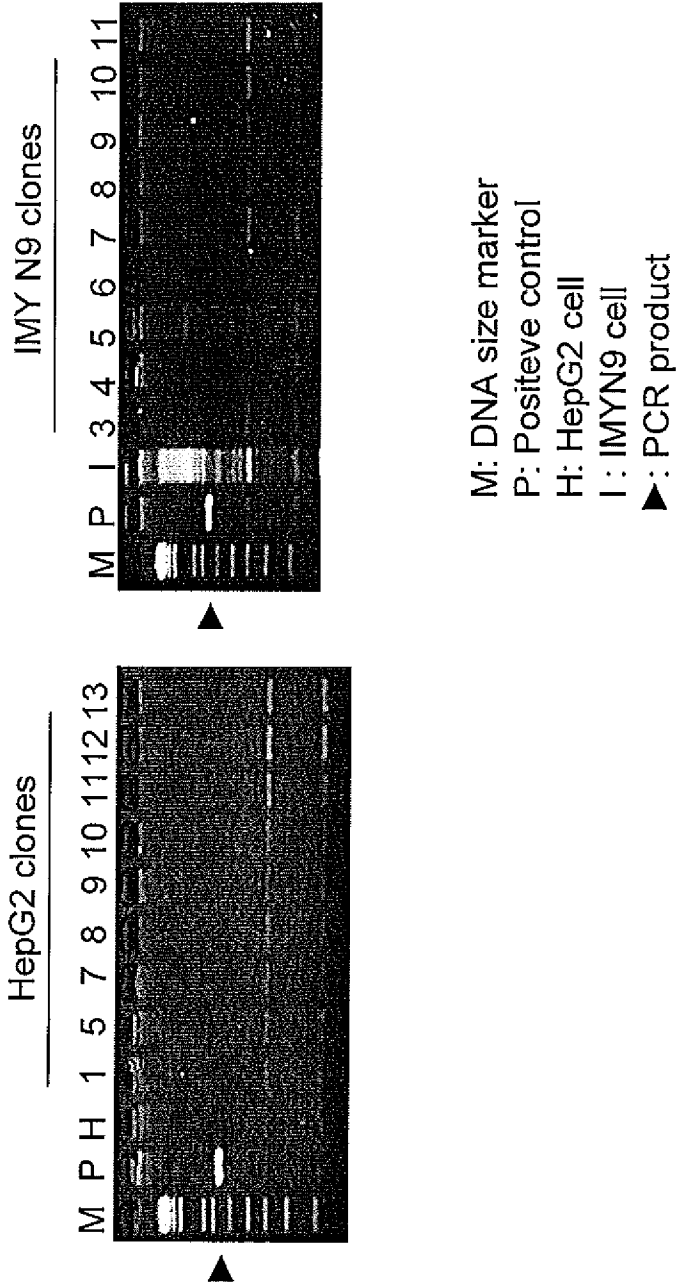


[Figure 13]



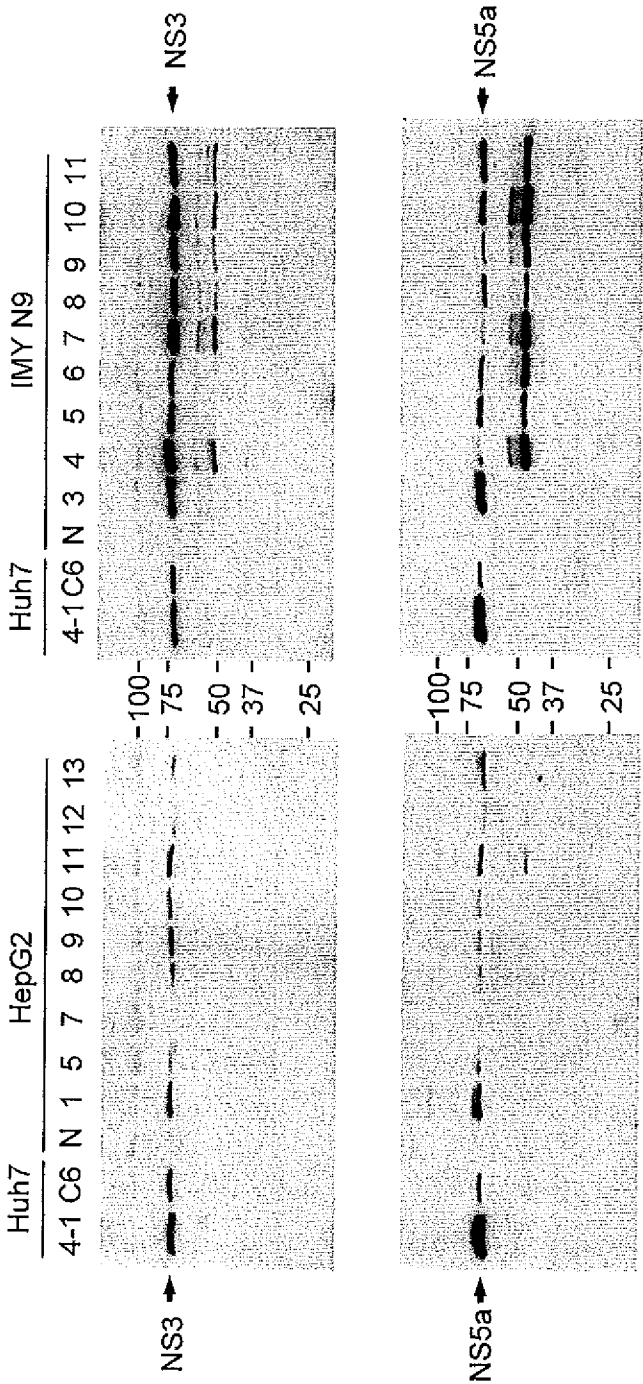
[Figure 14]

Detection of neomycine resistant gene integrations
by genomic DNA PCR analysis
In HepG2 and IMYN9 replicon cells



[Figure 15]

Western blot analysis of NS3 and NS5a protein



[Title of Document] ABSTRACT

[Abstract]

[Technical Problem] An object is to provide a replicon RNA that is derived from HCV of a different genotype from genotype 1b.

[Technical Solution] A replicon RNA comprising a nucleotide sequence at least containing the 5' untranslated region, the nucleotide sequence encoding NS3 protein, NS4A protein, NS4B protein, NS5A protein and NS5B protein, and the 3' untranslated region on the genomic RNA of hepatitis C virus of genotype 2a is provided.

[Selected Drawing] None